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Description A collection of functions for conducting meta-analysis using a structural equation modeling (SEM) approach via the 'OpenMx' and 'lavaan' packages. It also implements various procedures to perform meta-analytic structural equation modeling on the correlation and covariance matrices, see Cheung (2015) <[doi:10.3389/fpsyg.2014.01521](https://doi.org/10.3389/fpsyg.2014.01521)>.

License GPL (>=2)

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Aloe14	<i>Multivariate effect sizes between classroom management self-efficacy (CMSE) and other variables reported by Aloe et al. (2014)</i>
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Description

This study reports sixteen studies on the effect sizes (correlation coefficients) between CMSE and emotional exhaustion (EE), depersonalization (DP), and (lowered) personal accomplishment (PA) reported by Aloe et al. (2014).

Format

A data frame with 16 observations on the following 14 variables.

Study a factor with levels Betoret Brouwers & Tomic Bumen Chang Durr Evers et al. Friedman Gold Huk Kress Kumarakulasingam Martin et al. Ozdemir Skaalvik and Skaalvik Williams

Year Year of publication

EE Emotional exhaustion

DP Depersonalization

PA (Lowered) personal accomplishment

V_EE Sampling variance of emotional exhaustion

V_DP Sampling variance of depersonalization

V_PA Sampling variance of (lowered) personal accomplishment

C_EE_DP Sampling covariance between EE and DP

C_EE_PA Sampling covariance between EE and PA

C_DP_PA Sampling covariance between DP and PA

Publication_type Either Dissertation or Journal

Percentage_females Percentage of females in the study

Years_experience Average years of experience

Source

Aloe, A. M., Amo, L. C., & Shanahan, M. E. (2014). Classroom management self-efficacy and burnout: A multivariate meta-analysis. *Educational Psychology Review*, **26(1)**, 101-126. doi:10.1007/s10648-013-9244-0

Examples

```
data(Aloe14)

## Random-effects meta-analysis
meta1 <- meta(cbind(EF,DP,PA),
              cbind(V_EE, C_EE_DP, C_EE_PA, V_DP, C_DP_PA, V_PA),
              data=Aloe14)
## Remove error code
meta1 <- rerun(meta1)

summary(meta1)

## Extract the coefficients for the variance component of the random effects
coef1 <- coef(meta1, select="random")

## Convert it into a symmetric matrix by row major
my.cov <- vec2symMat(coef1, byrow=TRUE)

## Convert it into a correlation matrix
cov2cor(my.cov)

## Plot the multivariate effect sizes
plot(meta1)
```

anova

Compare Nested Models with Likelihood Ratio Statistic

Description

It compares nested models with the likelihood ratio statistic from various objects. It is a wrapper of [mxCompare](#).

Arguments

object	An object or a list of objects of various classes. It will be passed to the base argument in mxCompare .
...	An object or a list of objects of various classes. It will be passed to the comparison argument in mxCompare .
all	A Boolean value on whether to compare all bases with all comparisons. It will be passed to the all argument in mxCompare .

Value

A table of comparisons between the models in base and comparison.

Note

When the objects are class `wls`, the degrees of freedom in the base and comparison models are incorrect, while the degrees of freedom of the difference between them is correct. If users want to obtain the correct degrees of freedom in the base and comparison models, they may individually apply the `summary` function on the base and comparison models.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
## Test the significance of a predictor with likelihood ratio test
## Model0: No predictor
model0 <- meta(y=yi, v=vi, data=Hox02, model.name="No predictor")

## Model1: With a predictor
model1 <- meta(y=yi, v=vi, x=weeks, data=Hox02, model.name="One predictor")

## Compare these two models
anova(model1, model0)
```

as.mxAlgebra

Convert a Character Matrix into MxAlgebra-class

Description

It converts a character matrix into MxAlgebra object.

Usage

```
as.mxAlgebra(x, startvalues = NULL, lbound = NULL, ubound = NULL, name = "X")
```

Arguments

<code>x</code>	A character or numeric matrix, which consists of valid operators in mxAlgebra.
<code>startvalues</code>	A list of starting values of the free parameters. If it is not provided, all free parameters are assumed 0.
<code>lbound</code>	A list of lower bound of the free parameters. If it is not provided, all free parameters are assumed NA.
<code>ubound</code>	A list of upper bound of the free parameters. If it is not provided, all free parameters are assumed NA.
<code>name</code>	A character string of the names of the objects based on.

Details

Suppose the name argument is "X", the output is a list of the following elements.

Value

mxalgebra	An mxAlgebra object.
parameters	A column vector mxMatrix of the free parameters.
list	A list of mxMatrix to form the mxAlgebra object.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[as.mxMatrix](#), [mxAlgebra](#)

Examples

```
## a, b, and c are free parameters
(A1 <- matrix(c(1, "a*b", "a^b", "exp(c)"), ncol=2, nrow=2))
##      [,1] [,2]
## [1,] "1"  "a^b"
## [2,] "a*b" "exp(c)"

A <- as.mxAlgebra(A1, startvalues=list(a=1, b=2),
                  lbound=list(a=0), ubound=list(b=1, c=2),
                  name="A")

## An object of mxAlgebra
A$mxalgebra
## mxAlgebra 'A'
## $formula: rbind(cbind(A1_1, A1_2), cbind(A2_1, A2_2))
## $result: (not yet computed) <0 x 0 matrix>
## dimnames: NULL

## A matrix of parameters
A$parameters
## FullMatrix 'Avars'

## $labels
##      [,1]
## [1,] "a"
## [2,] "b"
## [3,] "c"

## $values
##      [,1]
## [1,] 1
## [2,] 2
## [3,] 0

## $free
##      [,1]
## [1,] TRUE
```

```
## [2,] TRUE
## [3,] TRUE

## $lbound
##      [,1]
## [1,]    0
## [2,]   NA
## [3,]   NA

## $ubound
##      [,1]
## [1,]   NA
## [2,]    1
## [3,]    2

## A list of matrices of elements for the mxAlgebra
A$list
## $A1_1
## mxAlgebra 'A1_1'
## $formula: 1
## $result: (not yet computed) <0 x 0 matrix>
## dimnames: NULL

## $A2_1
## mxAlgebra 'A2_1'
## $formula: a * b
## $result: (not yet computed) <0 x 0 matrix>
## dimnames: NULL

## $A1_2
## mxAlgebra 'A1_2'
## $formula: a^b
## $result: (not yet computed) <0 x 0 matrix>
## dimnames: NULL

## $A2_2
## mxAlgebra 'A2_2'
## $formula: exp(c)
## $result: (not yet computed) <0 x 0 matrix>
## dimnames: NULL
```

as.mxMatrix

Convert a Matrix into MxMatrix-class

Description

It converts a matrix into MxMatrix-class via mxMatrix.

Usage

```
as.mxMatrix(x, name, ...)
```

Arguments

x	A character or numeric matrix. If x is not a matrix, as.matrix(x) is applied first.
name	An optional character string as the name of the MxMatrix object created by mxModel function. If the name is missing, the name of x will be used.
...	Further arguments to be passed to mxMatrix . It should be noted that type, nrow, ncol, values, free, name and labels will be created automatically. Thus, these arguments except labels should be avoided in ...

Details

If there are non-numeric values in x, they are treated as the labels of the parameters. If a "*" is present, the numeric value on the left-hand side will be treated as the starting value for a free parameter. If an "@" is present, the numeric value on the left-hand side will be considered as the value for a fixed parameter. If it is a matrix of numeric values, there are no free parameters in the output matrix.

Value

A [MxMatrix-class](#) object with the same dimensions as x

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[mxMatrix](#), [create.mxMatrix](#), [create.Fmatrix](#), [checkRAM](#), [lavaan2RAM](#), [as.symMatrix](#)

Examples

```
## a and b are free parameters with starting values and labels
(a1 <- matrix(c(1:4, "5*a", 6, "7*b", 8, 9), ncol=3, nrow=3))
#      [,1] [,2] [,3]
# [1,] "1"  "4"  "7*b"
# [2,] "2"  "5*a" "8"
# [3,] "3"  "6"  "9"

a1 <- as.mxMatrix(a1)

## a and b are fixed parameters without any labels, name="new2"
(a2 <- matrix(1:9, ncol=3, nrow=3))
#      [,1] [,2] [,3]
# [1,] 1    4    7
# [2,] 2    5    8
# [3,] 3    6    9

new2 <- as.mxMatrix(a2, name="new2")

## Free parameters without starting values
```

```

(a3 <- matrix(c(1:4, "*a", 6, "*b", 8, 9), ncol=3, nrow=3))
#      [,1] [,2] [,3]
# [1,] "1"  "4"  "*b"
# [2,] "2"  "*a" "8"
# [3,] "3"  "6"  "9"

a3 <- as.mxMatrix(a3, lbound=0)

## A free parameter without label
(a4 <- matrix(c(1:4, "5*", 6, "7*b", 8, 9), ncol=3, nrow=3))
#      [,1] [,2] [,3]
# [1,] "1"  "4"  "7*b"
# [2,] "2"  "5*" "8"
# [3,] "3"  "6"  "9"

a4 <- as.mxMatrix(a4)

## Convert a scalar into mxMatrix object
## "name" is required as "3*a" is not a valid name.
(a5 <- as.mxMatrix("3*a", name="a5"))

## Free and fixed parameters
(a6 <- matrix(c(1, "2*a", "3@b", 4), ncol=2, nrow=2))

as.mxMatrix(a6)

```

as.symMatrix

Convert a Character Matrix with Starting Values to a Character Matrix without Starting Values

Description

It converts a character matrix with starting values to a character matrix without the starting values.

Usage

```
as.symMatrix(x)
```

Arguments

x A character or numeric matrix or a list of character or numeric matrices.

Details

If there are non-numeric values in **x**, they are treated as the labels of the free parameters. If a "*" is present, the numeric value on the left-hand side will be treated as the starting value for a free parameter or a fixed value for a fixed parameter. If it is a matrix of numeric values, there are no free parameters in the output matrix. This function removes the starting values and "*" in the matrices.

Value

A character matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[as.mxMatrix](#)

Examples

```
## a and b are free parameters with starting values and labels
(a1 <- matrix(c(1:4, "5*a", 6, "7*b", 8, 9), ncol=3, nrow=3))
#      [,1] [,2] [,3]
# [1,] "1"  "4"  "7*b"
# [2,] "2"  "5*a" "8"
# [3,] "3"  "6"  "9"

(as.symMatrix(a1))
#      [,1] [,2] [,3]
# [1,] "1"  "4"  "b"
# [2,] "2"  "a"  "8"
# [3,] "3"  "6"  "9"
```

asyCovOld

Compute Asymptotic Covariance Matrix of a Correlation/Covariance Matrix

Description

It computes the asymptotic sampling covariance matrix of a correlation/covariance matrix under the assumption of multivariate normality.

Usage

```
asyCovOld(
  x,
  n,
  cor.analysis = TRUE,
  dropNA = FALSE,
  as.matrix = TRUE,
  acov = c("individual", "unweighted", "weighted"),
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
```

```

    ...
  )

  asyCov(
    x,
    n,
    cor.analysis = TRUE,
    as.matrix = TRUE,
    acov = c("weighted", "individual", "unweighted"),
    ...
  )

```

Arguments

<code>x</code>	A correlation/covariance matrix or a list of correlation/covariance matrices. NA on the variables or other values defined in <code>na.strings</code> will be removed before the analysis. Note that it only checks the diagonal elements of the matrices. If there are missing values, make sure that the diagonals are coded with NA or values defined in <code>na.strings</code> .
<code>n</code>	Sample size or a vector of sample sizes
<code>cor.analysis</code>	Logical. The output is either a correlation or covariance matrix.
<code>dropNA</code>	Logical. If it is TRUE, the resultant dimensions will be reduced by dropping the missing variables. If it is FALSE, the resultant dimensions are the same as the input by keeping the missing variables.
<code>as.matrix</code>	Logical. If it is TRUE and <code>x</code> is a list of correlation/covariance matrices with the same dimensions, the asymptotic covariance matrices will be column vectorized and stacked together. If it is FALSE, the output will be a list of asymptotic covariance matrices. Note that if it is TRUE, <code>dropNA</code> will be FALSE automatically. This option is useful when passing the asymptotic covariance matrices to meta
<code>acov</code>	If it is <code>individual</code> (the default), the sampling variance-covariance matrices are calculated based on the individual correlation/covariance matrix. If it is either <code>unweighted</code> or <code>weighted</code> , the average correlation/covariance matrix is calculated based on the unweighted or weighted mean with the sample sizes. The average correlation/covariance matrix is used to calculate the sampling variance-covariance matrices.
<code>suppressWarnings</code>	Logical. If TRUE, warnings are suppressed. It is passed to mxRun .
<code>silent</code>	Logical. An argument to be passed to mxRun
<code>run</code>	Logical. If FALSE, only return the mx model without running the analysis.
<code>...</code>	It is ignored in <code>asyCov</code> . The additional arguments will be passed to mxRun in <code>asyCovOld</code> .

Value

An asymptotic covariance matrix of the vectorized correlation/covariance matrix or a list of these matrices. If `as.matrix=TRUE` and `x` is a list of matrices, the output is a stacked matrix.

Note

Before 1.2.6, asyCov used an SEM approach based on Cheung and Chan (2004). After 1.2.6, asyCov was rewritten based on Olkin and Siotani (1976) for correlation matrix and Yuan and Bentler (2007, p. 371) for covariance matrix. Arguments such as dropNA, suppressWarnings, silent, and run were dropped. The original version was renamed to asyCovOld for compatibility.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L., & Chan, W. (2004). Testing dependent correlation coefficients via structural equation modeling. *Organizational Research Methods*, *7*, 206-223.
- Olkin, I., & Siotani, M. (1976). Asymptotic distribution of functions of a correlation matrix. In S. Ideka (Ed.), *Essays in probability and statistics* (pp. 235-251). Shinko Tsusho.
- Yuan, K.-H., & Bentler, P. M. (2007). Robust procedures in structural equation modeling. In S.-Y. Lee (Ed.), *Handbook of Latent Variable and Related Models* (pp. 367-397). Elsevier/North-Holland.

Examples

```
C1 <- matrix(c(1,0.5,0.4,0.5,1,0.2,0.4,0.2,1), ncol=3)
asyCov(C1, n=100)

## Data with missing values
C2 <- matrix(c(1,0.4,NA,0.4,1,NA,NA,NA,NA), ncol=3)
C3 <- matrix(c(1,0.2,0.2,1), ncol=2)

## Output is a stacked matrix of asymptotic covariance matrices
asyCov(list(C1,C2), n=c(100,50), as.matrix=TRUE)

## Output is a stacked matrix of asymptotic covariance matrices
asyCov(list(C3,C3), n=c(100,50), as.matrix=TRUE)

## Output is a list of asymptotic covariance matrices using the old version
asyCovOld(list(C1,C2,C3), n=c(100,50,50), dropNA=TRUE, as.matrix=FALSE)
```

BCG

Dataset on the Effectiveness of the BCG Vaccine for Preventing Tuberculosis

Description

This dataset includes 13 studies on the effectiveness of the Bacillus Calmette-Guerin (BCG) vaccine for preventing tuberculosis (see van Houwelingen, Arends, & Stijnen (2002) for details).

Details

A list of data with the following structure:

Trial Number of the trials

Author Authors of the original studies

Year Year of publication

VD Vaccinated group with disease

VWD Vaccinated group without the disease

NVD Not vaccinated group with disease

NVWD Not vaccinated group without the disease

Latitude Geographic latitude of the place where the study was done

Allocation Method of treatment allocation

ln_OR Natural logarithm of the odds ratio: $\log((VD/VWD)/(NVD/NVWD))$

v_ln_OR Sampling variance of ln_OR: $1/VD+1/VWD+1/NVD+1/NVWD$

ln_Odd_V Natural logarithm of the odds of the vaccinated group: $\log(VD/VWD)$

ln_Odd_NV Natural logarithm of the odds of the not vaccinated group: $\log(NVD/NVWD)$

v_ln_Odd_V Sampling variance of ln_Odd_V: $1/VD+1/VWD$

cov_V_NV Sampling covariance between ln_Odd_V and ln_Odd_NV: It is always 0

v_ln_Odd_NV Sampling variance of ln_Odd_NV: $1/NVD+1/NVWD$

Source

Colditz, G. A., Brewer, T. F., Berkey, C. S., Wilson, M. E., Burdick, E., Fineberg, H. V., & Mosteller, F. (1994). Efficacy of BCG vaccine in the prevention of tuberculosis: Meta-analysis of the published literature. *Journal of the American Medical Association*, **271**, 698–702.

References

Berkey, C. S., Hoaglin, D. C., Mosteller, F., & Colditz, G. A. (1995). A random-effects regression model for meta-analysis. *Statistics in Medicine*, **14**, 395–411.

van Houwelingen, H. C., Arends, L. R., & Stijnen, T. (2002). Advanced methods in meta-analysis: Multivariate approach and meta-regression. *Statistics in Medicine*, **21**, 589–624.

Viechtbauer, W. (2010). Conducting meta-analyses in R with the metafor package. *Journal of Statistical Software*, **36**(3), 1–48. <https://www.jstatsoft.org/v36/i03/>.

Examples

```
data(BCG)

## Univariate meta-analysis on the log of the odds ratio
summary( meta(y=ln_OR, v=v_ln_OR, data=BCG,
             x=cbind(scale(Latitude,scale=FALSE),
                     scale(Year,scale=FALSE))) )
```

```
## Multivariate meta-analysis on the log of the odds
## The conditional sampling covariance is 0
bcg <- meta(y=cbind(ln_Odd_V, ln_Odd_NV), data=BCG,
            v=cbind(v_ln_Odd_V, cov_V_NV, v_ln_Odd_NV))
summary(bcg)

plot(bcg)
```

bdiagMat

Create a Block Diagonal Matrix

Description

It creates a block diagonal matrix from a list of numeric or character matrices.

Usage

```
bdiagMat(x)
```

Arguments

x A list of numeric or character matrices (or values)

Value

A numeric or character block diagonal matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

It was based on a function posted by Scott Chasalow at <http://www.math.yorku.ca/Who/Faculty/Monette/pub/stmp/0827.html>

See Also

[bdiagRep](#), [matrix2bdiag](#)

Examples

```
## Block diagonal matrix of numbers
bdiagMat( list(matrix(1:4,nrow=2,ncol=2),
               matrix(5:6,nrow=1,ncol=2)) )
#      [,1] [,2] [,3] [,4]
# [1,]  1   3   0   0
# [2,]  2   4   0   0
# [3,]  0   0   5   6
```

```
## Block diagonal matrix of characters
bdiagMat( list(matrix(letters[1:4],nrow=2,ncol=2),
               matrix(letters[5:6],nrow=1,ncol=2)) )
#      [,1] [,2] [,3] [,4]
# [1,] "a"  "c"  "0"  "0"
# [2,] "b"  "d"  "0"  "0"
# [3,] "0"  "0"  "e"  "f"
```

bdiagRep

Create a Block Diagonal Matrix by Repeating the Input

Description

It creates a block diagonal matrix by repeating the input matrix several times.

Usage

```
bdiagRep(x, times)
```

Arguments

x	A numeric or character matrix (or values)
times	Number of times of x to be repeated.

Value

A numeric or character block diagonal matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[bdiagMat](#), [matrix2bdiag](#)

Examples

```
## Block diagonal matrix of numerics
bdiagRep( matrix(1:4,nrow=2,ncol=2), 2 )
#      [,1] [,2] [,3] [,4]
# [1,]  1   3   0   0
# [2,]  2   4   0   0
# [3,]  0   0   1   3
# [4,]  0   0   2   4

## Block diagonal matrix of characters
```

```

bdiagRep( matrix(letters[1:4],nrow=2,ncol=2), 2 )
#      [,1] [,2] [,3] [,4]
# [1,] "a"  "c"  "0"  "0"
# [2,] "b"  "d"  "0"  "0"
# [3,] "0"  "0"  "a"  "c"
# [4,] "0"  "0"  "b"  "d"

```

Becker09

Ten Studies of Correlation Matrices used by Becker (2009)

Description

This dataset includes ten studies on the relationships between CSAI subscales and sports behavior. The original data were used in Craft et al. (2003), whereas a subset of them was illustrated in Becker (2009).

Details

A list of data with the following structure:

data A list of 4x4 correlation matrices. The variables are *Performance*, *Cognitive*, *Somatic*, and *Self_confidence*

n A vector of sample sizes

Type_of_sport Samples based on *Individual* or *Team*

Source

Craft, L. L., Magyar, T. M., Becker, B. J., & Feltz, D. L. (2003). The relationship between the Competitive State Anxiety Inventory-2 and sport performance: a meta-analysis. *Journal of Sport and Exercise Psychology*, **25**(1), 44-65.

References

Becker, B. J. (2009). Model-based meta-analysis. In H. Cooper, L. V. Hedges, & J. C. Valentine (Eds.), *The handbook of research synthesis and meta-analysis* (2nd ed., pp. 377-395). New York: Russell Sage Foundation.

Examples

```

data(Becker09)

#### Fixed-effects model
## First stage analysis
fixed1 <- tssem1(Becker09$data, Becker09$n, method="FEM")
summary(fixed1)

## Model specification in lavaan syntax

```

```

model <- "## Regression paths
Performance ~ Cog2Per*Cognitive + S02Per*Somatic + SC2Per*Self_confidence
Self_confidence ~ Cog2SC*Cognitive + S02SC*Somatic
## Fix the variances of Cog and S0 at 1
Cognitive ~~ 1*Cognitive
Somatic ~~ 1*Somatic
## Label the correlation between Cog and S0
Cognitive ~~ cor*Somatic
## Label the error variances of Per and SC
Performance ~~ var_Per*Performance
Self_confidence ~~ var_SC*Self_confidence"

## Display the model
plot(model, layout="spring")

RAM <- lavaan2RAM(model, obs.variables=c("Performance", "Cognitive",
                                         "Somatic", "Self_confidence"))

RAM

## ## Equivalent RAM specification using create.mxMatrix()
## A1 <- create.mxMatrix(c(0, "0.1*Cog2Per", "0.1*S02Per", "0.1*SC2Per",
##                          0, 0, 0, 0,
##                          0, 0, 0, 0,
##                          0, "0.1*Cog2SC", "0.1*S02SC", 0),
##                        type="Full", byrow=TRUE, ncol=4, nrow=4,
##                        as.mxMatrix=FALSE)
## dimnames(A1)[[1]] <- dimnames(A1)[[2]] <- c("Performance", "Cognitive",
##                                              "Somatic", "Self_confidence")
## A1
##
## S1 <- create.mxMatrix(c("0.1*var_Per",
##                          0, 1,
##                          0, "0.1*cor", 1,
##                          0, 0, 0, "0.1*var_SC"), byrow=TRUE, type="Symm",
##                        as.mxMatrix=FALSE)
## dimnames(S1)[[1]] <- dimnames(S1)[[2]] <- c("Performance", "Cognitive",
##                                              "Somatic", "Self_confidence")
## S1

## Second stage analysis
fixed2 <- tssem2(fixed1, RAM=RAM, diag.constraints=TRUE,
                 intervals.type="LB", model.name="TSSEM2 Becker09",
                 mx.algebras=list( Cog=mxAlgebra(Cog2SC*SC2Per, name="Cog"),
                                   SO=mxAlgebra(S02SC*SC2Per, name="SO"),
                                   Cog_SO=mxAlgebra(Cog2SC*SC2Per+S02SC*SC2Per,
                                                     name="Cog_SO"))) )

summary(fixed2)

## Display the model with the parameter estimates
plot(fixed2, layout="spring")

#### Fixed-effects model: with type of sport as cluster
## First stage analysis

```

```

cluster1 <- tssem1(Becker09$data, Becker09$n, method="FEM",
                  cluster=Becker09$Type_of_sport)
summary(cluster1)

## Second stage analysis
cluster2 <- tssem2(cluster1, RAM=RAM, diag.constraints=TRUE,
                  intervals.type="LB", model.name="TSSEM2 Becker09",
                  mx.algebras=list( Cog=mxAlgebra(Cog2SC*SC2Per, name="Cog"),
                                    SO=mxAlgebra(SO2SC*SC2Per, name="SO"),
                                    Cog_SO=mxAlgebra(Cog2SC*SC2Per+SO2SC*SC2Per,
                                                       name="Cog_SO")) )
summary(cluster2)

## Convert the model to semPlotModel object with 2 plots
## Use the short forms of the variable names
my.plots <- lapply(X=cluster2, FUN=meta2semPlot, manNames=c("Per", "Cog", "SO", "SC"))

## Load the library
library("semPlot")

## Setup two plots
layout(t(1:2))
## The labels are overlapped. We may modify it by using layout="spring"
semPaths(my.plots[[1]], whatLabels="est", nCharNodes=10, color="orange",
         layout="spring", edge.label.cex=0.8)
title("Individual sport")
semPaths(my.plots[[2]], whatLabels="est", nCharNodes=10, color="skyblue",
         layout="spring", edge.label.cex=0.8)
title("Team sport")

#### Random-effects model
## First stage analysis
random1 <- tssem1(Becker09$data, Becker09$n, method="REM", RE.type="Diag")
summary(random1)

## Second stage analysis
random2 <- tssem2(random1, RAM=RAM, diag.constraints=TRUE,
                  intervals.type="LB", model.name="TSSEM2 Becker09",
                  mx.algebras=list( Cog=mxAlgebra(Cog2SC*SC2Per, name="Cog"),
                                    SO=mxAlgebra(SO2SC*SC2Per, name="SO"),
                                    Cog_SO=mxAlgebra(Cog2SC*SC2Per+SO2SC*SC2Per,
                                                       name="Cog_SO")) )
summary(random2)

## Display the model
plot(random2, what="path", layout="spring")

## Display the model with the parameter estimates
plot(random2, layout="spring", color="yellow")

#### Univariate r approach
#### First stage of the analysis

```

```

uni1 <- uniR1(Becker09$data, Becker09$n)
uni1

#### Second stage of analysis using OpenMx
model2 <- "## Regression paths
          Performance ~ Cog2Per*Cognitive + S02Per*Somatic + SC2Per*Self_confidence
          Self_confidence ~ Cog2SC*Cognitive + S02SC*Somatic
          ## Provide starting values for Cog and S0
          Cognitive ~~ start(1)*Cognitive
          Somatic ~~ start(1)*Somatic
          ## Label the correlation between Cog and S0
          Cognitive ~~ cor*Somatic
          ## Label the error variances of Per and SC
          Performance ~~ var_Per*Performance
          Self_confidence ~~ var_SC*Self_confidence"

RAM2 <- lavaan2RAM(model2, obs.variables=c("Performance", "Cognitive",
                                           "Somatic", "Self_confidence"))

RAM2

uni2mx <- uniR2mx(uni1, RAM=RAM2)
summary(uni2mx)

#### Second stage of analysis Using lavaan
model3 <- "## Regression paths
          Performance ~ Cognitive + Somatic + Self_confidence
          Self_confidence ~ Cognitive + Somatic"

uni2lavaan <- uniR2lavaan(uni1, model3)
lavaan::summary(uni2lavaan)

```

Becker83

Studies on Sex Differences in Conformity Reported by Becker (1983)

Description

The data set includes studies on sex differences in conformity using the fictitious norm group paradigm reported by Becker (1983).

Details

The variables are:

study study number

di Standardized mean difference

vi Sampling variance of the effect size

percentage Percentage of male authors

items Number of items

Source

Becker, B. J. (1983, April). Influence again: A comparison of methods for meta-analysis. *Paper presented at the annual meeting of the American Educational Research Association, Montreal.*

Hedges, L. V., & Olkin, I. (1985). *Statistical methods for meta-analysis*. Orlando, FL: Academic Press.

References

Cheung, M. W.-L. (2010). Fixed-effects meta-analyses as multiple-group structural equation models. *Structural Equation Modeling*, **17**, 481-509.

Examples

```
data(Becker83)

## Random-effects meta-analysis
summary( meta(y=di, v=vi, data=Becker83) )

## Mixed-effects meta-analysis with log(items) as the predictor
summary( meta(y=di, v=vi, x=log(items), data=Becker83) )
```

Becker92

Six Studies of Correlation Matrices reported by Becker (1992; 1995)

Description

This data set includes six studies of correlation matrices reported by Becker (1992; 1995).

Details

A list of data with the following structure:

data A list of 6 studies of correlation matrices. The variables are *Math* (math aptitude), *Spatial* (spatial ability), and *Verbal* (verbal ability)

n A vector of sample sizes

Source

Becker, B. J. (1992). Using results from replicated studies to estimate linear models. *Journal of Educational Statistics*, **17**(4), 341-362. doi:10.3102/10769986017004341

Becker, B. J. (1995). Corrections to "Using Results from Replicated Studies to Estimate Linear Models." *Journal of Educational and Behavioral Statistics*, **20**(1), 100-102. doi:10.2307/1165390

Examples

```

data(Becker92)

#### Fixed-effects model
## First stage analysis
## Replicate Becker's (1992) analysis using 4 studies only

fixed1 <- tssem1(Becker92$data[1:4], Becker92$n[1:4], method="FEM")
summary(fixed1)

## ## Prepare a regression model using create.mxMatrix()
## A1 <- create.mxMatrix(c(0,0,0,"0.2*Spatial2Math",
##                        0,0,"0.2*Verbal2Math",0,0), type="Full",
##                        ncol=3, nrow=3, as.mxMatrix=FALSE)

## var.names <- c("Math_apptitude","Spatial","Verbal")

## ## This step is not necessary but it is useful for inspecting the model.
## dimnames(A1)[[1]] <- dimnames(A1)[[2]] <- var.names

## ## Display A1
## A1

## S1 <- create.mxMatrix(c("0.2*ErrorVarMath",0,0,1,"0.2*CorSpatialVerbal",1),
##                        type="Symm", as.mxMatrix=FALSE)

## ## This step is not necessary but it is useful for inspecting the model.
## dimnames(S1)[[1]] <- dimnames(S1)[[2]] <- var.names

## ## Display S1
## S1

#####
## Alternative model specification in lavaan model syntax
model <- "## Regression paths
         Math ~ Spatial2Math*Spatial + Verbal2Math*Verbal
         Spatial ~~ CorSpatialVerbal*Verbal
         ## Fix the variances of Spatial and Verbal at 1
         Spatial ~~ 1*Spatial
         Verbal ~~ 1*Verbal
         ## Label the error variance of Math
         Math ~~ ErrorVarMath*Math + start(0.2)*Math"

## Display the model
plot(model)

RAM <- lavaan2RAM(model, obs.variables=c("Math", "Spatial", "Verbal"))
RAM
#####

```

```
## Fixed-effects model: Second stage analysis
## Two equivalent versions to calculate the R2 and its 95% LBCI
fixed2 <- tssem2(fixed1, RAM=RAM, intervals.type="LB",
                 mx.algebras=list(R1=mxAlgebra(Spatial2Math^2+Verbal2Math^2
                 +2*CorSpatialVerbal*Spatial2Math*Verbal2Math, name="R1"),
                 R2=mxAlgebra(One-Smatrix[1,1], name="R2"),
                 One=mxMatrix("Iden", ncol=1, nrow=1, name="One")))

summary(fixed2)

## Display the model with the parameter estimates
plot(fixed2)

#### Random-effects model
## First stage analysis
## No random effects for off-diagonal elements
random1 <- tssem1(Becker92$data, Becker92$n, method="REM", RE.type="Diag")
summary(random1)

## Random-effects model: Second stage analysis
random2 <- tssem2(random1, RAM=RAM)
summary(random2)

## Display the model with the parameter estimates
plot(random2, color="yellow")

#### Similar to conventional fixed-effects GLS approach
## First stage analysis
## No random effects
## Replicate Becker's (1992) analysis using 4 studies only
gls1 <- tssem1(Becker92$data[1:4], Becker92$n[1:4], method="REM", RE.type="Zero",
               model.name="Fixed effects GLS Stage 1")
summary(gls1)

## Fixed-effects GLS model: Second stage analysis
gls2 <- tssem2(gls1, RAM=RAM, model.name="Fixed effects GLS Stage 2")
summary(gls2)
```

Becker94

Five Studies of Ten Correlation Matrices reported by Becker and Schram (1994)

Description

This data set includes five studies of ten correlation matrices reported by Becker and Schram (1994).

Details

A list of data with the following structure:

data A list of 10 correlation matrices. The variables are *Math* (math aptitude), *Spatial* (spatial ability), and *Verbal* (verbal ability)

n A vector of sample sizes

gender *Females* or *Males* samples

Source

Becker, B. J., & Schram, C. M. (1994). Examining explanatory models through research synthesis. In H. Cooper & L. V. Hedges (Eds.), *The handbook of research synthesis* (pp. 357-381). New York: Russell Sage Foundation.

Examples

```
data(Becker94)

#### Fixed-effects model
## First stage analysis
fixed1 <- tssem1(Becker94$data, Becker94$n, method="FEM")
summary(fixed1)

## Prepare a regression model using create.mxMatrix()
## A1 <- create.mxMatrix(c(0,0,0,"0.2*Spatial2Math",
##                        0,0,"0.2*Verbal2Math",0,0), type="Full",
##                        ncol=3, nrow=3, name="A1")
## S1 <- create.mxMatrix(c("0.2*ErrorVarMath",0,0,1,
##                        "0.2*CorBetweenSpatialVerbal",1),
##                        type="Symm", name="S1")

## An alternative method to create a regression model with the lavaan syntax
model <- "## Regression model
        Math ~ Spatial2Math*Spatial + Verbal2Math*Verbal
        ## Error variance of Math
        Math ~~ ErrorVarMath*Math
        ## Variances of Spatial and Verbal fixed at 1.0
        Spatial ~~ 1*Spatial
        Verbal ~~ 1*Verbal
        ## Correlation between Spatial and Verbal
        Spatial ~~ CorBetweenSpatialVerbal*Verbal"

## Display the model
plot(model)

RAM <- lavaan2RAM(model, obs.variables=c("Math", "Spatial", "Verbal"))
RAM

## Second stage analysis
## A1 <- RAM$A
## S1 <- RAM$S
## fixed2 <- tssem2(fixed1, Amatrix=A1, Smatrix=S1, intervals.type="LB")

fixed2 <- tssem2(fixed1, RAM=RAM, intervals.type="LB")
```

```

summary(fixed2)

## Display the model with the parameter estimates
plot(fixed2)

#### Fixed-effects model: with gender as cluster
## First stage analysis
cluster1 <- tssem1(Becker94$data, Becker94$n, method="FEM", cluster=Becker94$gender)
summary(cluster1)

## Second stage analysis
cluster2 <- tssem2(cluster1, RAM=RAM, intervals.type="LB")
summary(cluster2)

#### Conventional fixed-effects GLS approach
## First stage analysis
## No random effects
## Replicate Becker's (1992) analysis using 4 studies only
gls1 <- tssem1(Becker92$data[1:4], Becker92$n[1:4], method="REM", RE.type="Zero",
               model.name="Fixed effects GLS Stage 1")
summary(gls1)

## Fixed-effects GLS model: Second stage analysis
gls2 <- tssem2(gls1, RAM=RAM, intervals.type="LB",
               model.name="Fixed effects GLS Stage 2")
summary(gls2)

```

Berkey98

Five Published Trials from Berkey et al. (1998)

Description

The data set includes five published trials, reported by Berkey et al. (1998), comparing surgical and non-surgical treatments for medium-severity periodontal disease, one year after treatment.

Details

The variables are:

trial Trial number

pub_year Publication year

no_of_patients Number of patients

PD Patient improvements (mm) in *probing depth*

AL Patient improvements (mm) in *attachment level*

var_PD Sampling variance of PD

cov_PD_AL Sampling covariance between PD and AL

var_AL Sampling variance of AL

Source

Berkey, C. S., Hoaglin, D. C., Antczak-Bouckoms, A., Mosteller, F., & Colditz, G. A. (1998). Meta-analysis of multiple outcomes by regression with random effects. *Statistics in Medicine*, **17**, 2537-2550.

Examples

```
data(Berkey98)

#### ML estimation method
## Multivariate meta-analysis
x <- meta(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL), data=Berkey98)
x <- rerun(x)
summary(x)
plot(x)

## Plot individual studies proportional to the weights
plot(x, study.weight.plot=TRUE)

## Include forest plot from the metafor package
library(metafor)
plot(x, diag.panel=TRUE, main="Multivariate meta-analysis",
      axis.label=c("PD", "AL"))
  forest( rma(yi=PD, vi=var_PD, data=Berkey98) )
  title("Forest plot of PD")
  forest( rma(yi=AL, vi=var_AL, data=Berkey98) )
  title("Forest plot of AL")

## Multivariate meta-analysis with "publication year-1979" as the predictor
summary( meta(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL),
          x=scale(pub_year, center=1979), data=Berkey98,
          RE.lbound=NA) )

## Multivariate meta-analysis with equality constraint on the regression coefficients
summary( meta(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL),
          x=scale(pub_year, center=1979), data=Berkey98,
          coef.constraints=matrix(c("0.3*Eq_slope", "0.3*Eq_slope"),
          nrow=2)) )

#### REML estimation method
## Multivariate meta-analysis
summary( reml(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL),
            data=Berkey98,
            model.name="Multivariate meta analysis with REML") )

## Multivariate meta-analysis with "publication year-1979" as the predictor
## Diagonal structure for the variance component
summary( reml(y=cbind(PD, AL), v=cbind(var_PD, cov_PD_AL, var_AL),
            RE.constraints=Diag(c("1e-5*Tau2_1_1", "1e-5*Tau2_2_2")),
            x=scale(pub_year, center=1979), data=Berkey98) )
```

Boer16

Correlation Matrices from Boer et al. (2016)

Description

The data set includes correlation matrices of leader-member exchange in transformational leadership reported by Boer et al. (2016).

Details

A list of data with the following structure:

data A list of correlation matrices. The variables are *LMX* (leader-member exchange), *TFL* (transformational leadership), *JS* (job satisfaction), *OC* (organizational commitment), and *LE* (leader effectiveness)

n A vector of sample sizes

RelLMX The reliability of *LMX*

RelTFL The reliability of *TFL*

Source

Boer, D., Deinert, A., Homan, A. C., & Voelpel, S. C. (2016). Revisiting the mediating role of leader-member exchange in transformational leadership: the differential impact model. *European Journal of Work and Organizational Psychology*, **25**(6), 883-899.

Examples

```
## Stage 1 analysis
rand1 <- tssem1(Boer16$data, Boer16$n, method="REM", RE.type="Diag",
               acov="weighted")
summary(rand1)

## Stage 2 analysis
model2a <- 'JS+OC+LE ~ LMX+TFL
           LMX ~ TFL
           ## Variance of TFL is fixed at 1
           TFL ~~ 1*TFL
           ## Correlated residuals
           JS ~~ OC
           JS ~~ LE
           OC ~~ LE'

## Display the model
plot(model2a)
```

```

RAM2a <- lavaan2RAM(model2a, obs.variables = c("LMX", "TFL", "JS", "OC", "LE"),
  A.notation="on", S.notation="with")

rand2a <- tssem2(rand1, RAM=RAM2a)
summary(rand2a)

## Display the model with the parameter estimates
plot(rand2a, layout="spring")

```

bootuniR1

*Parametric bootstrap on the univariate R (uniR) object***Description**

It generates correlation matrices with the parametric bootstrap on the univariate R (uniR) object.

Usage

```
bootuniR1(x, Rep, nonPD.pop = c("replace", "nearPD", "accept"))
```

Arguments

x	An object of class 'uniR1'
Rep	Number of replications of the parametric bootstrap
nonPD.pop	If it is replace, generated non-positive definite matrices are replaced by generated new ones which are positive definite. If it is nearPD, they are replaced by nearly positive definite matrices by calling <code>Matrix::nearPD()</code> . If it is accept, they are accepted.

Details

This function implements the parametric bootstrap approach suggested by Yu et al. (2016). It is included in this package for research interests. Please refer to Cheung (2018) for the issues associated with this parametric bootstrap approach.

Value

An object of the generated correlation matrices.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Cheung, M. W.-L. (2018). Issues in solving the problem of effect size heterogeneity in meta-analytic structural equation modeling: A commentary and simulation study on Yu, Downes, Carter, and O'Boyle (2016). *Journal of Applied Psychology*, **103**, 787-803.

Yu, J. (Joya), Downes, P. E., Carter, K. M., & O'Boyle, E. H. (2016). The problem of effect size heterogeneity in meta-analytic structural equation modeling. *Journal of Applied Psychology*, **101**, 1457-1473.

See Also

[rCor](#), [bootuniR2](#), [Nohe15](#)

bootuniR2

Fit Models on the bootstrapped correlation matrices

Description

It fits structural equation models on the bootstrapped correlation matrices.

Usage

```
bootuniR2(model, data, n, ...)
```

Arguments

model	A model in sem syntax.
data	A list of correlation matrices.
n	Sample size in fitting the structural equation models
...	Further arguments to be passed to sem .

Details

This function fits the lavaan model with the bootstrapped correlation matrices. It implements the parametric bootstrap approach suggested by Yu et al. (2016). It is included in this package for research interests. Please refer to Cheung (2018) for the issues associated with this parametric bootstrap approach.

Value

A list of the fitted object from [sem](#).

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2018). Issues in solving the problem of effect size heterogeneity in meta-analytic structural equation modeling: A commentary and simulation study on Yu, Downes, Carter, and O'Boyle (2016). *Journal of Applied Psychology*, **103**, 787-803.
- Yu, J. (Joya), Downes, P. E., Carter, K. M., & O'Boyle, E. H. (2016). The problem of effect size heterogeneity in meta-analytic structural equation modeling. *Journal of Applied Psychology*, **101**, 1457-1473.

See Also

[bootuniR2](#), [tssemParaVar](#), [Nohe15](#)

Bornmann07

A Dataset from Bornmann et al. (2007)

Description

A dataset from Bornmann et al. (2007) for three-level meta-analysis.

Details

The variables are:

ID ID of the study

Study Study name

Cluster Cluster for effect sizes

logOR Effect size: log odds ratio

v Sampling variance of logOR

Year Year of publication

Type Type of proposal: either **Grant** or **Fellowship**

Discipline Discipline of the proposal: either **Physical sciences**, **Life sciences/biology**, **Social sciences/humanities** or **Multidisciplinary**

Country Country of the proposal: either the **United States**, **Canada**, **Australia**, **United Kingdom** or **Europe**

Source

Bornmann, L., Mutz, R., & Daniel, H.-D. (2007). Gender differences in grant peer review: A meta-analysis. *Journal of Informetrics*, **1**(3), 226-238. doi:10.1016/j.joi.2007.03.001

References

- Cheung, M. W.-L. (2014). Modeling dependent effect sizes with three-level meta-analyses: A structural equation modeling approach. *Psychological Methods*, **19**, 211-229.
- Marsh, H. W., Bornmann, L., Mutz, R., Daniel, H.-D., & O'Mara, A. (2009). Gender Effects in the Peer Reviews of Grant Proposals: A Comprehensive Meta-Analysis Comparing Traditional and Multilevel Approaches. *Review of Educational Research*, **79**(3), 1290-1326. doi:10.3102/0034654309334143

Examples

```

data(Bornmann07)

#### ML estimation method
## No predictor
summary( meta3L(y=logOR, v=v, cluster=Cluster, data=Bornmann07) )

## Type as a predictor
## Grant: 0
## Fellowship: 1
summary( meta3L(y=logOR, v=v, x=(as.numeric(Type)-1),
                cluster=Cluster, data=Bornmann07) )

## Centered Year as a predictor
summary( meta3L(y=logOR, v=v, x=scale(Year, scale=FALSE),
                cluster=Cluster, data=Bornmann07) )

#### REML estimation method
## No predictor
summary( reml3L(y=logOR, v=v, cluster=Cluster, data=Bornmann07) )

## Type as a predictor
## Grants: 0
## Fellowship: 1
summary( reml3L(y=logOR, v=v, x=(as.numeric(Type)-1),
                cluster=Cluster, data=Bornmann07) )

## Centered Year as a predictor
summary( reml3L(y=logOR, v=v, x=scale(Year, scale=FALSE),
                cluster=Cluster, data=Bornmann07) )

## Handling missing covariates with FIML
## MCAR
## Set seed for replication
set.seed(1000000)

## Copy Bornmann07 to my.df
my.df <- Bornmann07
## "Fellowship": 1; "Grant": 0
my.df$Type_MCAR <- ifelse(Bornmann07$Type=="Fellowship", yes=1, no=0)

## Create 17 out of 66 missingness with MCAR
my.df$Type_MCAR[sample(1:66, 17)] <- NA
summary(meta3LFIML(y=logOR, v=v, cluster=Cluster, x2=Type_MCAR, data=my.df))

## MAR
Type_MAR <- ifelse(Bornmann07$Type=="Fellowship", yes=1, no=0)

## Create 27 out of 66 missingness with MAR for cases Year<1996
index_MAR <- ifelse(Bornmann07$Year<1996, yes=TRUE, no=FALSE)
Type_MAR[index_MAR] <- NA

```

```
## Include auxiliary variable
summary(meta3LFIML(y=logOR, v=v, cluster=Cluster, x2=Type_MAR, av2=Year, data=my.df))
```

calEffSizes

Calculate Effect Sizes using lavaan Models

Description

It calculates effect sizes with Delta Method by formulating the effect sizes as functions of SEM in lavaan.

Usage

```
calEffSizes(
  model,
  data = NULL,
  n,
  Cov,
  Mean = NULL,
  group = NULL,
  lavaan.output = FALSE,
  warn = FALSE,
  ...
)
```

Arguments

model	A lavaan model. Effect sizes are defined as functions of SEM parameters with <code>:=</code> .
data	A data frame of the observed variables. If it is NULL, summary statistics are required.
n	Sample sizes
Cov	A covariance matrix or a list of covariance matrices.
Mean	Optional sample means.
group	A character of the variable name in the data frame defining the groups in a multiple group analysis.
lavaan.output	If TRUE, it returns the fitted object instead of the effect sizes and their sampling covariance matrix.
warn	If FALSE, it suppresses lavaan related warnings.
...	Further arguments passed to sem .

Value

Effect sizes and their sampling covariance matrix or a lavaan fitted object.

Note

The input matrices are treated as covariance matrices unless there are explicit constraints in the model.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Cheung, M. W.-L. (2015). *Meta-analysis: A structural equation modeling approach*. Chichester, West Sussex: John Wiley & Sons, Inc.

Cheung, M. W.-L. (2018). Computing multivariate effect sizes and their sampling covariance matrices with structural equation modeling: Theory, examples, and computer simulations. *Frontiers in Psychology*, *9*(1387). <https://doi.org/10.3389/fpsyg.2018.01387>

See Also

[smdMES](#), [smdMTS](#)

Examples

```
## Select ATT, BI, and BEH
obs.vars <- c("BEH", "BI", "ATT")

## Select one study from Cooke16 for illustration
my.cor <- Cooke16$data[[4]][obs.vars, obs.vars]
my.n <- Cooke16$n[4]

## Effect sizes: indirect effect and direct effect
model <- "BEH ~ c*ATT + b*BI
          BI ~ a*ATT
          ## Indirect effect
          Ind := a*b
          Dir := c"

calEffSizes(model=model, n=my.n, Cov=my.cor, lavaan.output=FALSE)

## Return the lavaan fitted model
fit <- calEffSizes(model=model, n=my.n, Cov=my.cor, lavaan.output=TRUE)
lavaan::summary(fit)

lavaan::parameterestimates(fit)
```

 Chan17

Dataset from Chan, Jones, Jamieson, and Albarracin (2017)

Description

A dataset of multiple treatment effects of standardized mean differences on misinformation and debunking effects.

Format

A data frame with 34 independent samples from 6 research reports.

Author a character vector of study

g_misinfo Hedges' g of misinformation comparing the misinformation experimental and control groups

g_debunk Hedges' g of debunking comparing the debunking experimental and misinformation experimental groups

v_misinfo sampling variance of g_misinfo

c_mis_deb Sampling covariance between g_misinfo and g_debunk due to the overlap of the misinformation experimental group

v_debunk sampling variance of g_debunk

PublicationYear publication year

Published published or unpublished

MeanAge mean age of participants

PctFemale percentage of female participants

Details

The sampling variances and covariances are calculated using Gleser and Olkin's (2009) method for multiple treatment effects (Equations 3.3 and 3.4). Since the sample sizes of the misinformation, debunking, and control groups are not given, it is assumed they are equal.

Source

Chan, M. S., Jones, C. R., Hall Jamieson, K., & Albarracin, D. (2017). Debunking: A meta-analysis of the psychological efficacy of messages countering misinformation. *Psychological Science*, **28**(11), 1531-1546. <https://doi.org/10.1177/0956797617714579>

References

Gleser, L. J., & Olkin, I. (2009). Stochastically dependent effect sizes. In H. Cooper, L. V. Hedges, & J. C. Valentine (Eds.), *The handbook of research synthesis and meta-analysis*. (2nd ed., pp. 357-376). Russell Sage Foundation.

checkRAM	<i>Check the correctness of the RAM formulation</i>
----------	---

Description

It provides simple checks on the correctness of the RAM formulation.

Usage

```
checkRAM(Amatrix, Smatrix, cor.analysis = TRUE)
```

Arguments

Amatrix	An asymmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Smatrix	A symmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
cor.analysis	Logical. Analysis of correlation or covariance structure. There are additional checks for <code>cor.analysis=TRUE</code> .

Value

It returns silently if no error has been detected; otherwise, it returns a warning message.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[as.mxMatrix](#), [lavaan2RAM](#)

Examples

```
## Digman97 example
model1 <- "### Factor loadings
          Alpha=~A+C+ES
          Beta=~E+I
          ## Factor correlation
          Alpha~~Beta"

RAM1 <- lavaan2RAM(model1, obs.variables=c("A","C","ES","E","I"),
                  A.notation="on", S.notation="with")
RAM1

## The model is okay.
checkRAM(Amatrix=RAM1$A, Smatrix=RAM1$S)
```

```
## Hunter83 example
model2 <- "## Regression paths
          Job_knowledge ~ A2J*Ability
          Work_sample ~ A2W*Ability + J2W*Job_knowledge
          Supervisor ~ J2S*Job_knowledge + W2S*Work_sample

          ## Fix the variance of Ability at 1
          Ability ~~ 1*Ability

          ## Label the error variances of the dependent variables
          Job_knowledge ~~ VarE_J*Job_knowledge
          Work_sample ~~ VarE_W*Work_sample
          Supervisor ~~ VarE_S*Supervisor"

RAM2 <- lavaan2RAM(model2, obs.variables=c("Ability","Job_knowledge",
                                           "Work_sample","Supervisor"))

## The model is okay.
checkRAM(Amatrix=RAM2$A, Smatrix=RAM2$S)
```

Cheung00

Fifty Studies of Correlation Matrices used in Cheung and Chan (2000)

Description

This data set includes fifty studies of correlation matrices on the theory of planned theory reported by Cheung and Chan (2000).

Details

A list of data with the following structure:

data A list of 50 studies of correlation matrices. The variables are the attitude toward behavior *att*, subjective norm *sn*, behavioral intention *bi*, and behavior *beh*

n A vector of sample sizes

Note

These studies were extracted from the original data set for illustration purpose. Some samples contained two or more correlation matrices, and only one of them was arbitrarily selected to avoid the problem of dependence. Moreover, studies with less than 3 correlation coefficients were also excluded.

Source

Cheung, S.-F., & Chan, D. K.-S. (2000). The role of perceived behavioral control in predicting human behavior: A meta-analytic review of studies on the theory of planned behavior. *Unpublished manuscript*, Chinese University of Hong Kong.

References

Cheung, M.W.-L., & Cheung, S.-F. (2016). Random-effects models for meta-analytic structural equation modeling: Review, issues, and illustrations. *Research Synthesis Methods*, **7**, 140-155.

Examples

```
data(Cheung00)

## Full mediation model in lavaan syntax
model <- "## Regression paths
        bi ~ att2bi*att + sn2bi*sn
        beh ~ bi2beh*bi
        ## Variances of att and sn are fixed at 1
        att ~~ 1*att
        sn ~~ 1*sn
        ## Covariance between att and sn
        att ~~ cov_att_sn*sn
        ## Error variances
        bi ~~ e_bi*bi
        beh ~~ e_beh*beh"

RAM <- lavaan2RAM(model, obs.variables=colnames(Cheung00$data[[1]]))
RAM

## ## Equivalent RAM specification
## labels <- colnames(Cheung00$data[[1]])
## A <- matrix(c("0","0","0","0",
##               "0","0","0","0",
##               ".2*att2bi", ".2*sn2bi", "0", "0",
##               "0", "0", ".2*bi2beh", "0"),
##             byrow=TRUE, 4, 4)
## dimnames(A) <- list(labels, labels)
## A
##
## S <- create.mxMatrix(c("1",
##                         ".2*cov_att_sn", "1",
##                         0, 0, ".2*e_bi",
##                         0, 0, 0, ".2*e_beh"),
##                       type="Symm", as.mxMatrix=FALSE, byrow=TRUE)
## dimnames(S) <- list(labels, labels)
## S

#### Random-effects model

## Stage 1 analysis
random_1 <- tssem1(Cheung00$data, Cheung00$n, method="REM", RE.type="Diag",
                  acov="weighted")
summary(random_1)

## Stage 2 analysis
random_2 <- tssem2(random_1, RAM=RAM, intervals.type="LB",
```

```

                                diag.constraints=TRUE)
summary(random_2)

## Display the model
plot(random_2, what="path")

## Display the model with the parameter estimates
plot(random_2, color="yellow")

## Load the library
library("semPlot")

```

Cheung09

A Dataset from TSSEM User's Guide Version 1.11 by Cheung (2009)

Description

Four studies were selected from the data set used by Cheung and Chan (2005; 2009). Some variables were randomly deleted to illustrate the analysis with missing data.

Details

A list of data with the following structure:

data A list of 4 studies of correlation matrices

n A vector of sample sizes

References

Cheung, M. W.-L., & Chan, W. (2005). Meta-analytic structural equation modeling: A two-stage approach. *Psychological Methods*, **10**, 40-64.

Cheung, M. W.-L., & Chan, W. (2009). A two-stage approach to synthesizing covariance matrices in meta-analytic structural equation modeling. *Structural Equation Modeling*, **16**, 28-53.

Examples

```

data(Cheung09)

#### Fixed-effects model: Stage 1 analysis
fixed1 <- tssem1(Cheung09$data, Cheung09$n, method="FEM")
summary(fixed1)

## Three-factor CFA model in lavaan syntax
model <- "## Factor loadings
          f1 =~ f1x1*x1 + f1x2*x2 + f1x3*x3
          f2 =~ f2x4*x4 + f2x5*x5 + f2x6*x6 + f2x7*x7

```

```

      f3 =~ f3x8*x8 + f3x9*x9
      ## Factor correlations
      f1 ~~ corf2f1*f2
      f1 ~~ corf3f1*f3
      f2 ~~ corf3f2*f3"

RAM1 <- lavaan2RAM(model, obs.variables=paste0("x", 1:9))
RAM1

## ## Equivalent RAM specification using create.mxMatrix()
## Phi <- create.mxMatrix( c("0.3*corf2f1","0.3*corf3f1","0.3*corf3f2"),
##                          type="Stand", as.mxMatrix=FALSE )
## Psi <- create.mxMatrix( paste("0.2*e", 1:9, sep=""), type="Diag",
##                          as.mxMatrix=FALSE )
## S1 <- bdiagMat(list(Psi, Phi))
## S1 <- as.mxMatrix(S1)
##
## Lambda <- create.mxMatrix( c(".3*f1x1",".3*f1x2",".3*f1x3",rep(0,9),
##                              ".3*f2x4",".3*f2x5",".3*f2x6",".3*f2x7",
##                              rep(0,9),".3*f3x8",".3*f3x9"), type="Full",
##                              ncol=3, nrow=9, as.mxMatrix=FALSE )
## Zero1 <- matrix(0, nrow=9, ncol=9)
## Zero2 <- matrix(0, nrow=3, ncol=12)
## A1 <- rbind( cbind(Zero1, Lambda), Zero2 )
## A1 <- as.mxMatrix(A1)
##
## F1 <- create.Fmatrix(c(rep(1,9), rep(0,3)))

#### Fixed-effects model: Stage 2 analysis
fixed2 <- tssem2(fixed1, RAM=RAM1, intervals.type="LB")
summary(fixed2)

## Display the model
plot(fixed2, what="path")

## Display the model with the parameter estimates
plot(fixed2, latNames=c("f1", "f2", "f3"), edge.label.cex=0.8,
      color="yellow")

```

coef

*Extract Parameter Estimates from various classes.***Description**

It extracts the parameter estimates from objects of various classes.

Arguments

<code>object</code>	An object returned from either class <code>tssem1FEM</code> , class <code>tssem1FEM.cluster</code> , class <code>tssem1REM</code> , class <code>wls</code> , class <code>wls.cluster</code> , class <code>meta</code> , class <code>reml</code> , class <code>osmasem</code> , class <code>osmasem2</code> , or class <code>sem</code>
<code>select</code>	Select all for both fixed- and random-effects parameters, fixed for the fixed-effects parameters or random for the random-effects parameters. For <code>meta3LFIML</code> objects, <code>allX</code> is used to extract all parameters including the predictors and auxiliary variables.
<code>...</code>	Further arguments; currently none is used

Value

Parameter estimates for both fixed-effects (if any) and random-effects (if any)

Note

`coef.sem` is simply a wrapper of `omxGetParameters`. Extra arguments will be passed to it

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[tssem1](#), [wls](#), [meta](#), [reml](#), [omxGetParameters](#), [osmasem](#)

Examples

```
## Random-effects meta-analysis
model1 <- meta(y=yi, v=vi, data=Hox02)
coef(model1)

## Fixed-effects only
coef(model1, select="fixed")
```

Description

The data set includes correlation matrices on using the theory of planned behavior to predict alcohol consumption reported by Cooke et al. (2016).

Details

A list of data with the following structure:

data A list of correlation matrices. The variables are *SN* (subjective norm), *ATT* (attitude), *PBC* (perceived behavior control), *BI* (behavioral intention), and *BEH* (behavior).

n A vector of sample sizes.

MeanAge Mean age of the participants except for Ajzen and Sheikh (2013), which is the median age, and Glassman, et al. (2010a) to Glassman, et al. (2010d), which are based on the range of 18 to 24.

Female Percentage of female participants.

Source

Cooke, R., Dahdah, M., Norman, P., & French, D. P. (2016). How well does the theory of planned behaviour predict alcohol consumption? A systematic review and meta-analysis. *Health Psychology Review*, **10**(2), 148-167.

References

Cheung, M. W.-L., & Hong, R. Y. (2017). Applications of meta-analytic structural equation modeling in health psychology: Examples, issues, and recommendations. *Health Psychology Review*, **11**, 265-279.

Examples

```
## Check whether the correlation matrices are valid (positive definite)
Cooke16$data[is.pd(Cooke16$data)==FALSE]

## Since the correlation matrix in Study 3 is not positive definite,
## we exclude it in the following analyses
my.data <- Cooke16$data[-3]
my.n <- Cooke16$n[-3]

## Show the no. of studies per correlation
pattern.na(my.data, show.na = FALSE)

## Show the total sample sizes per correlation
pattern.n(my.data, my.n)

## Stage 1 analysis
## Random-effects model
random1 <- tssem1(my.data, my.n, method="REM", RE.type="Diag", acov="weighted")
summary(random1)

## Model specification in lavaan syntax
model <- "## Regression paths
          BI ~ SN2BI*SN + ATT2BI*ATT + PBC2BI*PBC
          BEH ~ PBC2BEH*PBC + BI2BEH*BI
          ## Variances of SN, ATT, and PBC are fixed at 1"
```

```

SN ~~ 1*SN
ATT ~~ 1*ATT
PBC ~~ 1*PBC
## Covariances among SN, ATT, and PBC
ATT ~~ ATT_SN*SN
PBC ~~ PBC_SN*SN
PBC ~~ PBC_ATT*ATT
## Error variances of BI and BEH
BI ~~ VarBI*BI
BEH ~~ VarBEH*BEH"

RAM1 <- lavaan2RAM(model, obs.variables=colnames(Cooke16$data[[1]]))
RAM1

## ## Equivalent RAM specification using create.mxMatrix()
## A1 <- create.mxMatrix(c(0,0,0,0,0,
##                          0,0,0,0,0,
##                          0,0,0,0,0,
##                          "0.2*SN2BI", "0.2*ATT2BI", "0.2*PBC2BI", 0,0,
##                          0,0, "0.2*PBC2BEH", "0.2*BI2BEH", 0),
##                        type="Full", ncol=5, nrow=5,
##                        byrow=TRUE, as.mxMatrix=FALSE)
## dimnames(A1)[[1]] <- dimnames(A1)[[2]] <- colnames(Cooke16$data[[1]])
## A1
##
## S1 <- create.mxMatrix(c(1,
##                          "0.1*ATT_SN", 1,
##                          "0.1*PBC_SN", "0.1*PBC_ATT", 1,
##                          0, 0, 0, "0.5*VarBI",
##                          0, 0, 0, 0, "0.5*VarBEH"),
##                        type = "Symm", ncol=5, nrow=5,
##                        byrow=TRUE, as.mxMatrix=FALSE)
## dimnames(S1)[[1]] <- dimnames(S1)[[2]] <- colnames(Cooke16$data[[1]])
## S1

## Stage 2 analysis
random2 <- tssem2(random1, RAM=RAM1, diag.constraints=FALSE,
                  intervals.type="LB")
summary(random2)

## Display the model
plot(random2, what="path")

## Display the model with the parameter estimates
plot(random2, color="yellow")

```

Description

Fifty-six effect sizes from 11 districts from Cooper et al. (2003) were reported by Konstantopoulos (2011).

Details

The variables are:

District District ID

Study Study ID

y Effect size

v Sampling variance

Year Year of publication

Source

Cooper, H., Valentine, J. C., Charlton, K., & Melson, A. (2003). The Effects of Modified School Calendars on Student Achievement and on School and Community Attitudes. *Review of Educational Research*, **73**(1), 1-52. doi:10.3102/00346543073001001

References

Konstantopoulos, S. (2011). Fixed effects and variance components estimation in three-level meta-analysis. *Research Synthesis Methods*, **2**, 61-76. doi:10.1002/jrsm.35

Examples

```
data(Cooper03)

#### ML estimation method
## No predictor
summary( model1 <- meta3L(y=y, v=v, cluster=District, data=Cooper03) )

## Show all heterogeneity indices and their 95% confidence intervals
summary( meta3L(y=y, v=v, cluster=District, data=Cooper03,
               intervals.type="LB", I2=c("I2q", "I2hm", "I2am", "ICC")) )

## Year as a predictor
summary( meta3L(y=y, v=v, cluster=District, x=scale(Year, scale=FALSE),
               data=Cooper03, model.name="Year as a predictor") )

## Equality of level-2 and level-3 heterogeneity
summary( model2 <- meta3L(y=y, v=v, cluster=District, data=Cooper03,
               RE2.constraints="0.2*EqTau2",
               RE3.constraints="0.2*EqTau2",
               model.name="Equal Tau2") )

## Compare model2 vs. model1
anova(model1, model2)
```

```
#### REML estimation method
## No predictor
summary( reml3L(y=y, v=v, cluster=District, data=Cooper03) )

## Level-2 and level-3 variances are constrained equally
summary( reml3L(y=y, v=v, cluster=District, data=Cooper03,
               RE.equal=TRUE, model.name="Equal Tau2") )

## Year as a predictor
summary( reml3L(y=y, v=v, cluster=District, x=scale(Year, scale=FALSE),
               data=Cooper03, intervals.type="LB") )

## Handling missing covariates with FIML
## Create 20/56 MCAR data in Year
set.seed(10000)
Year_MCAR <- Cooper03$Year
Year_MCAR[sample(56, 20)] <- NA
summary( meta3LFIML(y=y, v=v, cluster=District, x2=scale(Year_MCAR, scale=FALSE),
                   data=Cooper03, model.name="NA in Year_MCAR") )
```

Cor2DataFrame

Convert correlation or covariance matrices into a dataframe of correlations or covariances with their sampling covariance matrices

Description

It converts the correlation or covariance matrices into a dataframe of correlations or covariances with their asymptotic sampling covariance matrices. It uses the asyCov at the backend.

Usage

```
Cor2DataFrame(
  x,
  n,
  v.na.replace = TRUE,
  cor.analysis = TRUE,
  acov = c("weighted", "individual", "unweighted"),
  Means,
  row.names.unique = FALSE,
  append.vars = TRUE,
  asyCovOld = FALSE,
  ...
)
```

Arguments

<code>x</code>	A list of data with correlation/covariance matrix in <code>x\$data</code> and sample sizes <code>x\$n</code> . Additional variables in <code>x</code> can be attached.
<code>n</code>	If <code>x</code> is a list of correlation matrices without <code>x\$data</code> and <code>x\$n</code> , a vector of sample sizes <code>n</code> must be provided.
<code>v.na.replace</code>	Logical. Missing value is not allowed in definition variables. If it is TRUE (the default), missing value is replaced by a large value (1e10). These values are not used in the analysis.
<code>cor.analysis</code>	Logical. The output is either a correlation or covariance matrix.
<code>acov</code>	If it is weighted, the average correlation/covariance matrix is calculated based on the weighted mean with the sample sizes. The average correlation/covariance matrix is used to calculate the sampling variance-covariance matrices.
<code>Means</code>	An optional matrix of means. The number of rows must be the same as the length of <code>n</code> . The sampling covariance matrices of the means are calculated by the covariance matrices divided by the sample sizes. Therefore, it is important to make sure that covariance matrices (not correlation matrices) are used in <code>x</code> when Means are included; otherwise, the calculated sampling covariance matrices of the means are incorrect.
<code>row.names.unique</code>	Logical. If it is FALSE (the default), unique row names are not created.
<code>append.vars</code>	Whether to append the additional variables to the output dataframe.
<code>asyCovOld</code>	Whether to use the old version of <code>asyCov</code> . See asyCov .
<code>...</code>	Further arguments to be passed to asyCov .

Value

A list of components: (1) a data frame of correlations or covariances with their sampling covariance matrices; (2) a vector of sample sizes; (3) labels of the correlations; and (4) labels of their sampling covariance matrices.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[asyCov](#), [osmasem](#), [create.vechsR](#), [create.Tau2](#), [create.V](#)

Examples

```
## Provide a list of correlation matrices and a vector of sample sizes as the inputs
my.df1 <- Cor2DataFrame(Nohe15A1$data, Nohe15A1$n)

## Add Lag time as a variable
my.df1$data <- data.frame(my.df1$data, Lag=Nohe15A1$Lag, check.names=FALSE)
```

```
## Data
my.df1$data

## Sample sizes
my.df1$n

## ylabels
my.df1$ylabels

## vlabels
my.df1$vlabels

#### Simplified version to do it
my.df2 <- Cor2DataFrame(Nohe15A1)
```

create.Fmatrix	<i>Create an F matrix to select observed variables</i>
----------------	--

Description

It creates an F matrix to select observed variables for wls function.

Usage

```
create.Fmatrix(x, name, as.mxMatrix = TRUE, ...)
```

Arguments

x	A vector of logical type
name	Name of the matrix. If it is missing, "Fmatrix" will be used.
as.mxMatrix	Logical. If it is TRUE, the output is a matrix of MxMatrix-class. If it is FALSE, it is a numeric matrix.
...	Not used.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[as.mxMatrix](#), [create.mxMatrix](#), [wls](#)

Examples

```
## Select the first 3 variables while the other 2 variables are latent.
create.Fmatrix(c(1,1,1,0,0))
# FullMatrix 'Fmatrix'
#
# @labels: No labels assigned.
#
# @values
#      [,1] [,2] [,3] [,4] [,5]
# [1,]    1    0    0    0    0
# [2,]    0    1    0    0    0
# [3,]    0    0    1    0    0
#
# @free: No free parameters.
#
# @lbound: No lower bounds assigned.
#
# @ubound: No upper bounds assigned.

create.Fmatrix(c(1,1,1,0,0), as.mxMatrix=FALSE)
#      [,1] [,2] [,3] [,4] [,5]
# [1,]    1    0    0    0    0
# [2,]    0    1    0    0    0
# [3,]    0    0    1    0    0
```

create.modMatrix

Create a moderator matrix used in OSMASEM

Description

It creates a moderator matrix used in OSMASEM.

Usage

```
create.modMatrix(RAM, output = c("A", "S"), mod)
```

Arguments

RAM	A RAM object including a list of matrices of the model returned from lavaan2RAM .
output	Whether the output is an "A" or "S" matrix.
mod	A string of moderator in the dataset.

Value

A character matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
## A multiple regression model
model <- "y ~ x1 + x2
          x1 ~~ 1*x1
          x2 ~~ 1*x2
          x1 ~~ x2"

## RAM specification
RAM <- lavaan2RAM(model, obs.variables=c("y", "x1", "x2"))

## Create a moderator matrix on A with "meanAge as the moderator.
A1 <- create.modMatrix(RAM=RAM, output="A", mod="meanAge")
A1

## Create a moderator matrix on S with "meanAge as the moderator.
S1 <- create.modMatrix(RAM=RAM, output="S", mod="meanAge")
S1
```

create.mxMatrix

Create a Vector into MxMatrix-class

Description

It converts a vector into MxMatrix-class via mxMatrix.

Usage

```
create.mxMatrix(
  x,
  type = c("Full", "Symm", "Diag", "Stand"),
  ncol = NA,
  nrow = NA,
  as.mxMatrix = TRUE,
  byrow = FALSE,
  ...
)
```

Arguments

x	A character or numeric vector
type	Matrix type similar to those listed in mxMatrix
ncol	The number of columns. It is necessary when type="Full". It is ignored and determined by the length of x for the other types of matrices.

nrow	The number of rows. It is necessary when type="Full". It is ignored and determined by the length of x for the other types of matrices.
as.mxMatrix	Logical. If it is TRUE, the output is a matrix of MxMatrix-class. If it is FALSE, it is a numeric matrix.
byrow	Logical. If FALSE (the default) the matrix is filled by columns, otherwise the matrix is filled by rows.
...	Further arguments to be passed to <code>mxMatrix</code> . Please note that type, nrow, ncol, values, free and labels will be created automatically. Thus, these arguments except labels should be avoided in ...

Details

If there are non-numeric values in x, they are treated as the labels of the free parameters. If an "*" is present, the numeric value on the left-hand side will be treated as the starting value for a free parameter or a fixed value for a fixed parameter. If it is a matrix of numeric values, there are no free parameters in the output matrix. nrow and ncol will be calculated from the length of x unless type="Full" is specified.

Value

A `MxMatrix-class` object with the same dimensions as x

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

`mxMatrix`, `create.mxMatrix`, `create.Fmatrix`

Examples

```
## a and b are free parameters with starting values and labels
(a1 <- c(1:4, "5*a", 6, "7*b", 8, 9))

(mat1 <- create.mxMatrix(a1, ncol=3, nrow=3, name="mat1"))

## Arrange the elements by row
(mat2 <- create.mxMatrix(a1, ncol=3, nrow=3, as.mxMatrix=FALSE, byrow=TRUE))

(a3 <- c(1:3, "4*f4", "5*f5", "6*f6"))

(mat3 <- create.mxMatrix(a3, type="Symm", name="mat3"))

## Create character matrix
(mat4 <- create.mxMatrix(a3, type="Symm", as.mxMatrix=FALSE))

## Arrange the elements by row
(mat5 <- create.mxMatrix(a3, type="Symm", as.mxMatrix=FALSE, byrow=TRUE))
```

```
(mat6 <- create.mxMatrix(a3, type="Diag", lbound=6:1, name="mat6"))
```

create.Tau2	<i>Create a variance component of the heterogeneity of the random effects</i>
-------------	---

Description

It creates variance component of the heterogeneity of the random effects by decomposing the variance component into matrices of correlation and standard deviations.

Usage

```
create.Tau2(
  RAM,
  no.var,
  Tau1.labels = seq(no.var),
  RE.type = c("Diag", "CSymm", "HCSymm", "Symm", "Zero", "User"),
  level = c("single", "between", "within"),
  RE.User = NULL,
  Transform = c("expLog", "sqSD"),
  RE.startvalues = 0.05
)
```

Arguments

RAM	The RAM model for testing. no.var is calculated from it.
no.var	If RAM is missing, the user has to specify the no.var argument. It represents the no.var by no.var of the random effects).
Tau1.labels	Parameter labels in Tau1. The default is Tau1_1, Tau1_2, etc.
RE.type	Either "Diag", "CSymm", "HCSymm", "Symm", "Zero" or "User". If it is "Diag" (the default if missing), a diagonal matrix is used for the random effects meaning that the random effects are independent. "CSymm" and "HCSymm" represent compound symmetry and heteroscedastic compound symmetry, respectively. If it is "Symm", a symmetric matrix is used for the random effects on the covariances among the correlation (or covariance) vectors. If it is "Zero", a zero matrix is assumed on the variance component of the random effects. If it is "User", users have to specify the RE.User argument.
level	whether it is for single-level, between-, or within-level analyses. The only difference are the names of the matrices.
RE.User	It represents the no.var by no.var symmetric matrix of TRUE or FALSE for the variance component. If the elements are FALSE, they are fixed at 0.
Transform	Either "expLog" or "sqSD". If it is "expLog", the variances are estimated by applying a log and exp transformation. If it is "sqSD", the variances are estimated by applying a square on the SD. The transformation may improve the estimation when the heterogeneity is small or close to zero.

RE.startvalues Starting values for the variances.

Value

A list of `MxMatrix`-class. The variance component is computed in `Tau2`.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[osmasem](#), [create.V](#), [create.vechsR](#)

Examples

```
T0 <- create.Tau2(no.var=4, RE.type="Diag", Transform="expLog", RE.startvalues=0.05)
T0

T1 <- create.Tau2(no.var=4, Tau1.labels=c("a", "b", "c", "d"))
T1
```

create.V	Create a V-known matrix
----------	-------------------------

Description

It creates a V-known matrix of the sampling covariance matrix using definition variables.

Usage

```
create.V(x, type = c("Symm", "Diag", "Full"), as.mxMatrix = TRUE)
```

Arguments

x	A character vector of variable names of the sampling covariance matrix.
type	Either "Symm", "Diag" or "Full". Suppose the number of variables is p , the numbers of variable names for "Symm", "Diag", and "Full" are $p(p-1)/2$, p , and $p * p$, respectively. The elements are arranged in a column major.
as.mxMatrix	Logical. Whether to convert the output into <code>MxMatrix</code> -class.

Value

A list of `MxMatrix`-class. The V-known sampling covariance matrix is computed in `V`.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[osmasem](#), [create.Tau2](#), [create.vechsR](#)

Examples

```
my.df <- Cor2DataFrame(Nohe15A1)

## Create known sampling variance covariance matrix
V0 <- create.V(my.df$vlabels)
V0
```

create.vechsR	<i>Create a model implied correlation matrix with implicit diagonal constraints</i>
---------------	---

Description

It creates implicit diagonal constraints on the model implied correlation matrix by treating the error variances as functions of other parameters.

Usage

```
create.vechsR(
  A0,
  S0,
  F0 = NULL,
  Ax = NULL,
  Sx = NULL,
  A.lbound = NULL,
  A.ubound = NULL
)
```

Arguments

A0	A Amatrix, which will be converted into MxMatrix-class via as.mxMatrix.
S0	A Smatrix, which will be converted into MxMatrix-class via as.mxMatrix.
F0	A Fmatrix, which will be converted into MxMatrix-class via as.mxMatrix.
Ax	A Amatrix of a list of Amatrix with definition variables as the moderators of the Amatrix.

Sx	A Smatrix of a list of Smatrix with definition variables as the moderators of the Smatrix.
A.lbound	A matrix of lower bound of the Amatrix. If a scalar is given, the lbound matrix will be filled with this scalar.
A.ubound	A matrix of upper bound of the Amatrix. If a scalar is given, the ubound matrix will be filled with this scalar.

Value

A list of MxMatrix-class. The model implied correlation matrix is computed in impliedR and vechsR.

Note

Since A0 are the intercepts and Ax are the regression coefficients, the parameters in Ax must be a subset of those in A0.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[osmasem](#), [create.Tau2](#), [create.V](#)

Examples

```
## Proposed model
model1 <- 'W2 ~ w2w*W1 + s2w*S1
          S2 ~ w2s*W1 + s2s*S1
          W1 ~~ w1WITHs1*S1
          W2 ~~ w2WITHs2*S2
          W1 ~~ 1*W1
          S1 ~~ 1*S1
          W2 ~~ Errw2*W2
          S2 ~~ Errs2*S2'

## Convert into RAM
RAM1 <- lavaan2RAM(model1, obs.variables=c("W1", "S1", "W2", "S2"))

## No moderator
M0 <- create.vechsR(A0=RAM1$A, S0=RAM1$S, F0=NULL, Ax=NULL, Sx=NULL)

## Lag (definition variable) as a moderator on the paths in the Amatrix
Ax <- matrix(c(0,0,0,0,
              0,0,0,0,
              "0*data.Lag","0*data.Lag",0,0,
              "0*data.Lag","0*data.Lag",0,0),
            nrow=4, ncol=4, byrow=TRUE)
```

```

M1 <- create.vechsR(A0=RAM1$A, S0=RAM1$S, F0=NULL, Ax=Ax, Sx=NULL)

## Lag (definition variable) as a moderator on the correlation in the Smatrix
Sx <- matrix(c(0,"0*data.Lag",0,0,
               "0*data.Lag",0,0,0,
               0,0,0,"0*data.Lag",
               0,0,"0*data.Lag",0),
             nrow=4, ncol=4, byrow=TRUE)

M2 <- create.vechsR(A0=RAM1$A, S0=RAM1$S, F0=NULL, Ax=NULL, Sx=Sx)

```

Diag

Matrix Diagonals

Description

Extract or replace the diagonal of a matrix, or construct a diagonal matrix with the same behaviors as diag prior to R-3.0.0.

Usage

```
Diag(x, ...)
```

```
Diag(x) <- value
```

Arguments

x	A matrix, vector or 1D array, or missing.
...	Optional dimensions (nrow and ncol) for the result when x is not a matrix.
value	Either a single value or a vector of length equal to that of the current diagonal. Should be of a mode which can be coerced to that of x.

Details

Starting from R-3.0.0, diag(x) returns a numeric matrix with NA in the diagonals when x is a character vector. Although this follows what the manual says, this breaks metaSEM. The Diag has the same functions as diag except that Diag(x) works for a character vector of x by returning a square matrix of character "0" with x as the diagonals.

Note

See <http://r.789695.n4.nabble.com/Behaviors-of-diag-with-character-vector-in-R-3-0-0-td4663735.html> for the discussion.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also[diag](#)**Examples**

```
v <- c("a", "b")
Diag(v)
```

Digman97

*Factor Correlation Matrices of Big Five Model from Digman (1997)***Description**

The data set includes fourteen studies of the factor correlation matrices of the Five-Factor Model of personality reported by Digman (1997).

Details

A list of data with the following structure:

data A list of 14 studies of correlation matrices. The variables are *Agreeableness* (A), *Conscientiousness* (C), *Emotional Stability* (ES), *Extraversion* (E) and *Intellect* (I)

n A vector of sample sizes

cluster Types of participants of the studies

Source

Digman, J.M. (1997). Higher-order factors of the Big Five. *Journal of Personality and Social Psychology*, **73**, 1246-1256.

References

Cheung, M. W.-L., & Chan, W. (2005). Classifying correlation matrices into relatively homogeneous subgroups: A cluster analytic approach. *Educational and Psychological Measurement*, **65**, 954-979.

Examples

```
Digman97

##### Fixed-effects TSSEM
fixed1 <- tssem1(Digman97$data, Digman97$n, method="FEM")
summary(fixed1)

## Two-factor CFA model in lavaan syntax
model <- "## Factor loadings"
```

```

      Alpha=~A+C+ES
      Beta=~E+I
      ## Factor correlation
      Alpha~~Beta"

## Display the model
plot(model)

RAM <- lavaan2RAM(model, obs.variables=c("A","C","ES","E","I"),
                  A.notation="on", S.notation="with")

RAM

## ## Equivalent RAM specification
## Phi <- matrix(c(1,"0.3*cor","0.3*cor",1), ncol=2, nrow=2)
## Psi <- Diag(c("0.2*e1","0.2*e2","0.2*e3","0.2*e4","0.2*e5"))
## S1 <- bdiagMat(list(Psi, Phi))
## dimnames(S1)[[1]] <- dimnames(S1)[[2]] <- c("A","C","ES","E","I","Alpha","Beta")
## S1
##
## Lambda <- matrix(c(".3*Alpha_A",".3*Alpha_C",".3*Alpha_ES",rep(0,5),
##                    ".3*Beta_E",".3*Beta_I"), ncol=2, nrow=5)
## A1 <- rbind( cbind(matrix(0,ncol=5,nrow=5), Lambda),
##              matrix(0, ncol=7, nrow=2) )
## dimnames(A1)[[1]] <- dimnames(A1)[[2]] <- c("A","C","ES","E","I","Alpha","Beta")
## A1
##
## F1 <- create.Fmatrix(c(1,1,1,1,1,0,0), as.mxMatrix=FALSE)

fixed2 <- tssem2(fixed1, RAM=RAM,
                 model.name="TSSEM2 Digman97")
summary(fixed2)

## Display the model with the parameter estimates
plot(fixed2)

#### Fixed-effects TSSEM with several clusters
#### Create a variable for different samples
#### Younger participants: Children and Adolescents
#### Older participants: others
cluster <- ifelse(Digman97$cluster %in% c("Children","Adolescents"),
                  yes="Younger participants", no="Older participants")

#### Show the cluster
cluster

## Example of Fixed-effects TSSEM with several clusters
fixed1.cluster <- tssem1(Digman97$data, Digman97$n, method="FEM",
                        cluster=cluster)
summary(fixed1.cluster)

fixed2.cluster <- tssem2(fixed1.cluster, RAM=RAM)
#### Please note that the estimates for the younger participants are problematic.
summary(fixed2.cluster)

```

```
## Setup two plots
layout(t(1:2))

## Plot the first group
plot(fixed2.cluster[[1]])
title("Younger participants")

## Plot the second group
plot(fixed2.cluster[[2]])
title("Older participants")

#### Random-effects TSSEM with random effects on the diagonals
random1 <- tssem1(Digman97$data, Digman97$n, method="REM",
                  RE.type="Diag")
summary(random1)

random2 <- tssem2(random1, RAM=RAM)
summary(random2)

## Display the model with the parameter estimates
plot(random2, color="green")
```

Gleser94

Two Datasets from Gleser and Olkin (1994)

Description

It includes two datasets in multiple-treatment studies and multiple-endpoint studies reported by Gleser and Olkin (1994).

Format

A list of two data frames.

MTS A data frame of multiple-treatment studies.

MES A data frame of multiple-endpoint studies.

Source

Gleser, L. J., & Olkin, I. (1994). Stochastically dependent effect sizes. In H. Cooper & L. V. Hedges (Eds.), *The handbook of research synthesis*. (pp. 339-355). New York: Russell Sage Foundation.

See Also

[smdMTS](#), [smdMES](#)

Examples

```

data(Gleser94)

#### Multiple-treatment studies
Gleser94$MTS

## Assuming homogeneity of variances
my.MTS <- t(apply(Gleser94$MTS, MARGIN=1,
  function(x)
    smdMTS(m=x[c("Mean.C", "Mean.E1", "Mean.E2", "Mean.E3", "Mean.E4", "Mean.E5")],
      v=x[c("SD.C", "SD.E1", "SD.E2", "SD.E3", "SD.E4", "SD.E5")]^2,
      n=x[c("N.C", "N.E1", "N.E2", "N.E3", "N.E4", "N.E5")],
      homogeneity="variance", list.output=FALSE)))

## Fixed-effects multivariate meta-analysis
fit.MTS <- meta(y=my.MTS[, 1:5],
  v=my.MTS[, 6:20],
  RE.constraints = diag(0, ncol=5, nrow=5),
  model.name="MTS")
summary(fit.MTS)

#### Multiple-endpoint studies
Gleser94$MES

## Calculate the sampling variances and covariance and amend into the data set
Gleser94$MES$Uncoached.V11 <- with(Gleser94$MES, SD.Uncoached.Math^2)
Gleser94$MES$Uncoached.V21 <- with(Gleser94$MES,
  SD.Uncoached.Math*Cor.Math.Verbal*SD.Uncoached.Verbal)
Gleser94$MES$Uncoached.V22 <- with(Gleser94$MES, SD.Uncoached.Verbal^2)

Gleser94$MES$Coached.V11 <- with(Gleser94$MES, SD.Coached.Math^2)
Gleser94$MES$Coached.V21 <- with(Gleser94$MES,
  SD.Coached.Math*Cor.Math.Verbal*SD.Coached.Verbal)
Gleser94$MES$Coached.V22 <- with(Gleser94$MES, SD.Coached.Verbal^2)

## Assuming homogeneity of covariance matrices
my.MES <- t(apply(Gleser94$MES, MARGIN=1,
  function(x)
    smdMES(m1=x[c("Mean.Uncoached.Math", "Mean.Uncoached.Verbal")],
      m2=x[c("Mean.Coached.Math", "Mean.Coached.Verbal")],
      V1=vec2symMat(x[c("Uncoached.V11", "Uncoached.V21", "Uncoached.V22")]),
      V2=vec2symMat(x[c("Coached.V11", "Coached.V21", "Coached.V22")]),
      n1=x["N.Uncoached"],
      n2=x["N.Coached"],
      homogeneity="covariance", list.output=FALSE)))

## Fixed-effects multivariate meta-analysis
fit.MES <- meta(y=my.MES[, 1:2],
  v=my.MES[, 3:5],
  RE.constraints = diag(0, ncol=2, nrow=2),
  model.name="MES")

```

```
summary(fit.MES)
```

Gnambs18

Correlation Matrices from Gnambs, Scharl, and Schroeders (2018)

Description

The data set includes 113 correlation matrices on the Rosenberg Self-Esteem Scale reported by Gnambs, Scharl, and Schroeders (2018). Thirty-six studies were based on the reported correlation matrices (CorMat=1) whereas the correlation matrices of the other 77 studies were calculated from the reported factor loadings.

Details

A list of data with the following structure:

data A list of 113 correlation matrices. The variable names are from *I1* to *I10*.

n A vector of sample sizes.

Year The year of publications.

Country The country of studies conducted.

Language The language used in the studies.

Publication Whether the studies were published (1) or unpublished (0).

MeanAge Mean age of the participants.

FemaleProp Proportion of the female participants.

Individualism Individualism score of the country.

CorMat Whether the correlation matrices are obtained from the original studies (1) or reproduced from the factor loadings (0).

Source

Gnambs, T., Scharl, A., & Schroeders, U. (2018). The structure of the Rosenberg Self-Esteem Scale. *Zeitschrift Fur Psychologie*, **226**(1), 14-29. <https://doi.org/10.1027/2151-2604/a000317>

HedgesOlkin85

*Effects of Open Education Reported by Hedges and Olkin (1985)***Description**

Effects of open education on attitude toward school and on reading achievement reported by Hedges and Olkin (1985).

Details

The variables are:

study Study number

d_att Standardized mean difference on *attitude*

d_ach Standardized mean difference on *achievement*

var_att Sampling variance of the effect size of *attitude*

cov_att_ach Sampling covariance between the effect sizes

var_ach Sampling variance of the effect size of *achievement*

Source

Hedges, L. V., & Olkin, I. (1985). *Statistical methods for meta-analysis*. Orlando, FL: Academic Press.

References

Cheung, M. W.-L. (2010). Fixed-effects meta-analyses as multiple-group structural equation models. *Structural Equation Modeling*, **17**, 481-509.

Examples

```
data(HedgesOlkin85)

## Fixed-effects meta-analysis
summary( meta(y=cbind(d_att, d_ach),
                    v=cbind(var_att, cov_att_ach, var_ach),
                    data=HedgesOlkin85,
                    RE.constraints=matrix(0, nrow=2, ncol=2)) )
```

homoStat

*Test the Homogeneity of Effect Sizes***Description**

It tests the homogeneity of univariate and multivariate effect sizes.

Usage

```
homoStat(y, v)
```

Arguments

y	A vector of effect size for univariate meta-analysis or a $k \times p$ matrix of effect sizes for multivariate meta-analysis where k is the number of studies and p is the number of effect sizes.
v	A vector of the sampling variance of the effect size for univariate meta-analysis or a $k \times p^*$ matrix of the sampling covariance matrix of the effect sizes for multivariate meta-analysis where $p^* = p(p + 1)/2$. It is arranged by column major as used by vech . It is assumed that there is no missing value in v if y is complete. If there are missing values in v due to the missingness on y , the missing values in v will be removed automatically.

Value

A list of	
Q	Q statistic on the null hypothesis of homogeneity of effect sizes. It has an approximate chi-square distribution under the null hypothesis.
Q.df	Degrees of freedom of the Q statistic
pval	p-value on the test of homogeneity of effect sizes

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Becker, B. J. (1992). Using results from replicated studies to estimate linear models. *Journal of Educational Statistics*, **17**, 341-362.

Cheung, M. W.-L. (2010). Fixed-effects meta-analyses as multiple-group structural equation models. *Structural Equation Modeling*, **17**, 481-509.

Cochran, W. G. (1954). The combination of estimates from different experiments. *Biometrics*, **10**, 101-129.

See Also[meta](#)**Examples**

```
with( Hox02, homoStat(yi, vi) )

with( HedgesOlkin85, homoStat(y=cbind(d_att, d_ach),
  v=cbind(var_att, cov_att_ach, var_ach)) )
```

Hox02

*Simulated Effect Sizes Reported by Hox (2002)***Description**

Twenty simulated studies on standardized mean difference and one continuous study characteristic reported by Hox (2002).

Details

The variables are:

study Study number

yi Effect size (standardized mean difference)

vi Sampling variance of the effect size

weeks Duration of the experimental intervention in terms of weeks

Source

Hox, J. J. (2002). *Multilevel analysis: Techniques and applications*. Mahwah, N.J.: Lawrence Erlbaum Associates.

References

Cheung, M. W.-L. (2008). A model for integrating fixed-, random-, and mixed-effects meta-analyses into structural equation modeling. *Psychological Methods*, **13**, 182-202.

Examples

```
data(Hox02)

#### ML estimation method
## Random-effects meta-analysis
summary( meta(y=yi, v=vi, data=Hox02, I2=c("I2q", "I2hm"), intervals.type="LB") )

## Fixed-effects meta-analysis
```

```

summary( meta(y=yi, v=vi, data=Hox02, RE.constraints=0,
             model.name="Fixed effects model") )

## Mixed-effects meta-analysis with "weeks" as a predictor
## Request likelihood-based CI
summary( meta(y=yi, v=vi, x=weeks, data=Hox02, intervals.type="LB",
             model.name="Mixed effects meta analysis with LB CI") )

#### REML estimation method
## Random-effects meta-analysis with REML
summary( VarComp <- reml(y=yi, v=vi, data=Hox02) )

## Extract the variance component
VarComp_REML <- matrix( coef(VarComp), ncol=1, nrow=1 )

## Meta-analysis by treating the variance component as fixed
summary( meta(y=yi, v=vi, data=Hox02, RE.constraints=VarComp_REML) )

## Mixed-effects meta-analysis with "weeks" as a predictor
## Request Wald CI
summary( reml(y=yi, v=vi, x=weeks, intervals.type="z",
             data=Hox02, model.name="REML with LB CI") )

```

 Hunter83

Fourteen Studies of Correlation Matrices reported by Hunter (1983)

Description

This dataset includes fourteen studies of Correlation Matrices reported by Hunter (1983)

Details

A list of data with the following structure:

data A list of 14 studies of correlation matrices. The variables are *Ability*, *Job knowledge*, *Work sample* and *Supervisor rating*

n A vector of sample sizes

Source

Hunter, J. E. (1983). A causal analysis of cognitive ability, job knowledge, job performance, and supervisor ratings. In F. Landy, S. Zedeck, & J. Cleveland (Eds.), *Performance Measurement and Theory* (pp. 257-266). Hillsdale, NJ: Erlbaum.

Examples

```

data(Hunter83)

#### Fixed-effects model
## First stage analysis
fixed1 <- tssem1(Hunter83$data, Hunter83$n, method="FEM",
                 model.name="TSSEM1 fixed effects model")
summary(fixed1)

#### Second stage analysis
## Model without direct effect from Ability to Supervisor
## A1 <- create.mxMatrix(c(0,"0.1*A2J","0.1*A2W",0,0,0,"0.1*J2W","0.1*J2S",
##                        0,0,0,"0.1*W2S",0,0,0,0),
##                        type="Full", ncol=4, nrow=4, as.mxMatrix=FALSE)

## ## This step is not necessary but it is useful for inspecting the model.
## dimnames(A1)[[1]] <- dimnames(A1)[[2]] <- c("Ability","Job","Work","Supervisor")
## A1

## S1 <- create.mxMatrix(c(1,"0.1*Var_e_J", "0.1*Var_e_W", "0.1*Var_e_S"),
##                        type="Diag", as.mxMatrix=FALSE)
## dimnames(S1)[[1]] <- dimnames(S1)[[2]] <- c("Ability","Job","Work","Supervisor")
## S1

#####
## Model specification in lavaan model syntax
## The "ind" effect can be defined within the syntax
model1 <- "## Regression paths
          Job_knowledge ~ A2J*Ability
          Work_sample ~ A2W*Ability + J2W*Job_knowledge
          Supervisor ~ J2S*Job_knowledge + W2S*Work_sample

          ## Fix the variance of Ability at 1
          Ability ~~ 1*Ability

          ## Label the error variances of the dependent variables
          Job_knowledge ~~ VarE_J*Job_knowledge
          Work_sample ~~ VarE_W*Work_sample
          Supervisor ~~ VarE_S*Supervisor

          ## Define an indirect effect
          ind := A2J*J2S+A2J*J2W*W2S+A2W*W2S"

## Display the model
plot(model1, layout="spring", sizeMan=10)

RAM1 <- lavaan2RAM(model1, obs.variables=c("Ability","Job_knowledge",
                                           "Work_sample","Supervisor"))
RAM1

#####

```

```

fixed2 <- tssem2(fixed1, RAM=RAM1, intervals.type="z",
                 diag.constraints=FALSE,
                 model.name="TSSEM2 fixed effects model")
summary(fixed2)

## Display the model with the parameter estimates
plot(fixed2, layout="spring")

## Coefficients
coef(fixed2)

## VCOV based on parametric bootstrap
vcov(fixed2)

#### Random-effects model with diagonal elements only
## First stage analysis
random1 <- tssem1(Hunter83$data, Hunter83$n, method="REM", RE.type="Diag",
                  acov="weighted", model.name="TSSEM1 random effects model")
summary(random1)

model2 <- "## Regression paths
          Job_knowledge ~ A2J*Ability
          Work_sample ~ A2W*Ability + J2W*Job_knowledge
          Supervisor ~ J2S*Job_knowledge + W2S*Work_sample

          ## Fix the variance of Ability at 1
          Ability ~~ 1*Ability

          ## Label the error variances of the dependent variables
          Job_knowledge ~~ VarE_J*Job_knowledge
          Work_sample ~~ VarE_W*Work_sample
          Supervisor ~~ VarE_S*Supervisor"

RAM2 <- lavaan2RAM(model2, obs.variables=c("Ability", "Job_knowledge",
                                           "Work_sample", "Supervisor"))
RAM2

## Second stage analysis
## Model without direct effect from Ability to Supervisor

## The "ind" effect is defined in tssem2().
random2 <- tssem2(random1, RAM=RAM2, intervals.type="LB",
                  diag.constraints=FALSE,
                  mx.algebras=
                    list(ind=mxAlgebra(A2J*J2S+A2J*J2W*W2S+A2W*W2S, name="ind")),
                  model.name="TSSEM2 random effects model")

summary(random2)

## Display the model with the parameter estimates
plot(random2, layout="spring")

```

impliedR	<i>Create or Generate the Model Implied Correlation or Covariance Matrices</i>
----------	--

Description

It creates or generates the model implied correlation or covariance matrices based on the RAM model specification.

Usage

```
impliedR(RAM, Amatrix, Smatrix, Fmatrix, Mmatrix, corr = TRUE, labels, ...)

rimpliedR(
  RAM,
  Amatrix,
  Smatrix,
  Fmatrix,
  AmatrixSD,
  SmatrixSD,
  k = 1,
  corr = TRUE,
  nonPD.pop = c("replace", "nearPD", "accept")
)
```

Arguments

RAM	A RAM object including a list of matrices of the model returned from lavaan2RAM .
Amatrix	If RAM is not specified, an Amatrix is required. An asymmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Smatrix	If RAM is not specified, an Smatrix is required. A symmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Fmatrix	A filter matrix in the RAM specification with MxMatrix-class . If it is missing, an identity matrix with the same dimensions of Smatrix will be created, which means that all variables are observed. If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function. It is not required when there is no latent variable.
Mmatrix	An optional matrix of the mean vector. It is assumed zeros if missing.
corr	Logical. The output is either the model implied correlation matrix or the covariance matrix.
labels	A character vector of the observed and latent variables with the same dimensions as that in the Amatrix and Smatrix.
...	Not used.

<code>AmatrixSD</code>	Standard deviations (SD) of the elements in the <code>Amatrix</code> . If it is missing, a matrix of zero is created.
<code>SmatrixSD</code>	Standard deviations (SD) of the elements in the <code>Smatrix</code> . If it is missing, a matrix of zero is created.
<code>k</code>	Number of studies.
<code>nonPD.pop</code>	If it is <code>replace</code> , generated non-positive definite matrices are replaced by generated new ones which are positive definite. If it is <code>nearPD</code> , they are replaced by nearly positive definite matrices by calling <code>Matrix::nearPD()</code> . If it is <code>accept</code> , they are accepted.

Details

This function can be used to generate the model implied correlation matrix for the standardized parameters with the `corr=TRUE` argument. Suppose we want to calculate the population correlation matrix for a mediation model with `x`, `m`, and `y`. We only need to specify the population path coefficients among `x`, `m`, and `y` in the `Amatrix`. We do not need to specify the population error variances of `m` and `y`. We treat the error variances as unknown parameters by giving them starting values in the `Smatrix` matrix. When the covariance matrix is requested by specifying `corr=FALSE`, it simply calculates the population model covariance matrix by treating the values in `Smatrix` as the population values.

Value

A list of RAM matrices, the model implied correlation or covariance matrix of the observed variables (`SigmaObs`), of both observed and latent variables (`SigmaAll`), the minimum fit (`minFit`) which should be zero, and the status code of the optimization (`status`) which should also be zero when the optimization is fine. The last object is `mx.fit` which is the output after running the model. It can be used in the diagnosis.

Note

It is important to ensure that all the population values in `Amatrix` must be set as fixed parameters; otherwise, these values may be altered with the `corr=TRUE` argument. When there is an error or warning message about the status code, there is a high chance that some of the values in `Amatrix` are incorrectly set as free parameters.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
set.seed(100)

## A one-factor CFA model
model <- "f =~ 0.3*x1 + 0.4*x2 + 0.5*x3
         f =~ 1*f"

RAM <- lavaan2RAM(model)
```

```

impliedR(RAM, corr=TRUE)

## A simple mediation model
## All are population parameters in the A matrix
A1 <- matrix(c(0, 0, 0,
               0.3, 0, 0,
               0.2, 0.4, 0), nrow=3, ncol=3, byrow=TRUE,
             dimnames=list(c("x", "m", "y"), c("x", "m", "y")))
A1

## Variance of x is fixed at 1 while the other variances are free.
S1 <- matrix(c(1, 0, 0,
               0, "0.1*ErrVarM", 0,
               0, 0, "0.1*ErrVarY"), nrow=3, ncol=3,
             dimnames=list(c("x", "m", "y"), c("x", "m", "y")))
S1

impliedR(Amatrix=A1, Smatrix=S1)

## SD of A1
A1SD <- matrix(c(0, 0, 0,
                 0.1, 0, 0,
                 0.1, 0.1, 0), nrow=3, ncol=3, byrow=TRUE,
               dimnames=list(c("x", "m", "y"), c("x", "m", "y")))
A1SD

rimpliedR(Amatrix=A1, Smatrix=S1, AmatrixSD=A1SD, k=2)

## A CFA model
A2 <- matrix(c(0, 0, 0, 0.3,
               0, 0, 0, 0.4,
               0, 0, 0, 0.5,
               0, 0, 0, 0), nrow=4, ncol=4, byrow=TRUE,
             dimnames=list(c("x1", "x2", "x3", "f"),
                           c("x1", "x2", "x3", "f")))
A2

## Variance of f is fixed at 1 while the other variances are free.
S2 <- matrix(c("0.7*Err1", 0, 0, 0,
               0, "0.7*Err2", 0, 0,
               0, 0, "0.7*Err3", 0,
               0, 0, 0, 1), nrow=4, ncol=4,
             dimnames=list(c("x1", "x2", "x3", "f"), c("x1", "x2", "x3", "f")))
S2

F2 <- create.Fmatrix(c(1,1,1,0), as.mxMatrix=FALSE)
F2

## Model implied correlation matrix
impliedR(Amatrix=A2, Smatrix=S2, Fmatrix=F2, corr=TRUE)

## Model implied covariance matrix

```

```

impliedR(Amatrix=A2, Smatrix=S2, Fmatrix=F2, corr=FALSE)

## SD of A2
A2SD <- matrix(c(0, 0, 0, 0.1,
                 0, 0, 0, 0.1,
                 0, 0, 0, 0.1,
                 0, 0, 0, 0), nrow=4, ncol=4, byrow=TRUE,
               dimnames=list(c("x1", "x2", "x3", "f"),
                             c("x1", "x2", "x3", "f"))))
A2SD

## SD of S2: correlated between x1 and x2
S2SD <- matrix(c(0, 0.1, 0, 0,
                 0.1, 0, 0, 0,
                 0, 0, 0, 0.1,
                 0, 0, 0, 0), nrow=4, ncol=4, byrow=TRUE,
               dimnames=list(c("x1", "x2", "x3", "f"),
                             c("x1", "x2", "x3", "f"))))
S2SD

rimpliedR(Amatrix=A2, Smatrix=S2, Fmatrix=F2, AmatrixSD=A2SD,
          SmatrixSD=S2SD, k=2)

```

indirectEffect	<i>Estimate the asymptotic covariance matrix of standardized or unstandardized indirect and direct effects</i>
----------------	--

Description

It estimates the standardized or unstandardized indirect and direct effects and their asymptotic sampling covariance matrix.

Usage

```
indirectEffect(x, n, standardized = TRUE, direct.effect = TRUE, run = TRUE)
```

Arguments

x	A 3x3 correlation/covariance matrix or a list of correlation/covariance matrices. Variables are arranged as the dependent variable (y), mediator (m) and independent variable (x)
n	Sample size or a vector of sample sizes
standardized	Logical. Whether the indirect effect is standardized.
direct.effect	Logical. Whether the direct effect is estimated. If it is FALSE, the direct effect is fixed at zero.
run	Logical. If FALSE, only return the mx model without running the analysis.

Details

Cheung (2009) estimated the standardized indirect effect and its standard error with non-linear constraints. Since OpenMx does not generate standard errors when there are non-linear constraints, Kwan and Chan's (2011) approach is used in this function. Delta method is used to calculate the asymptotic covariance matrix.

Value

A vector (or a matrix if the input is a list of matrices) of (standardized) indirect effect, standardized direct effect, and their asymptotic sampling covariance matrices

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2009). Comparison of methods for constructing confidence intervals of standardized indirect effects. *Behavior Research Methods*, 41, 425-438.
- Kwan, J., & Chan, W. (2011). Comparing standardized coefficients in structural equation modeling: a model reparameterization approach. *Behavior Research Methods*, 43, 730-745.

Examples

```
## A correlation matrix as input
x <- matrix(c(1, 0.4, 0.2, 0.4, 1, 0.3, 0.2, 0.3, 1), ncol=3)
dimnames(x) <- list( c("y", "m", "x"), c("y", "m", "x") )
indirectEffect(x, n=300)

## A list of correlation matrices
indirectEffect( list(x, x), n=c(300,500), standardized=FALSE )
```

is.pd

Test Positive Definiteness of a List of Square Matrices

Description

It tests the positive definiteness of a square matrix or a list of square matrices. It returns TRUE if the matrix is positive definite. It returns FALSE if the matrix is either non-positive definite or not symmetric. Variables with NA in the diagonals will be removed before testing. It returns NA when there are missing correlations even after deleting the missing variables.

Usage

```
is.pd(x, check.aCov = FALSE, cor.analysis = TRUE, tol = 1e-06)
```

Arguments

<code>x</code>	A square matrix or a list of square matrices
<code>check.aCov</code>	If it is TRUE, it mirrors the checking in asyCov .
<code>cor.analysis</code>	Whether the input matrix is a correlation or a covariance matrix. It is ignored when <code>check.aCov=FALSE</code> .
<code>tol</code>	Tolerance (relative to largest variance) for numerical lack of positive-definiteness in <code>x</code> . It is adopted from mvrnorm .

Value

If the input is a matrix, it returns TRUE, FALSE or NA. If the input is a list of matrices, it returns a list of TRUE, FALSE or NA.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
A <- diag(1,3)
is.pd(A)
# TRUE

B <- matrix(c(1,2,2,1), ncol=2)
is.pd(B)
# FALSE

is.pd(list(A, B))
# TRUE FALSE

C <- A
C[2,1] <- C[1,2] <- NA
is.pd(C)
# NA
```

Description

Thirty-two covariance matrices on work-related attitudes were extracted from the International Social Survey Programme 2005: Work Orientation III (ISSP, 2005). Seven variables were selected for demonstration purposes. They were grouped into three constructs: *Importance of Job Prospects* measured by job security (JP1), high income (JP2), and opportunity for advancement (JP3); *Importance of Job Autonomy* measured by work independently (JA1) and decide time of work (JA2); and *Importance of Contributions to Society* measured by help other people (CS1) and a job useful to society (CS2).


```

## Stage 2 analysis
rand2 <- tssem2(rand1, RAM=ram1)
summary(rand2)

plot(rand2)

#### OSMASEM with covariance matrices
## Create a data frame for the OSMASEM
df <- Cor2DataFrame(issp05$data, n=issp05$n, Means=issp05$means,
                    cor.analysis=FALSE)

## Standardize idv
idv <- scale(issp05$idv)

## Replace missing values with mean
idv[is.na(idv)] <- mean(idv, na.rm=TRUE)
df$data$idv <- idv

## No moderator
fit1 <- osmasem2(model.name="No_moderator", RAM=ram1, data=df,
                 cor.analysis=FALSE, mean.analysis=FALSE)
summary(fit1, fitIndices = TRUE)

## Proposed model with idv as a moderator
model2 <- "JP =~ a*JP1 + b*JP2 + c*JP3
          JA =~ d*JA1 + e*JA2
          CS =~ f*CS1 + g*CS2
          JP ~~ JA + CS
          JA ~~ CS
          a == a0 + a1*data.idv
          b == b0 + b1*data.idv
          c == c0 + c1*data.idv
          d == d0 + d1*data.idv
          e == e0 + e1*data.idv
          f == f0 + f1*data.idv
          g == g0 + g1*data.idv"

ram2 <- lavaan2RAM(model2, obs.variables=c("JP1", "JP2", "JP3", "JA1", "JA2",
                                           "CS1", "CS2"))

fit2 <- osmasem2(RAM=ram2, data=df, cor.analysis=FALSE, mean.analysis=FALSE,
                 replace.constraints = TRUE)
summary(fit2)

## Compare fit1 and fit2
anova(fit2, fit1)

```

Description

Eleven covariance matrices on work-related attitudes were extracted from the Inter-University Consortium for Political and Social Research (1989). Nine variables were selected by Cheung and Chan (2005; 2009) for demonstration purposes. They were grouped into three constructs: *Job Prospects* measured by job security (JP1), income (JP2), and advancement opportunity (JP3); *Job Nature* measured by interesting job (JN1), independent work (JN2), help other people (JN3), and useful to society (JN4); and *Time Demand* measured by flexible working hours (TD1) and lots of leisure time (TD2).

Details

A list of data with the following structure:

data A list of 11 studies of covariance matrices

n A vector of sample sizes

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Source

Inter-University Consortium for Political and Social Research. (1989). *International Social Survey Program: Work orientation*. Ann Arbor, MI: Author.

References

Cheung, M. W.-L., & Chan, W. (2005). Meta-analytic structural equation modeling: A two-stage approach. *Psychological Methods*, **10**, 40-64.

Cheung, M. W.-L., & Chan, W. (2009). A two-stage approach to synthesizing covariance matrices in meta-analytic structural equation modeling. *Structural Equation Modeling*, **16**, 28-53.

See Also

[issp05](#)

Examples

```
data(issp89)

#### Analysis of correlation structure in Cheung and Chan (2005)
#### Fixed-effects model: Stage 1 analysis
cor1 <- tssem1(issp89$data, issp89$n, method="FEM", cor.analysis=TRUE)
summary(cor1)

## Three-factor CFA model in lavaan syntax
## f1: Job Prospects (JP1-JP3); f2: Job Nature (JN1-JN4); f3: Time Demand (TD1-TD2)
model <- "## Factor loadings
         f1 =~ f1JP1*JP1 + f1JP2*JP2 + f1JP3*JP3"
```

```

f2 =~ f2JN1*JN1 + f2JN2*JN2 + f2JN3*JN3 + f2JN4*JN4
f3 =~ f3TD1*TD1 + f3TD2*TD2
## Factor correlations
f1 ~~ corf2f1*f2
f1 ~~ corf3f1*f3
f2 ~~ corf3f2*f3"

RAM1 <- lavaan2RAM(model, obs.variables=c("JP1","JP2","JP3",
                                           "JN1","JN2","JN3","JN4",
                                           "TD1","TD2"))

RAM1

## ## Equivalent RAM specification using create.mxMatrix()
## Phi <- create.mxMatrix( c("0.3*corf2f1","0.3*corf3f1","0.3*corf3f2"),
##                          type="Stand", as.mxMatrix=FALSE )
## Psi <- create.mxMatrix( paste("0.2*e", 1:9, sep=""), type="Diag",
##                          as.mxMatrix=FALSE )
## S1 <- bdiagMat(list(Psi, Phi))
## S1 <- as.mxMatrix(S1)
##
## Lambda <- create.mxMatrix( c(".3*f1x1",".3*f1x2",".3*f1x3",rep(0,9),
##                              ".3*f2x4",".3*f2x5",".3*f2x6",".3*f2x7",
##                              rep(0,9),".3*f3x8",".3*f3x9"), type="Full",
##                              ncol=3, nrow=9, as.mxMatrix=FALSE )
## Zero1 <- matrix(0, nrow=9, ncol=9)
## Zero2 <- matrix(0, nrow=3, ncol=12)
## A1 <- rbind( cbind(Zero1, Lambda), Zero2 )
## A1 <- as.mxMatrix(A1)
##
## F1 <- create.Fmatrix(c(rep(1,9), rep(0,3)))

#### Fixed-effects model: Stage 2 analysis
cor2 <- tssem2(cor1, RAM=RAM1, intervals.type="LB")
summary(cor2)

## Display the model with the parameter estimates
plot(cor2, nDigits=1)

#### Analysis of covariance structure in Cheung and Chan (2009)
#### Fixed-effects model: Stage 1 analysis
cov1 <- tssem1(issp89$data, issp89$n, method="FEM", cor.analysis=FALSE)
summary(cov1)

#### Fixed-effects model: Stage 2 analysis
cov2 <- tssem2(cov1, RAM=RAM1)
summary(cov2)

## Display the model with the parameter estimates
plot(cov2, nDigits=1)

```

Jaramillo05

*Dataset from Jaramillo, Mulki and Marshall (2005)***Description**

A dataset of the relationship between organizational commitment (OC) and salesperson job performance (JP) from Jaramillo, Mulki & Marshall (2005).

Format

A data frame with 61 observations on the following 10 variables.

Author a character vector of study

Sample_size sample size of the study

Sales sample type; either "mixed", "nonsales" or "sales"

Country a character vector of country of study

IDV Hofstede's (1997) individualism index

OC_scale scale of OC; either "Porter or Mowday", "Meyer" or "other"

OC_alpha Coefficient alpha of organizational commitment

JP_alpha Coefficient alpha of job performance

r correlation between organizational commitment and job performance

r_v sampling variance of r

Citations Citations from Google Scholar as of 27 August 2024

Source

Jaramillo, F., Mulki, J. P., & Marshall, G. W. (2005). A meta-analysis of the relationship between organizational commitment and salesperson job performance: 25 years of research. *Journal of Business Research*, **58**(6), 705-714. doi:10.1016/j.jbusres.2003.10.004

Examples

```
## Research question 4.4.1
summary(meta(r, r_v, data=Jaramillo05))

## Research question 4.4.2
## Select cases with either "sales" or "nonsales"
Sales.df <- subset(Jaramillo05, Sales %in% c("sales", "nonsales"))

## Create a predictor with 1 and 0 when they are "sales" or "nonsales", respectively
predictor <- ifelse(Jaramillo05$Sales=="sales", yes=1, no=0)

## Mixed-effects meta-analysis
summary( meta(y = r, v = r_v, x = predictor, data = Jaramillo05) )
```

```
## Research question 4.4.3
summary(meta(r, r_v, x=IDV, data=Jaramillo05))
```

Kalaian96

Multivariate effect sizes reported by Kalaian and Raudenbush (1996)

Description

This data set includes 47 multivariate effect sizes reported by Kalaian and Raudenbush (1996, Table 1).

Details

A list of data with the following structure:

Study Study name

Year Year of publication

n_e Sample size of the experimental group

n_c Sample size of the control group

dSAT_V Standardized mean difference of the Scholastic Aptitude Test (SAT) on verbal

dSAT_M Standardized mean difference of SAT on math

var_V Sampling variance of dSAT_V

cov_VM Sampling covariance of dSAT_V and dSAT_M with a common correlation of 0.66

var_M Sampling variance of dSAT_M

Hr Hours of training

ETS Educational Testing Service

Study_type Either Randomized, Matched or Nonequivalent comparison

Home_work Home work

Source

Kalaian, H. A., & Raudenbush, S. W. (1996). A multivariate mixed linear model for meta-analysis. *Psychological Methods*, 1(3), 227-235. <https://doi.org/10.1037/1082-989X.1.3.227>

Examples

```
data(Kalaian96)
```

lavaan2RAM

*Convert lavaan models to RAM models***Description**

It converts models specified in lavaan model syntax to RAM models.

Usage

```
lavaan2RAM(
  model,
  obs.variables = NULL,
  A.notation = "ON",
  S.notation = "WITH",
  M.notation = "mean",
  A.start = 0.1,
  S.start = 0.5,
  M.start = 0,
  auto.var = TRUE,
  std.lv = TRUE,
  ngroups = 1,
  ...
)
```

Arguments

<code>model</code>	A character string of model using the lavaan model syntax.
<code>obs.variables</code>	A character vector of the observed variables. The observed variables in the RAM specification will follow the order specified in <code>obs.variables</code> . It is important to check whether the order of the observed variables matches the order in the dataset.
<code>A.notation</code>	A character string to be used in the A matrix if the labels are not included in the lavaan model. For example, the label will be "yONx" for regressing "y" on "x".
<code>S.notation</code>	A character string to be used in the S matrix if the labels are not included in the lavaan model. For example, the label will be "yWITHx" for the covariance between "y" with "x" and "yWITHy" for the (error) variance of "y".
<code>M.notation</code>	A character string to be used in the M matrix if the labels are not included in the lavaan model. For example, the label will be "ymean" for the mean of "y" if <code>M.notation="mean"</code> .
<code>A.start</code>	A numeric value of starting value for the A matrix when the starting values are not provided.
<code>S.start</code>	A numeric value of starting value for the S matrix when the starting values are not provided.
<code>M.start</code>	A numeric value of starting value for the M matrix when the starting values are not provided.

<code>auto.var</code>	Logical. If TRUE, the residual variances and the variances of exogenous latent variables are included in the model and set free. See model.syntax .
<code>std.lv</code>	Logical. If TRUE (the default), the metric of each latent variable is determined by fixing their variances to 1.0. If FALSE, the metric of each latent variable is determined by fixing the factor loading of the first indicator to 1.0. See model.syntax . <code>std.lv=TRUE</code> always fixes every latent variable's variance at 1, even overriding an explicit " <code>~~</code> " specification for it in model (e.g. " <code>f ~~ c(vf1,vf2)*f</code> " to free a factor's variance per group) with no error – the label is simply discarded, since the parameter is fixed rather than estimated. Pass <code>std.lv=FALSE</code> whenever model identifies a latent variable's scale by fixing one of its loadings instead (e.g. " <code>f =~ 1*y1</code> " or " <code>f =~ c(1,1)*y1</code> " for a multiple-group model), which is incompatible with also fixing its variance. <code>lavaan2RAM()</code> warns when this happens (any parameter given an explicit label in model that ends up fixed rather than free), but the warning is general – checking <code>std.lv</code> first is faster than working backward from it.
<code>ngroups</code>	Number. The number of groups in the model. See model.syntax . Parameters without an explicit label in model are free and distinct in each group by default (as in lavaan); use lavaan's <code>c(label, label)</code> syntax to constrain a parameter to be equal across groups. Has no effect (and is rejected with an error) on two-level (<code>level:</code>) models – see below.
<code>...</code>	Further arguments to be passed to model.syntax

Details

It uses the [model.syntax](#) to do the conversion. Experimental: functions of parameters (`':='` in lavaan) and constraints (`'=='`, `'>'`, and `'<'` in lavaan) will be converted to `mxAlgebra` and `mxConstraint` in OpenMx. As there are differences between lavaan and OpenMx, they may not work properly.

Value

A list of RAM specification with A, S, F, and M matrices. When `ngroups > 1`, a list of such lists (one per group) is returned instead, with class `"RAM_multigroup"`. When model uses lavaan's two-level syntax (`level: 1 / level: 2` blocks), a list with within and between elements (each themselves a list of A, S, F, and M matrices) is returned instead, with class `"RAM_twolevel"`. Level-1 (within) manifest variables default to a fixed-at-zero mean (the overall intercept lives at the between level by convention); level-2 (between) "shadow" variables backing a level-1 manifest variable default to a free mean, while genuine level-2-only latent factors default to fixed-at-zero, mirroring the usual latent-variable convention. Genuine level-2-only observed covariates (e.g. a real cluster-level covariate with its own data column, as opposed to a level-1 manifest variable's between-cluster component) are not yet supported and raise an error, rather than being silently treated as an ordinary latent variable with no connection to any data; neither is combining two-level with multiple groups (also an error).

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[ramModel](#), [Becker92](#), [Becker09](#), [Digman97](#), [Hunter83](#), [as.mxMatrix](#), [checkRAM](#)

Examples

```
## Regression model on correlation matrix
model1 <- "## y is modelled by x1, x2, and x3
          y ~ b1*x1 + b2*x2 + b3*x3
          ## Fix the independent variables at 1
          x1 ~~ 1*x1
          x2 ~~ 1*x2
          x3 ~~ 1*x3
          ## Declare the correlations among the independent variables
          x1 ~~ x2
          x1 ~~ x3
          x2 ~~ x3
          ## Constraint
          b3 == b1 + b2
          ## Function of parameters
          fn1 := b1*b2^b3"

## Compare the arrangements of variables with and without
## specifying the obs.variables arguments.
lavaan2RAM(model1, obs.variables=c("y", "x1", "x2", "x3"))

## Two-factor CFA model
model2 <- "f1 =~ x1 + x2 + x3
          f2 =~ x4 + x5 + x6
          ## Declare the correlation between f1 and f2
          ## and label it with cor_f1f2
          f1 ~~ cor_f1f2*f2"

lavaan2RAM(model2)

## Regression model with the mean structure
model3 <- "y ~ x
          ## Intercept of y
          y ~ 1
          ## Mean of x
          x ~ 1"

lavaan2RAM(model3)

## Multiple group regression model
## Different intercepts with a common slope
model4 <- "y ~ c(a1, a2)*1 + c(b, b)*x"

lavaan2RAM(model4, ngroups=2)

## Two-level random-intercept regression model
model5 <- "level: 1
          y ~ b1*x"
```

```

        y ~~ residW*y
level: 2
        y ~ 1
        y ~~ residB*y"

lavaan2RAM(model5)

```

list2matrix

*Convert a List of Symmetric Matrices into a Stacked Matrix***Description**

It converts a list of symmetric matrices into a stacked matrix. Dimensions of the symmetric matrices have to be the same. It tries to preserve the dimension names if possible. Dimension names will be created if there are no dimension names in the first symmetric matrix.

Usage

```
list2matrix(x, diag = FALSE)
```

Arguments

x A list of k $p \times p$ symmetric matrices.

diag Logical. If it is TRUE, [vech](#) is used to vectorize the (covariance) matrices. If it is FALSE, [vechs](#) is used to vectorize the (correlation) matrices.

Value

A $k \times p^*$ stacked matrix where $p^* = p(p-1)/2$ for `diag=FALSE` or $p^* = p(p+1)/2$ for `diag=TRUE`.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```

C1 <- matrix(c(1,0.5,0.4,0.5,1,0.2,0.4,0.2,1), ncol=3)
C2 <- matrix(c(1,0.4,NA,0.4,1,NA,NA,NA,NA), ncol=3)

## A list without dimension names
list2matrix(list(C1, C2))
#      x2_x1 x3_x1 x3_x2
# [1,]  0.5  0.4  0.2
# [2,]  0.4  NA  NA

dimnames(C1) <- list( c("x","y","z"), c("x","y","z") )
dimnames(C2) <- list( c("x","y","z"), c("x","y","z") )

```

```
## A list with dimension names
list2matrix(list(C1, C2))
#      y_x z_x z_y
# [1,] 0.5 0.4 0.2
# [2,] 0.4 NA  NA
```

Mak09	<i>Eight studies from Mak et al. (2009)</i>
-------	---

Description

Eight studies from Mak et al. (2009) were reported by Cheung et al. (2012).

Format

A data frame with 8 observations on the following 10 variables.

Study a character vector of study

type a character vector

AF.BP a numeric vector

Tot.BP a numeric vector

AF.non.BP a numeric vector

Tot.non.BP a numeric vector

yi a numeric vector

vi a numeric vector

age.mean a numeric vector

study.duration a numeric vector

Source

Mak, A., Cheung, M. W.-L., Ho, R. C. M., Cheak, A. A. C., & Lau, C. S. (2009). Bisphosphonate and atrial fibrillation: Bayesian meta-analyses of randomized controlled trials and observational studies. *BMC Musculoskeletal Disorders*, **10**(113). doi:10.1186/1471-2474-10-113 Available at <https://link.springer.com/article/10.1186/1471-2474-10-113>.

References

Cheung, M. W.-L., Ho, R. C. M., Lim, Y., & Mak, A. (2012). Conducting a meta-analysis: Basics and good practices. *International Journal of Rheumatic Diseases*, **15**(2), 129-135. doi: 10.1111/j.1756-185X.2012.01712.x

Examples

```
## Random-effects meta-analysis
( meta1 <- summary(meta(y=yi, v=vi, data=Mak09, I2=c("I2q", "I2hm")))) )

## Convert the estimates back into odds ratio
OR <- with(coef(meta1), exp(c(Estimate[1], lbound[1], ubound[1])))
names(OR) <- c("Estimate in OR", "lbound in OR", "ubound in OR")
OR

## Mixed-effects meta-analysis with mean age as a predictor
summary( meta(y=yi, v=vi, x=age.mean, data=Mak09) )
```

Mathieu15

Correlation Matrices from Mathieu et al. (2015)

Description

The data set includes a list of correlation matrices of panel studies between cohesion (C) and performance (P) in Mathieu et al. (2015, Table 1).

Details

A list of data with the following structure:

data A list of studies of correlation matrices. The variables are *C1*, *P1*, *C2*, and *P2*.

n A vector of sample sizes.

Year Year of publication.

Sample Sample characteristics.

Student Whether the samples are student or non-student based on Sample.

Source

Mathieu, J. E., Kukenberger, M. R., D’Innocenzo, L., & Reilly, G. (2015). Modeling reciprocal team cohesion-performance relationships, as impacted by shared leadership and members’ competence. *Journal of Applied Psychology*, **100**(3), 713-734. <https://doi.org/10.1037/a0038898>

Examples

```
# TSSEM
## Model 1: no constraint
## Stage 1 analysis
tssem1.fit <- tssem1(Mathieu15$data, Mathieu15$n)
summary(tssem1.fit)

## Proposed model in lavaan syntax
model1 <- 'C2 ~ c2c*C1 + p2c*P1'
```

```

P2 ~ c2p*C1 + p2p*P1
C1 ~~ c1withp1*P1
C1 ~~ 1*C1
P1 ~~ 1*P1
C2 ~~ c2withp2*P2'

## Convert the lavaan model to RAM specification
RAM1 <- lavaan2RAM(model1, obs.variables=c("C1", "P1", "C2", "P2"))
RAM1

## Stage 2 analysis
tssem1b.fit <- tssem2(tssem1.fit, RAM=RAM1)
summary(tssem1b.fit)

plot(tssem1b.fit, col="yellow", edge.label.position=0.58)

## Model 2: Equality constraints on the path coefficient
## Proposed model with equal effects time 1 to time 2
model2 <- 'C2 ~ same*C1 + diff*P1
P2 ~ diff*C1 + same*P1
C1 ~~ c1withp1*P1
C1 ~~ 1*C1
P1 ~~ 1*P1
C2 ~~ c2withp2*P2'

## Convert the lavaan model to RAM specification
RAM2 <- lavaan2RAM(model2, obs.variables=c("C1", "P1", "C2", "P2"))
RAM2

## Stage 2 analysis
tssem2b.fit <- tssem2(tssem1.fit, RAM=RAM2)
summary(tssem2b.fit)

## Compare the models with and without the constraints.
anova(tssem1b.fit, tssem2b.fit)

## Plot the model
plot(tssem2b.fit, col="yellow", edge.label.position=0.60)

## OSMASEM
my.df <- Cor2DataFrame(Mathieu15)

head(my.df$data)

## Model without any moderator
osmasem.fit1 <- osmasem(model.name="No moderator", RAM=RAM1, data=my.df)
summary(osmasem.fit1)

## Extract the heterogeneity variance-covariance matrix
diag(VarCorr(osmasem.fit1))

plot(osmasem.fit1, col="yellow", edge.label.position=0.6)

```

```
## Model with student sample as a moderator on the regression coefficients
A1 <- create.modMatrix(RAM1, output="A", "Student")
A1

## Model with a moderator
osmasem.fit2 <- osmasem(model.name="Student sample as a moderator", RAM=RAM1,
                      Ax=A1, data=my.df)
summary(osmasem.fit2)

## Compare the models with and without the moderator
anova(osmasem.fit2, osmasem.fit1)

## Get the R2 of the moderator
osmasemR2(osmasem.fit2, osmasem.fit1)
```

matrix2bdiag	<i>Convert a Matrix into a Block Diagonal Matrix</i>
--------------	--

Description

It converts a matrix into a block diagonal matrix.

Usage

```
matrix2bdiag(x, ...)
```

Arguments

x	A $k \times p$ matrix of numerics or characters.
...	Further arguments to be passed to vec2symMat

Details

Each row of x is converted into a symmetric matrix via [vec2symMat](#). Then the list of the symmetric matrices is converted into a block diagonal matrix via a function written by Scott Chasalow posted at <http://www.math.yorku.ca/Who/Faculty/Monette/pub/stmp/0827.html>.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[vec2symMat](#)

Examples

```
(m1 <- matrix(1:12, ncol=6, byrow=TRUE))
#      [,1] [,2] [,3] [,4] [,5] [,6]
# [1,]  1   2   3   4   5   6
# [2,]  7   8   9  10  11  12

matrix2bdiag(m1)
#      [,1] [,2] [,3] [,4] [,5] [,6]
# [1,]  1   2   3   0   0   0
# [2,]  2   4   5   0   0   0
# [3,]  3   5   6   0   0   0
# [4,]  0   0   0   7   8   9
# [5,]  0   0   0   8  10  11
# [6,]  0   0   0   9  11  12
```

meta	<i>Univariate and Multivariate Meta-Analysis with Maximum Likelihood Estimation</i>
------	---

Description

It conducts univariate and multivariate meta-analysis with maximum likelihood estimation method. Mixed-effects meta-analysis can be conducted by including study characteristics as predictors. Equality constraints on intercepts, regression coefficients, and variance components can be easily imposed by setting the same labels on the parameter estimates.

Usage

```
meta(
  y,
  v,
  x,
  data,
  intercept.constraints = NULL,
  coef.constraints = NULL,
  RE.constraints = NULL,
  RE.startvalues = 0.1,
  RE.lbound = 1e-10,
  intervals.type = c("z", "LB"),
  I2 = "I2q",
  R2 = TRUE,
  model.name = "Meta analysis with ML",
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)
```

```

metaFIML(
  y,
  v,
  x,
  av,
  data,
  intercept.constraints = NULL,
  coef.constraints = NULL,
  RE.constraints = NULL,
  RE.startvalues = 0.1,
  RE.lbound = 1e-10,
  intervals.type = c("z", "LB"),
  R2 = TRUE,
  model.name = "Meta analysis with FIML",
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)

```

Arguments

- | | |
|------------------------------------|---|
| <code>y</code> | A vector of effect size for univariate meta-analysis or a $k \times p$ matrix of effect sizes for multivariate meta-analysis where k is the number of studies and p is the number of effect sizes. |
| <code>v</code> | A vector of the sampling variance of the effect size for univariate meta-analysis or a $k \times p^*$ matrix of the sampling covariance matrix of the effect sizes for multivariate meta-analysis where $p^* = p(p + 1)/2$. It is arranged by column major as used by vech . |
| <code>x</code> | A predictor or a $k \times m$ matrix of predictors where m is the number of predictors. |
| <code>data</code> | An optional data frame containing the variables in the model. |
| <code>intercept.constraints</code> | A $1 \times p$ matrix specifying whether the intercepts of the effect sizes are fixed or free. If the input is not a matrix, the input is converted into a $1 \times p$ matrix with <code>t(as.matrix(intercept.constraints))</code> . The default is that the intercepts are free. When there is no predictor, these intercepts are the same as the pooled effect sizes. The format of this matrix follows as.mxMatrix . The intercepts can be constrained equally by using the same labels. |
| <code>coef.constraints</code> | A $p \times m$ matrix specifying how the predictors predict the effect sizes. If the input is not a matrix, it is converted into a matrix by <code>as.matrix()</code> . The default is that all m predictors predict all p effect sizes. The format of this matrix follows as.mxMatrix . The regression coefficients can be constrained equally by using the same labels. |
| <code>RE.constraints</code> | A $p \times p$ matrix specifying the variance components of the random effects. If the input is not a matrix, it is converted into a matrix by <code>as.matrix()</code> . The default |

is that all covariance/variance components are free. The format of this matrix follows [as.mxMatrix](#). Elements of the variance components can be constrained equally by using the same labels. If a zero matrix is specified, it becomes a fixed-effects meta-analysis.

RE.startvalues	A vector of p starting values on the diagonals of the variance component of the random effects. If only one scalar is given, it will be duplicated across the diagonals. Starting values for the off-diagonals of the variance component are all 0. A $p \times p$ symmetric matrix of starting values is also accepted.
RE.lbound	A vector of p lower bounds on the diagonals of the variance component of the random effects. If only one scalar is given, it will be duplicated across the diagonals. Lower bounds for the off-diagonals of the variance component are set at NA. A $p \times p$ symmetric matrix of the lower bounds is also accepted.
intervals.type	Either z (default if missing) or LB. If it is z, it calculates the 95% Wald confidence intervals (CIs) based on the z statistic. If it is LB, it calculates the 95% likelihood-based CIs on the parameter estimates. Note that the z values and their associated p values are based on the z statistic. They are not related to the likelihood-based CIs.
I2	Possible options are "I2q", "I2hm" and "I2am". They represent the I2 calculated by using a typical within-study sampling variance from the Q statistic, the harmonic mean and the arithmetic mean of the within-study sampling variances (Xiong, Miller, & Morris, 2010). More than one options are possible. If intervals.type="LB", 95% confidence intervals on the heterogeneity indices will be constructed.
R2	Logical. If TRUE and there are predictors, R2 is calculated (Raudenbush, 2009).
model.name	A string for the model name in mxModel .
suppressWarnings	Logical. If TRUE, warnings are suppressed. The argument to be passed to mxRun .
silent	Logical. An argument to be passed to mxRun
run	Logical. If FALSE, only return the mx model without running the analysis.
...	Further arguments to be passed to mxRun
av	An auxiliary variable or a $k \times m$ matrix of auxiliary variables where m is the number of auxiliary variables.

Value

An object of class meta with a list of

call	Object returned by match.call
data	A data matrix of y, v and x
no.y	Number of effect sizes
no.x	Number of predictors
miss.x	A vector indicating whether the predictors are missing. Studies will be removed before the analysis if they are TRUE
I2	Types of I2 calculated

R2	Logical
mx.fit	A fitted object returned from mxRun
mx0.fit	A fitted object without any predictor returned from mxRun

Note

Missing values (NA) in y and their related elements in v will be removed automatically. When there are missing values in v but not in y, missing values will be replaced by 1e5. Effectively, these effect sizes will have little impact on the analysis. `metaFIML()` uses FIML to handle missing covariates in X. It is experimental. It may not be stable.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2008). A model for integrating fixed-, random-, and mixed-effects meta-analyses into structural equation modeling. *Psychological Methods*, **13**, 182-202.
- Cheung, M. W.-L. (2009). Constructing approximate confidence intervals for parameters with structural equation models. *Structural Equation Modeling*, **16**, 267-294.
- Cheung, M. W.-L. (2013). Multivariate meta-analysis as structural equation models. *Structural Equation Modeling*, **20**, 429-454.
- Cheung, M. W.-L. (2015). *Meta-analysis: A structural equation modeling approach*. Chichester, West Sussex: John Wiley & Sons, Inc.
- Hardy, R. J., & Thompson, S. G. (1996). A likelihood approach to meta-analysis with random effects. *Statistics in Medicine*, **15**, 619-629.
- Neale, M. C., & Miller, M. B. (1997). The use of likelihood-based confidence intervals in genetic models. *Behavior Genetics*, **27**, 113-120.
- Raudenbush, S. W. (2009). Analyzing effect sizes: random effects models. In H. M. Cooper, L. V. Hedges, & J. C. Valentine (Eds.), *The handbook of research synthesis and meta-analysis* (2nd ed., pp. 295-315). New York: Russell Sage Foundation.
- Xiong, C., Miller, J. P., & Morris, J. C. (2010). Measuring study-specific heterogeneity in meta-analysis: application to an antecedent biomarker study of Alzheimer's disease. *Statistics in Biopharmaceutical Research*, **2**(3), 300-309. doi:10.1198/sbr.2009.0067

See Also

[reml](#), [Hox02](#), [Berkey98](#), [wvs94a](#)

meta2semPlot

*Convert metaSEM objects into semPlotModel objects for plotting***Description**

It converts objects in class `wls` into objects of class `semPlotModel`.

Usage

```
meta2semPlot(
  object,
  manNames = NULL,
  latNames = NULL,
  labels = c("labels", "RAM"),
  ...
)
```

Arguments

<code>object</code>	An object of class <code>wls</code> returned from <code>wls()</code> or <code>tssem2()</code> .
<code>manNames</code>	A character vector of the manifest names. The program will try to get it from the object if it is not given.
<code>latNames</code>	A character vector of the latent names. The program will create it by using "L1", "L2", etc if it is not given.
<code>labels</code>	Either <code>labels</code> (default if missing) or <code>RAM</code> . If <code>labels</code> , the labels of the parameters are used in plotting. If <code>RAM</code> , the RAM notations are used in plotting.
<code>...</code>	Further arguments to be passed to ramModel

Details

It uses the `ramModel()` to do the conversion.

Value

A "semPlotModel" object.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[ramModel](#), [Becker92](#), [Becker09](#), [Digman97](#), [Hunter83](#)

meta3L

*Three-Level Univariate Meta-Analysis with Maximum Likelihood Estimation***Description**

It conducts three-level univariate meta-analysis with maximum likelihood estimation method. Mixed-effects meta-analysis can be conducted by including study characteristics as predictors. Equality constraints on the intercepts, regression coefficients and variance components on the level-2 and on the level-3 can be easily imposed by setting the same labels on the parameter estimates.

Usage

```
meta3L(
  y,
  v,
  cluster,
  x,
  data,
  intercept.constraints = NULL,
  coef.constraints = NULL,
  RE2.constraints = NULL,
  RE2.lbound = 1e-10,
  RE3.constraints = NULL,
  RE3.lbound = 1e-10,
  intervals.type = c("z", "LB"),
  I2 = "I2q",
  R2 = TRUE,
  model.name = "Meta analysis with ML",
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)
```

```
meta3(
  y,
  v,
  cluster,
  x,
  data,
  intercept.constraints = NULL,
  coef.constraints = NULL,
  RE2.constraints = NULL,
  RE2.lbound = 1e-10,
  RE3.constraints = NULL,
  RE3.lbound = 1e-10,
```

```

    intervals.type = c("z", "LB"),
    I2 = "I2q",
    R2 = TRUE,
    model.name = "Meta analysis with ML",
    suppressWarnings = TRUE,
    silent = TRUE,
    run = TRUE,
    ...
)

meta3LFIML(
  y,
  v,
  cluster,
  x2,
  x3,
  av2,
  av3,
  data,
  intercept.constraints = NULL,
  coef.constraints = NULL,
  RE2.constraints = NULL,
  RE2.lbound = 1e-10,
  RE3.constraints = NULL,
  RE3.lbound = 1e-10,
  intervals.type = c("z", "LB"),
  R2 = TRUE,
  model.name = "Meta analysis with ML",
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)

meta3X(
  y,
  v,
  cluster,
  x2,
  x3,
  av2,
  av3,
  data,
  intercept.constraints = NULL,
  coef.constraints = NULL,
  RE2.constraints = NULL,
  RE2.lbound = 1e-10,
  RE3.constraints = NULL,

```

```

    RE3.lbound = 1e-10,
    intervals.type = c("z", "LB"),
    R2 = TRUE,
    model.name = "Meta analysis with ML",
    suppressWarnings = TRUE,
    silent = TRUE,
    run = TRUE,
    ...
)

```

Arguments

<code>y</code>	A vector of k studies of effect size.
<code>v</code>	A vector of k studies of sampling variance.
<code>cluster</code>	A vector of k string or number indicating the clusters.
<code>x</code>	A predictor or a $k \times m$ matrix of level-2 and level-3 predictors where m is the number of predictors.
<code>data</code>	An optional data frame containing the variables in the model.
<code>intercept.constraints</code>	A 1 x 1 matrix specifying whether the intercept of the effect size is fixed or constrained. The format of this matrix follows as.mxMatrix . The intercept can be constrained with other parameters by using the same label.
<code>coef.constraints</code>	A 1 x m matrix specifying how the level-2 and level-3 predictors predict the effect sizes. If the input is not a matrix, it is converted into a matrix by <code>as.matrix()</code> . The default is that all m predictors predict the effect size. The format of this matrix follows as.mxMatrix . The regression coefficients can be constrained equally by using the same labels.
<code>RE2.constraints</code>	A scalar or a 1 x 1 matrix specifying the variance components of the random effects. The default is that the variance components are free. The format of this matrix follows as.mxMatrix . Elements of the variance components can be constrained equally by using the same label.
<code>RE2.lbound</code>	A scalar or a 1 x 1 matrix of lower bound on the level-2 variance component of the random effects.
<code>RE3.constraints</code>	A scalar or a 1 x 1 matrix specifying the variance components of the random effects at level-3. The default is that the variance components are free. The format of this matrix follows as.mxMatrix . Elements of the variance components can be constrained equally by using the same label.
<code>RE3.lbound</code>	A scalar or a 1 x 1 matrix of lower bound on the level-3 variance component of the random effects.
<code>intervals.type</code>	Either <code>z</code> (default if missing) or <code>LB</code> . If it is <code>z</code> , it calculates the 95% Wald confidence intervals (CIs) based on the z statistic. If it is <code>LB</code> , it calculates the 95% likelihood-based CIs on the parameter estimates. Note that the z values and their associated p values are based on the z statistic. They are not related to the likelihood-based CIs.

I2	Possible options are "I2q", "I2hm", "I2am" and "ICC". They represent the I2 calculated by using a typical within-study sampling variance from the Q statistic, the harmonic mean, the arithmetic mean of the within-study sampling variances, and the intra-class correlation. More than one options are possible. If <code>intervals.type="LB"</code> , 95% confidence intervals on the heterogeneity indices will be constructed.
R2	Logical. If TRUE and there are predictors, R2 is calculated.
<code>model.name</code>	A string for the model name in <code>mxModel</code> .
<code>suppressWarnings</code>	Logical. If TRUE, warnings are suppressed. It is passed to <code>mxRun</code> .
<code>silent</code>	Logical. An argument to be passed to <code>mxRun</code>
<code>run</code>	Logical. If FALSE, only return the mx model without running the analysis.
<code>...</code>	Further arguments to be passed to <code>mxRun</code>
<code>x2</code>	A predictor or a $k \times m$ matrix of level-2 predictors where m is the number of predictors.
<code>x3</code>	A predictor or a $k \times m$ matrix of level-3 predictors where m is the number of predictors.
<code>av2</code>	A predictor or a $k \times m$ matrix of level-2 auxiliary variables where m is the number of variables.
<code>av3</code>	A predictor or a $k \times m$ matrix of level-3 auxiliary variables where m is the number of variables.

Details

$$y_{ij} = \beta_0 + \beta' * \mathbf{x}_{ij} + u_{(2)ij} + u_{(3)j} + e_{ij}$$

where y_{ij} is the effect size for the i th study in the j th cluster, β_0 is the intercept, β is the regression coefficients, $\mathbf{x}_{ij}$ is a vector of predictors, $u_{(2)ij} \sim N(0, \tau_2^2)$ and $u_{(3)j} \sim N(0, \tau_3^2)$ are the level-2 and level-3 heterogeneity variances, respectively, and $e_{ij} \sim N(0, v_{ij})$ is the conditional known sampling variance.

`meta3L()` does not differentiate between level-2 or level-3 variables in $\mathbf{x}$ since both variables are treated as a design matrix. When there are missing values in $\mathbf{x}$, the data will be deleted. `meta3LFIML()` treats the predictors `x2` and `x3` as level-2 and level-3 variables. Thus, their means and covariance matrix will be estimated. Missing values in `x2` and `x3` will be handled by (full information) maximum likelihood (FIML) in `meta3LFIML()`. Moreover, auxiliary variables `av2` at level-2 and `av3` at level-3 may be included to improve the estimation. Although `meta3LFIML()` is more flexible in handling missing covariates, it is more likely to encounter estimation problems.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2014). Modeling dependent effect sizes with three-level meta-analyses: A structural equation modeling approach. *Psychological Methods*, **19**, 211-229.
- Enders, C. K. (2010). *Applied missing data analysis*. New York: Guilford Press.
- Graham, J. (2003). Adding missing-data-relevant variables to FIML-based structural equation models. *Structural Equation Modeling: A Multidisciplinary Journal*, **10**(1), 80-100.
- Konstantopoulos, S. (2011). Fixed effects and variance components estimation in three-level meta-analysis. *Research Synthesis Methods*, **2**, 61-76.

See Also

[reml3L](#), [Cooper03](#), [Bornmann07](#)

Nam03	<i>Dataset on the Environmental Tobacco Smoke (ETS) on children's health</i>
-------	--

Description

This dataset includes 59 studies reported by Nam, Mengersen, and Garthwaite (2003) on the potential health effects among children exposed to environmental tobacco smoke (ETS), or passive smoking. The effect sizes are the log odds ratios of asthma and lower respiratory disease (LRD).

Details

A list of data with the following structure:

ID Study identification number.

Size Total number of valid subjects in the study.

Age Mean age of participants.

Year Year of publication.

Country Country code.

Smoke Source of ETS.

Adj Whether the reported odds ratio is adjusted for covariates.

Asthma_logOR Log odds ratio of asthma.

LRD_logOR Log odds ratio of lower respiratory disease.

Asthma_v Sampling variance of Asthma_logOR.

AsthmaLRD_cov_05 Sampling covariance between Asthma_logOR and LRD_logOR by assuming a correlation of 0.5

LRD_v Sampling variance of LRD_logOR.

Source

Nam, I.-S., Mengersen, K., & Garthwaite, P. (2003). Multivariate meta-analysis. *Statistics in Medicine*, **22**(14), 2309-2333. <https://doi.org/10.1002/sim.1410>

Examples

```
data(Nam03)
```

Nohe15

Correlation Matrices from Nohe et al. (2015)

Description

The data sets include two lists of correlation matrices of panel studies between work-family conflict and strain reported in Table A1 (Nohe15A1) and Table A2 (Nohe15A2) by Nohe et al. (2015).

Details

A list of data with the following structure:

data A list of studies of correlation matrices. The variables are *W1*, *SI*, *W2*, and *S2* in Nohe15A1 and *F1*, *SI*, *F2*, and *S2* in Nohe15A2

n A vector of sample sizes

RelXX The reliabilities of *W1*, *SI*, *W2* and *S2* in Nohe15A1 and the reliabilities of *F1*, *SI*, *F2*, and *S2* in Nohe15A2

FemalePer Percentage of female participants

Publication Whether the studies were published (*P*) or unpublished (*U*)

Lag Time lag between the coded measurement waves in months

Source

Nohe, C., Meier, L. L., Sonntag, K., & Michel, A. (2015). The chicken or the egg? A meta-analysis of panel studies of the relationship between work-family conflict and strain. *Journal of Applied Psychology*, **100**(2), 522-536.

Examples

```
#### TSSEM
```

```
## Set seed for replicability
set.seed(23891)
```

```
## Table A1
randA1a <- tssem1(Nohe15A1$data, Nohe15A1$n, method="REM", RE.type="Diag")
summary(randA1a)
```

```

model1 <- 'W2 ~ w2w*W1 + s2w*S1
          S2 ~ w2s*W1 + s2s*S1
          W1 ~~ w1WITHs1*S1
          W2 ~~ w2WITHs2*S2
          W1 ~~ 1*W1
          S1 ~~ 1*S1
          W2 ~~ Errw2*W2
          S2 ~~ Errs2*S2'

## Display the model
plot(model1, layout="spring")

RAM1 <- lavaan2RAM(model1, obs.variables=c("W1", "S1", "W2", "S2"))
RAM1

randA1b <- tssem2(randA1a, RAM=RAM1)
summary(randA1b)

## Display the model with the parameter estimates
plot(randA1b, layout="spring")

## Table A2
randA2a <- tssem1(Nohe15A2$data, Nohe15A2$n, method="REM", RE.type="Diag")
## Rerun to remove error code
randA2a <- rerun(randA2a)
summary(randA2a)

model2 <- 'F2 ~ f2f*F1 + s2F*S1
          S2 ~ f2s*F1 + s2s*S1
          F1 ~~ f1WITHs1*S1
          F2 ~~ f2WITHs2*S2
          F1 ~~ 1*F1
          S1 ~~ 1*S1
          F2 ~~ Errf2*F2
          S2 ~~ Errs2*S2'

## Display the model
plot(model2, layout="spring")

RAM2 <- lavaan2RAM(model2, obs.variables=c("F1", "S1", "F2", "S2"))
RAM2

randA2b <- tssem2(randA2a, RAM=RAM2)
summary(randA2b)

## Display the model with the parameter estimates
plot(randA2b, layout="spring")

## Estimate the heterogeneity of the parameter estimates
tssemParaVar(randA1a, randA2b)

## Parametric bootstrap based on Yu et al. (2016)

```

```

## I assume that you know what you are doing!

## Set seed for reproducibility
set.seed(39128482)

## Average the correlation coefficients with the univariate-r approach
uni1 <- uniR1(Nohe15A1$data, Nohe15A1$n)
uni1

## Generate random correlation matrices
boot.cor <- bootuniR1(uni1, Rep=50)

## Display the quality of the generated correlation matrices
summary(boot.cor)

## Proposed saturated model
model1 <- 'W2 + S2 ~ W1 + S1'

## Use the harmonic mean of the sample sizes as n in SEM
n <- uni1$n.harmonic

boot.fit1 <- bootuniR2(model=model1, data=boot.cor, n=n)
summary(boot.fit1)

## Proposed model with equal regression coefficients
model2 <- 'W2 ~ Same*W1 + Cross*S1
          S2 ~ Cross*W1 + Same*S1'

boot.fit2 <- bootuniR2(model=model2, data=boot.cor, n=n)
summary(boot.fit2)

#### OSMASEM

## Calculate the sampling variance-covariance matrix of the correlation matrices.
my.df <- Cor2DataFrame(Nohe15A1)

## Standardize the moderator "Lag"
my.df$data$Lag <- scale(my.df$data$Lag)

head(my.df$data)

## Proposed model
model1 <- 'W2 ~ w2w*W1 + s2w*S1
          S2 ~ w2s*W1 + s2s*S1
          W1 ~~ w1WITHs1*S1
          W2 ~~ w2WITHs2*S2
          W1 ~~ 1*W1
          S1 ~~ 1*S1
          W2 ~~ Errw2*W2
          S2 ~~ Errs2*S2'
plot(model1)

## Convert it into RAM specification

```

```

RAM1 <- lavaan2RAM(model1, obs.variables=c("W1", "S1", "W2", "S2"))
RAM1

## Create vechs of the model implied correlation matrix
## with implicit diagonal constraints
## M0 <- create.vechsR(A0=RAM1$A, S0=RAM1$S)

## Create heterogeneity variances
## RE.type= either "Diag" or "Symm"
##
## Transform= either "expLog" or "sqSD" for better estimation on variances
## T0 <- create.Tau2(RAM=RAM1, RE.type="Diag")
##
## Fit the model
## fit0 <- osmasem(model.name="No moderator", Mmatrix=M0, Tmatrix=T0, data=my.df)

## Fit the model
fit0 <- osmasem(model.name="No moderator", RAM=RAM1, data=my.df)
summary(fit0)

## Get the SRMR
osmasemSRMR(fit0)

## Get the transformed variance component of the random effects
VarCorr(fit0)

## "lag" as a moderator on A matrix
A1 <- matrix(c(0,0,0,0,
               0,0,0,0,
               "0*data.Lag","0*data.Lag",0,0,
               "0*data.Lag","0*data.Lag",0,0),
             nrow=4, ncol=4, byrow=TRUE)

## M1 <- create.vechsR(A0=RAM1$A, S0=RAM1$S, Ax=A1)
##
## Fit the model
## fit1 <- osmasem(model.name="Lag as a moderator for Amatrix", Mmatrix=M1,
##                 Tmatrix=T0, data= my.df)

fit1 <- osmasem(model.name="Lag as a moderator for Amatrix",
               RAM=RAM1, Ax=A1, data= my.df)
summary(fit1)
VarCorr(fit1)

## Compare the models with and without the moderator "lag"
anova(fit1, fit0)

## Calculate the R2
osmasemR2(fit0, fit1)

```

Norton13	<i>Studies on the Hospital Anxiety and Depression Scale Reported by Norton et al. (2013)</i>
----------	--

Description

The data set includes 28 studies on 14 items measuring the Hospital Anxiety and Depression Scale (HADS) Reported by Norton et al. (2013).

Details

The variables are:

data A list of 28 studies of correlation matrices. The variables are 14 items (x1 to x14) measuring HADS.

n A vector of sample sizes

population A vector of the population of the data

group A vector of classification into *patients* vs. *non-patients* based on population

Source

Norton, S., Cosco, T., Doyle, F., Done, J., & Sacker, A. (2013). The Hospital Anxiety and Depression Scale: A meta confirmatory factor analysis. *Journal of Psychosomatic Research*, 74(1), 74-81.

References

Jak, S., & Cheung, M. W.-L. (2018). Addressing heterogeneity in meta-analytic structural equation modeling using subgroup analysis. *Behavior Research Methods*, **50**, 1359-1373.

Examples

```
data(Norton13)
```

osmasem	<i>One-stage meta-analytic structural equation modeling</i>
---------	---

Description

It fits MASEM with the one-stage MASEM (OSMASEM) approach.

Usage

```

osmasem(
  model.name = "osmasem",
  RAM = NULL,
  Mmatrix = NULL,
  Tmatrix = NULL,
  Jmatrix = NULL,
  Ax = NULL,
  Sx = NULL,
  A.lbound = NULL,
  A.ubound = NULL,
  RE.type = c("Diag", "CSymm", "HCSymm", "Symm", "Zero", "User"),
  RE.User = NULL,
  data,
  subset.variables = NULL,
  subset.rows = NULL,
  intervals.type = c("z", "LB"),
  mxModel.Args = NULL,
  mxRun.Args = NULL,
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)

osmasem2(
  model.name = "osmasem2",
  RAM,
  data,
  cor.analysis = TRUE,
  RE.type.Sigma = c("Diag", "Symm", "Zero"),
  RE.type.Mu = c("Symm", "Diag", "Zero"),
  RE.type.SigmaMu = c("Zero", "Full"),
  mean.analysis = FALSE,
  startvalues = NULL,
  replace.constraints = FALSE,
  mxModel.Args = NULL,
  run = TRUE,
  ...
)

```

Arguments

<code>model.name</code>	A string for the model name in <code>mxModel</code> .
<code>RAM</code>	A RAM object including a list of matrices of the model returned from <code>lavaan2RAM</code> . If it is given, <code>Mmatrix</code> and <code>Tmatrix</code> arguments will be ignored.
<code>Mmatrix</code>	A list of matrices of the model implied correlation matrix created by the <code>create.vechsR</code> . It is only required when <code>RAM</code> is null.

Tmatrix	A list of matrices of the heterogeneity variance-covariance matrix created by the <code>create.Tau2</code> . It is only required when RAM is null.
Jmatrix	The Jacobian matrix of the mean structure in <code>mxMatrix</code> . The covariance structure is <code>Jmatrix %&% Tau2 + Vi</code> . If it is not given, an identity matrix will be used.
Ax	A <code>Amatrix</code> of a list of <code>Amatrix</code> with definition variables as the moderators of the <code>Amatrix</code> . It is used to create the <code>Mmatrix</code> .
Sx	A <code>Smatrix</code> of a list of <code>Smatrix</code> with definition variables as the moderators of the <code>Smatrix</code> . It is used to create the <code>Mmatrix</code> .
A.lbound	A matrix of lower bound of the <code>Amatrix</code> . If a scalar is given, the lbound matrix will be filled with this scalar.
A.ubound	A matrix of upper bound of the <code>Amatrix</code> . If a scalar is given, the ubound matrix will be filled with this scalar.
RE.type	Type of the random effects. See <code>create.Tau2</code> for details.
RE.User	A symmetric matrix of TRUE or FALSE for the variance component when <code>RE.type="User"</code> . See <code>create.Tau2</code> for details.
data	A list of data created by the <code>Cor2DataFrame</code> .
subset.variables	A character vector of the observed variables selected for the analysis.
subset.rows	A logical vector of the same length as the number of rows in the data to select which rows are used in the analysis.
intervals.type	Either z (default if missing) or LB. If it is z, it calculates the 95% confidence intervals (CIs) based on the estimated standard error. If it is LB, it calculates the 95% likelihood-based CIs on the parameter estimates.
mxModel.Args	A list of arguments passed to <code>mxModel</code> .
mxRun.Args	A list of arguments passed to <code>mxRun</code> .
suppressWarnings	Logical. If it is TRUE, warnings are suppressed. This argument is passed to <code>mxRun</code> .
silent	Logical. An argument is passed to <code>mxRun</code>
run	Logical. If FALSE, only return the mx model without running the analysis.
...	Not used yet.
cor.analysis	Whether to analyze correlation or covariance structure analysis.
RE.type.Sigma	Type of the random effects of the correlation or covariance vectors.
RE.type.Mu	Type of the random effects of the mean vectors.
RE.type.SigmaMu	Type of the random effects between the correlation/covariance vectors and the mean vectors.
mean.analysis	Whether to include the analysis of the mean structure.
startvalues	An optional list of starting values. It is useful when there are new parameters in RAM.
replace.constraints	It is relevant only when there are constraints in RAM. If it is FALSE, these constraints will be imposed. If it is TRUE, the parameters on the left-hand side will be replaced by the algebras on the right-hand side.

Details

osmasem() was implemented based on Jak and Cheung (2020) for meta-analyzing correlation matrices. osmasem2() was a rewritten version to handle correlation or covariance matrices with the means. There are several major differences between them: 1. osmasem() allows the use of RAM or (Mmatrix and Tmatrix), while osmasem2() only accepts the RAM input. 2. RE.type is used to specify the type of random effects on the correlations in osmasem(). On the contrary, osmasem2() has three types of random effects: correlations/covariances, means, and covariance between correlations/covariance and means. 3. osmasem() reports the transformed random effects in the parameter table. Users have to use VarCorr() to obtain the heterogeneity matrix of the random effects. In contrast, osmasem2() reports the heterogeneity matrix in the parameter table. 4. osmasem() accepts either intervals.type="z" or intervals.type="LB", whereas osmasem2() only uses intervals.type="z". Thus, this argument is removed in osmasem2(). 5. osmasem2() allows the imposing linear and nonlinear constraints and the creation of parameter functions in RAM, which osmasem cannot.

Value

An object of class osmasem

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Jak, S., & Cheung, M. W.-L. (2020). Meta-analytic structural equation modeling with moderating effects on SEM parameters. *Psychological Methods*, **25** (4), 430-455. <https://doi.org/10.1037/met0000245>

Jak, S., Cheung, M. W.-L., Acar, S., & Kindred, R. (December 18, 2024). Evaluating differences in latent means across studies: Extending meta-analytic confirmatory factor analysis with the analysis of means. OSF. <https://doi.org/10.31234/osf.io/35gtz>.

See Also

[Cor2DataFrame](#), [create.vechsR](#), [create.Tau2](#), [create.V](#), [osmasem](#), [Nohe15](#), [issp05](#)

osmasemR2	<i>Calculate the R2 in OSMASEM and OSMASEM3L</i>
-----------	--

Description

It calculates the R2 of the moderators in explaining the variances in the heterogeneity variances.

Usage

osmasemR2(model11, model0, R2.truncate = TRUE)

Arguments

model1	An object in class osmasem.
model0	An object in class osmasem.
R2.truncate	Whether to truncate the negative R2 to zero.

Value

model1 and model0 are the models with and without the moderators, respectively. The function does not check whether the models are nested. It is the users' responsibility to make sure that the models with and without the moderators are nested. It returns a list of the diagonals of the heterogeneity variances of the models without and with the moderators, and the R2.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[osmasem](#)

osmasemSRMR

Calculate the SRMR in OSMASEM and OSMASEM3L

Description

It calculates the standardized root mean squared residuals (SRMR) in OSMASEM and OSMASEM3L.

Usage

```
osmasemSRMR(x)
```

Arguments

x	An OSMASEM object without any moderators.
---	---

Value

It calculates the model implied correlation matrix and its saturated counterpart to calculate the SRMR. It should be noted that the heterogeneity variances are ignored in the calculations.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[osmasem](#), [Nohe15](#)

pattern.n

Display the Accumulative Sample Sizes for the Covariance Matrix

Description

It displays the accumulative sample sizes for the covariance matrix.

Usage

```
pattern.n(x, n)
```

Arguments

x	A list of square matrices
n	A vector of sample sizes.

Value

A square matrix of the accumulative sample sizes of the input matrices.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
## Show the pattern of missing data
pattern.n(Hunter83$data, Hunter83$n)

#           Ability Knowledge Work sample Supervisor
# Ability      3815      3372      3281      3605
# Knowledge     3372      3532      2998      3322
# Work sample   3281      2998      3441      3231
# Supervisor    3605      3322      3231      3765
```

pattern.na

Display the Pattern of Missing Data of a List of Square Matrices

Description

It displays the pattern of missing data (or pattern of data that are present) of a list of square matrices with the same dimensions.

Usage

```
pattern.na(x, show.na = TRUE, type = c("tssem", "osmasem"))
```

Arguments

x	A list of square matrices
show.na	If it is TRUE, it shows the pattern of missing data. If it is FALSE, it shows the pattern of data that are present.
type	If it is tssem, it reports the pattern of missing correlations for the tssem approach. If it is osmasem, it reports the pattern of missing correlations for the data created by Cor2DataFrame .

Value

A square matrix of numerical values with the same dimensions of the input matrices.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
## Show the pattern of missing data
pattern.na(Hunter83$data, show.na=TRUE)

#           Ability Knowledge Work sample Supervisor
# Ability           1           3           3           2
# Knowledge          3           2           4           3
# Work sample        3           4           2           3
# Supervisor         2           3           3           1

## Show the pattern of data that are present
pattern.na(Hunter83$data, show.na=FALSE)

#           Ability Knowledge Work sample Supervisor
# Ability          13          11          11          12
# Knowledge         11          12          10          11
# Work sample       11          10          12          11
# Supervisor        12          11          11          13
```

plot	<i>Plot methods for various objects</i>
------	---

Description

It plots the models from either the lavaan model or meta, wls, and osmasem objects.

Arguments

<code>x</code>	An object returned from either a lavaan model class character, osmasem, osmasem3L, wls or meta
<code>effect.sizes</code>	Numeric values indicating which effect sizes to be plotted. At least two effect sizes are required. To plot the effect sizes of y_1 and y_2 , one may use <code>effect.sizes=c(1,2)</code> . If it is missing, all effect sizes will be plotted in a pairwise way.
<code>add.margin</code>	Value for additional margins on the left and bottom margins.
<code>interval</code>	Interval for the confidence ellipses.
<code>main</code>	Main title of each plot. If there are multiple plots, a vector of character titles may be used.
<code>axis.labels</code>	Labels for the effect sizes.
<code>study.col</code>	The color for individual studies. See <code>col</code> in par .
<code>study.pch</code>	Plotting character of individual studies. See <code>pch</code> in points .
<code>study.min.cex</code>	The minimum value of <code>cex</code> for individual studies. See <code>cex</code> in par .
<code>study.weight.plot</code>	Logical. If TRUE, the plotting size of individual studies (<code>cex</code>) will be proportional to one over the square root of the determinant of the sampling covariance matrix of the study.
<code>study.ellipse.plot</code>	Logical. If TRUE, the confidence ellipses of individual studies are plotted.
<code>study.ellipse.col</code>	The color of the confidence ellipses of individual studies. See <code>col</code> in par .
<code>study.ellipse.lty</code>	The line type of the confidence ellipse of individual studies. See <code>lty</code> in par .
<code>study.ellipse.lwd</code>	The line width of the confidence ellipse of individual studies. See <code>lwd</code> in par .
<code>estimate.col</code>	The color of the estimated effect size. See <code>col</code> in par .
<code>estimate.pch</code>	Plotting character of the estimated effect sizes. See <code>pch</code> in points .
<code>estimate.cex</code>	The amount of plotting of the estimated effect sizes. See <code>cex</code> in par .
<code>estimate.ellipse.plot</code>	Logical. If TRUE, the confidence ellipse of the estimated effect sizes will be plotted.
<code>estimate.ellipse.col</code>	The color of the confidence ellipse of the estimated effect sizes. See <code>col</code> in par .
<code>estimate.ellipse.lty</code>	The line type of the confidence ellipse of the estimated effect sizes. See <code>lty</code> in par .
<code>estimate.ellipse.lwd</code>	The line width of the confidence ellipse of the estimated effect sizes. See <code>lwd</code> in par .
<code>randeff.ellipse.plot</code>	Logical. If TRUE, the confidence ellipses of the random effects will be plotted.

<code>randeff.ellipse.col</code>	Color of the confidence ellipses of the random effects. See <code>col</code> in par .
<code>randeff.ellipse.lty</code>	The line type of the confidence ellipses of the random effects. See <code>lty</code> in par .
<code>randeff.ellipse.lwd</code>	The line width of the confidence ellipses of the random effects. See <code>lwd</code> in par .
<code>univariate.plot</code>	Logical. If TRUE, the estimated univariate effect sizes will be plotted.
<code>univariate.lines.col</code>	The color of the estimated univariate effect sizes. See <code>col</code> in par .
<code>univariate.lines.lty</code>	The line type of the estimated univariate effect sizes. See <code>lty</code> in par .
<code>univariate.lines.lwd</code>	The line width of the estimated univariate effect sizes. See <code>lwd</code> in par .
<code>univariate.polygon.width</code>	The width of the polygon of the estimated univariate effect sizes.
<code>univariate.polygon.col</code>	The color of the polygon of the estimated univariate effect sizes.
<code>univariate.arrows.col</code>	The color of the arrows of the estimated univariate effect sizes.
<code>univariate.arrows.lwd</code>	The line width of the arrows of the estimated univariate effect sizes.
<code>diag.panel</code>	Logical. If TRUE, diagonal panels will be created. They can then be used for forrest plots for univariate meta-analysis.
<code>xlim</code>	NULL or a numeric vector of length 2; if it is NULL, it provides defaults estimated from the data.
<code>ylim</code>	NULL or a numeric vector of length 2; if it is NULL, it provides defaults estimated from the data.
<code>fixed.x</code>	Argument passed to semPlotModel .
<code>manNames</code>	Argument passed to semPaths
<code>latNames</code>	Argument passed to semPaths
<code>labels</code>	Argument passed to semPaths
<code>what</code>	Argument passed to semPaths
<code>nCharNodes</code>	Argument passed to semPaths
<code>nCharEdges</code>	Argument passed to semPaths
<code>layout</code>	Argument passed to semPaths
<code>color</code>	Argument passed to semPaths
<code>sizeMan</code>	Argument passed to semPaths
<code>sizeLat</code>	Argument passed to semPaths
<code>edge.label.cex</code>	Argument passed to semPaths
<code>weighted</code>	Argument passed to semPaths

group	For <code>plot.mxsem</code> on a multiple-group sem fit (see sem): which group to plot. The original data value (e.g. "site-2", before any sanitizing into a legal <code>mxModel</code> name) always takes precedence over the possibly-sanitized submodel name, via <code>fit\$group.map</code> ; the sanitized submodel name itself (<code>names(fit\$mx.fit@submodels)</code>) only resolves to the matching group when it does not also collide with a <i>different</i> group's original value (e.g. original groups "a.b" and "a_b" both sanitize/map to a submodel named "a_b" or derived from it – passing "a_b" always means the group whose real data value is "a_b", never the submodel merely named that). When originals and submodel names coincide (the common case, e.g. "g1"), either happens to work, but the original value is the one to reach for. Defaults to the first group. Ignored otherwise.
level	For <code>plot.mxsem</code> on a two-level sem fit (see sem): either "within" (default) or "between", selecting which level to plot. Ignored otherwise (including when <code>combine=TRUE</code>). <code>semPlot</code> has no notion of the two levels' relational (<code>joinKey</code>) linkage, so only one level's diagram is shown at a time even when combined (below) into one figure.
combine	For <code>plot.mxsem</code> on a multiple-group or two-level sem fit: if <code>TRUE</code> , plot every group (or both the within and between levels) as side-by-side panels of one figure, overriding <code>group/level</code> . Defaults to <code>TRUE</code> when neither group nor level is supplied, and to <code>FALSE</code> (one diagram, as selected by <code>group/level</code>) as soon as either of them is – so <code>plot(fit)</code> shows every panel by default, but <code>plot(fit, group="g2")</code> or <code>plot(fit, level="between")</code> still shows just the one diagram requested without needing <code>combine=FALSE</code> as well. Passing <code>combine=TRUE</code> explicitly still shows every panel regardless of <code>group/level</code> . Ignored otherwise.
main.line	For <code>plot.mxsem</code> on a multiple-group or two-level sem fit: the line argument passed to title for each panel's "within"/ "between" or group-name title. Defaults to 2.5 (pushed up and out of the way of a variance-edge label that <code>semPaths</code> often draws close to the top margin). Ignored otherwise.
main.adj	For <code>plot.mxsem</code> on a multiple-group or two-level sem fit: the <code>adj</code> argument passed to title for each panel's title, controlling its horizontal position (0 = left-aligned, 0.5 = centered, 1 = right-aligned). Defaults to <code>NULL</code> , i.e. <code>title</code> 's own default (centered). Since the title is drawn over the diagram rather than reserving its own space, it can still land on top of an edge label that happens to sit centered near the top of that particular model's diagram; nudge it clear with e.g. <code>main.adj=0.8</code> (or 1 for fully right-aligned) if so. Ignored otherwise.
...	Further arguments passed to the methods.

Note

The estimated effect sizes and random effects are based on the labels `Intercept1`, `Intercept2`, ... and `Tau2_1_1`, `Tau2_2_1`, `Tau2_2_2`, etc. At least two effect sizes are required for this function.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Cheung, M. W.-L. (2013). Multivariate meta-analysis as structural equation models. *Structural Equation Modeling*, **20**, 429-454.

See Also

[Berkey98](#), [wvs94a](#) [meta2semPlot](#) [semPaths](#)

Examples

```
## lavaan model
model <- "y ~ m + x
         m ~ x"
plot(model)
```

<code>print</code>	<i>Print Methods for various Objects</i>
--------------------	--

Description

Print methods for the `tssem1FEM`, `tssem1FEM.cluster`, `tssem1REM`, `wls`, `meta`, `meta3LFIML`, `reml`, `uniR1` and `impliedR` objects.

Arguments

- `x` An object returned from either class `tssem1FEM`, class `tssem1FEM.cluster`, class `tssem1REM`, class `wls`, class `meta`, class `meta3LFIML`, class `reml`, class `uniR1` or class `impliedR`
- `...` Further arguments to be passed to `summary.default` or unused.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[tssem1](#), [wls](#), [meta](#), [reml](#)

rCor	<i>Generate (Nested) Sample/Population Correlation/Covariance Matrices</i>
------	--

Description

It generates (nested) random sample or population correlation or covariance matrices. `rCor()` is a wrapper to call `rCorPop()` and then `rCorSam()`.

Usage

```
rCor(
  Sigma,
  V,
  n,
  corr = TRUE,
  raw.data = FALSE,
  nonPD.pop = c("replace", "nearPD", "accept"),
  nonPD.sam = c("stop", "nearPD")
)

rCorPop(Sigma, V, k, corr = TRUE, nonPD.pop = c("replace", "nearPD", "accept"))

rCorSam(
  Sigma,
  n,
  corr = TRUE,
  raw.data = FALSE,
  nonPD.sam = c("stop", "nearPD")
)

rCor3L(
  Sigma,
  V.B,
  V.W,
  n,
  cluster,
  corr = TRUE,
  raw.data = FALSE,
  nonPD.pop = c("replace", "nearPD", "accept"),
  nonPD.sam = c("stop", "nearPD")
)
```

Arguments

Sigma	A list of population correlation/covariance matrices or a single matrix
V	A variance-covariance matrix of Sigma.

n	A vector or a single sample sizes.
corr	Logical. Whether to generate correlation or covariance matrices.
raw.data	Logical. Whether correlation/covariance matrices are generated via raw.data or directly from a Wishart distribution.
nonPD.pop	If it is replace, generated non-positive definite matrices are replaced by generated new ones which are positive definite. If it is nearPD, they are replaced by nearly positive definite matrices by calling <code>Matrix::nearPD()</code> . If it is accept, they are accepted.
nonPD.sam	If it is stop, the program stops when the inputs in the <code>rCorSam</code> are non-positive definite. If it is nearPD, they are replaced by nearly positive definite matrices by calling <code>Matrix::nearPD()</code> .
k	A vector or a single number of studies.
V.B	A variance-covariance matrix of between-study Sigma.
V.W	A variance-covariance matrix of within-study Sigma
cluster	A vector of number of studies in clusters.

Value

An object of the generated population/sample correlation/covariance matrices.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```

Sigma <- matrix(c(1, .2, .3,
                  .2, 1, .4,
                  .3, .4, 1), ncol=3, nrow=3)
V <- diag(c(.1, .1, .1))

## Generate two population correlation matrices
Pop.corr <- rCorPop(Sigma, V, k=2)
Pop.corr

summary(Pop.corr)

## Generate two sample correlation matrices
rCorSam(Sigma=Pop.corr, n=c(10, 10))

## The above code is the same as the following one
rCor(Sigma, V, n=c(10, 10))

```

readData

*Read External Correlation/Covariance Matrices***Description**

It reads full/lower triangle/stacked vectors of correlation/covariance data into a list of correlation/covariance matrices.

Arguments

file	File name of the data.
no.var	The number of variables in the data.
...	Further arguments to be passed to scan for readLowTriMat and to read.table for readFullMat and readStackVec.

Value

A list of correlation/covariance matrices.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
## Write two full correlation matrices into a file named "fullmat.dat".
## x2 is missing in the second matrix.
## The content of "fullmat.dat" is
# 1.0 0.3 0.4
# 0.3 1.0 0.5
# 0.4 0.5 1.0
# 1.0 NA 0.4
# NA NA NA
# 0.4 NA 1.0

## cat("1.0 0.3 0.4\n0.3 1.0 0.5\n0.4 0.5 1.0
## 1.0 NA 0.4\nNA NA NA\n0.4 NA 1.0",
## file="fullmat.dat", sep="")

## Read the correlation matrices from a file
## my.full <- readFullMat("fullmat.dat")

## Read the correlation matrices from a string
x <-
"1.0 0.3 0.4
0.3 1.0 0.5
0.4 0.5 1.0
1.0 NA 0.4"
```

```

NA NA NA
0.4 NA 1.0"

my.full <- readFullMat(textConnection(x))

## my.full
# $`1`
#      x1  x2  x3
# x1 1.0 0.3 0.4
# x2 0.3 1.0 0.5
# x3 0.4 0.5 1.0
#
# $`2`
#      x1 x2  x3
# x1 1.0 NA 0.4
# x2 NA NA  NA
# x3 0.4 NA 1.0

## Write two lower triangle correlation matrices into a file named "lowertriangle.dat".
## x2 is missing in the second matrix.
## The content of "lowertriangle.dat" is
# 1.0
# 0.3 1.0
# 0.4 0.5 1.0
# 1.0
# NA NA
# 0.4 NA 1.0
## cat("1.0\n0.3 1.0\n0.4 0.5 1.0\n1.0\nNA NA\n0.4 NA 1.0",
##      file="lowertriangle.dat", sep="")

## Read the lower triangle correlation matrices from a file
## my.lowertri <- readLowTriMat(file = "lowertriangle.dat", no.var = 3)

## Read the correlation matrices from a string
x <-
"1.0
0.3 1.0
0.4 0.5 1.0
1.0
NA NA
0.4 NA 1.0"

my.lowertri <- readLowTriMat(textConnection(x), no.var = 3)

## my.lowertri
# $`1`
#      x1  x2  x3
# x1 1.0 0.3 0.4
# x2 0.3 1.0 0.5
# x3 0.4 0.5 1.0
#
# $`2`
#      x1 x2  x3

```

```

# x1 1.0 NA 0.4
# x2 NA NA NA
# x3 0.4 NA 1.0

## Write two vectors of correlation coefficients based on
## column major into a file named "stackvec.dat".
## x2 is missing in the second matrix.
## The content of "stackvec.dat" is
# 1.0 0.3 0.4 1.0 0.5 1.0
# 1.0 NA 0.4 NA NA 1.0
## cat("1.0 0.3 0.4 1.0 0.5 1.0\n1.0 NA 0.4 NA NA 1.0\n",
##      file="stackvec.dat", sep="")

## Read the stack vectors from a file
## my.vec <- readStackVec("stackvec.dat")

## Read the stack vectors from a string
x <- "
1.0 0.3 0.4 1.0 0.5 1.0
1.0 NA 0.4 NA NA 1.0"

my.vec <- readStackVec(textConnection(x))

## my.vec
# $`1`
#      x1  x2  x3
# x1 1.0 0.3 0.4
# x2 0.3 1.0 0.5
# x3 0.4 0.5 1.0
#
# $`2`
#      x1 x2  x3
# x1 1.0 NA 0.4
# x2 NA NA NA
# x3 0.4 NA 1.0

```

reml

Estimate Variance Components with Restricted (Residual) Maximum Likelihood Estimation

Description

It estimates the variance components of random-effects in univariate and multivariate meta-analysis with restricted (residual) maximum likelihood (REML) estimation method.

Usage

```
reml(
```

```

y,
v,
x,
data,
RE.constraints = NULL,
RE.startvalues = 0.1,
RE.lbound = 1e-10,
intervals.type = c("z", "LB"),
model.name = "Variance component with REML",
suppressWarnings = TRUE,
silent = TRUE,
run = TRUE,
...
)

```

Arguments

<code>y</code>	A vector of effect size for univariate meta-analysis or a $k \times p$ matrix of effect sizes for multivariate meta-analysis where k is the number of studies and p is the number of effect sizes.
<code>v</code>	A vector of the sampling variance of the effect size for univariate meta-analysis or a $k \times p^*$ matrix of the sampling covariance matrix of the effect sizes for multivariate meta-analysis where $p^* = p(p + 1)/2$. It is arranged by column major as used by vech .
<code>x</code>	A predictor or a $k \times m$ matrix of predictors where m is the number of predictors.
<code>data</code>	An optional data frame containing the variables in the model.
<code>RE.constraints</code>	A $p \times p$ matrix specifying the variance components of the random effects. If the input is not a matrix, it is converted into a matrix by <code>as.matrix()</code> . The default is that all covariance/variance components are free. The format of this matrix follows as.mxMatrix . Elements of the variance components can be constrained equally by using the same labels. If a zero matrix is specified, it becomes a fixed-effects meta-analysis.
<code>RE.startvalues</code>	A vector of p starting values on the diagonals of the variance component of the random effects. If only one scalar is given, it will be repeated across the diagonals. Starting values for the off-diagonals of the variance component are all 0. A $p \times p$ symmetric matrix of starting values is also accepted.
<code>RE.lbound</code>	A vector of p lower bounds on the diagonals of the variance component of the random effects. If only one scalar is given, it will be repeated across the diagonals. Lower bounds for the off-diagonals of the variance component are set at NA. A $p \times p$ symmetric matrix of the lower bounds is also accepted.
<code>intervals.type</code>	Either <code>z</code> (default if missing) or <code>LB</code> . If it is <code>z</code> , it calculates the 95% Wald confidence intervals (CIs) based on the z statistic. If it is <code>LB</code> , it calculates the 95% likelihood-based CIs on the parameter estimates. Note that the z values and their associated p values are based on the z statistic. They are not related to the likelihood-based CIs.
<code>model.name</code>	A string for the model name in mxModel .

<code>suppressWarnings</code>	Logical. If TRUE, warnings are suppressed. It is passed to <code>mxRun</code> .
<code>silent</code>	Logical. An argument to be passed to <code>mxRun</code>
<code>run</code>	Logical. If FALSE, only return the mx model without running the analysis.
<code>...</code>	Further arguments to be passed to <code>mxRun</code>

Details

Restricted (residual) maximum likelihood obtains the parameter estimates on the transformed data that do not include the fixed-effects parameters. A transformation matrix $M = I - X(X'X)^{-1}X'$ is created based on the design matrix X which is just a column vector when there is no predictor in x . The last N redundant rows of M is removed where N is the rank of X . After pre-multiplying by M on y , the parameters of fixed-effects are removed from the model. Thus, only the parameters of random-effects are estimated.

An alternative but equivalent approach is to minimize the $-2 \times \log$ -likelihood function:

$$\log(\det |V + T^2|) + \log(\det |X'(V + T^2)^{-1}X|) + (y - X\hat{\alpha})'(V + T^2)^{-1}(y - X\hat{\alpha})$$

where V is the known conditional sampling covariance matrix of y , T^2 is the variance component of the random effects, and $\hat{\alpha} = (X'(V + T^2)^{-1}X)^{-1}X'(V + T^2)^{-1}y$. `reml()` minimizes the above likelihood function to obtain the parameter estimates.

Value

An object of class `reml` with a list of

<code>call</code>	Object returned by <code>match.call</code>
<code>data</code>	A data matrix of y , v and x
<code>no.y</code>	Number of effect sizes
<code>no.x</code>	Number of predictors
<code>miss.vec</code>	A vector indicating missing data. Studies will be removed before the analysis if they are TRUE
<code>mx.fit</code>	A fitted object returned from <code>mxRun</code>

Note

`reml` is more computationally intensive than `meta`. Moreover, `reml` is more likely to encounter errors during optimization. Since a likelihood function is directly employed to obtain the parameter estimates, there is no number of studies and number of observed statistics returned by `mxRun`. Ad hoc steps are used to modify `mx.fit@runstate$objectives[[1]]@numObs` and `mx.fit@runstate$objectives[[1]]@num`

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2013). Implementing restricted maximum likelihood estimation in structural equation models. *Structural Equation Modeling*, **20**(1), 157-167.
- Mehta, P. D., & Neale, M. C. (2005). People Are Variables Too: Multilevel Structural Equations Modeling. *Psychological Methods*, **10**(3), 259-284.
- Searle, S. R., Casella, G., & McCulloch, C. E. (1992). *Variance components*. New York: Wiley.
- Viechtbauer, W. (2005). Bias and efficiency of meta-analytic variance estimators in the random-effects model. *Journal of Educational and Behavioral Statistics*, **30**(3), 261-293.

See Also

[meta](#), [reml3](#), [Hox02](#), [Berkey98](#)

reml3L	<i>Estimate Variance Components in Three-Level Univariate Meta-Analysis with Restricted (Residual) Maximum Likelihood Estimation</i>
--------	--

Description

It estimates the variance components of random-effects in three-level univariate meta-analysis with restricted (residual) maximum likelihood (REML) estimation method.

Usage

```
reml3L(
  y,
  v,
  cluster,
  x,
  data,
  RE2.startvalue = 0.1,
  RE2.lbound = 1e-10,
  RE3.startvalue = RE2.startvalue,
  RE3.lbound = RE2.lbound,
  RE.equal = FALSE,
  intervals.type = c("z", "LB"),
  model.name = "Variance component with REML",
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)
```

Arguments

<code>y</code>	A vector of k studies of effect size.
<code>v</code>	A vector of k studies of sampling variance.
<code>cluster</code>	A vector of k characters or numbers indicating the clusters.
<code>x</code>	A predictor or a $k \times m$ matrix of level-2 and level-3 predictors where m is the number of predictors.
<code>data</code>	An optional data frame containing the variables in the model.
<code>RE2.startvalue</code>	Starting value for the level-2 variance.
<code>RE2.lbound</code>	Lower bound for the level-2 variance.
<code>RE3.startvalue</code>	Starting value for the level-3 variance.
<code>RE3.lbound</code>	Lower bound for the level-3 variance.
<code>RE.equal</code>	Logical. Whether the variance components at level-2 and level-3 are constrained equally.
<code>intervals.type</code>	Either <code>z</code> (default if missing) or <code>LB</code> . If it is <code>z</code> , it calculates the 95% Wald confidence intervals (CIs) based on the z statistic. If it is <code>LB</code> , it calculates the 95% likelihood-based CIs on the parameter estimates. Note that the z values and their associated p values are based on the z statistic. They are not related to the likelihood-based CIs.
<code>model.name</code>	A string for the model name in <code>mxModel</code> .
<code>suppressWarnings</code>	Logical. If <code>TRUE</code> , warnings are suppressed. It is passed to <code>mxRun</code> .
<code>silent</code>	Logical. Argument to be passed to <code>mxRun</code>
<code>run</code>	Logical. If <code>FALSE</code> , only return the <code>mx</code> model without running the analysis.
<code>...</code>	Further arguments to be passed to <code>mxRun</code>

Details

Restricted (residual) maximum likelihood obtains the parameter estimates on the transformed data that do not include the fixed-effects parameters. A transformation matrix $M = I - X(X'X)^{-1}X'$ is created based on the design matrix X which is just a column vector when there is no predictor in x . The last N redundant rows of M is removed where N is the rank of X . After pre-multiplying by M on y , the parameters of fixed-effects are removed from the model. Thus, only the parameters of random-effects are estimated.

An alternative but equivalent approach is to minimize the $-2 \times \log$ -likelihood function:

$$\log(\det |V + T^2|) + \log(\det |X'(V + T^2)^{-1}X|) + (y - X\hat{\alpha})'(V + T^2)^{-1}(y - X\hat{\alpha})$$

where V is the known conditional sampling covariance matrix of y , T^2 is the variance component combining level-2 and level-3 random effects, and $\hat{\alpha} = (X'(V + T^2)^{-1}X)^{-1}X'(V + T^2)^{-1}y$. `reml()` minimizes the above likelihood function to obtain the parameter estimates.

Value

An object of class `reml` with a list of

<code>call</code>	Object returned by match.call
<code>data</code>	A data matrix of y, v, and x
<code>mx.fit</code>	A fitted object returned from mxRun

Note

`reml` is more computationally intensive than `meta`. Moreover, `reml` is more likely to encounter errors during optimization. Since a likelihood function is directly employed to obtain the parameter estimates, there is no number of studies and number of observed statistics returned by [mxRun](#). Ad hoc steps are used to modify `mx.fit@runstate$objectives[[1]]@numObs` and `mx.fit@runstate$objectives[[1]]@num`

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Cheung, M. W.-L. (2013). Implementing restricted maximum likelihood estimation in structural equation models. *Structural Equation Modeling*, **20**(1), 157-167.

Cheung, M. W.-L. (2014). Modeling dependent effect sizes with three-level meta-analyses: A structural equation modeling approach. *Psychological Methods*, **19**, 211-229.

Mehta, P. D., & Neale, M. C. (2005). People Are Variables Too: Multilevel Structural Equations Modeling. *Psychological Methods*, **10**(3), 259-284.

Searle, S. R., Casella, G., & McCulloch, C. E. (1992). *Variance components*. New York: Wiley.

See Also

[meta3L](#), [reml](#), [Cooper03](#), [Bornmann07](#)

rerun	<i>Rerun models via <code>mxTryHard()</code></i>
-------	--

Description

It reruns models via `mxTryHard()`.

Usage

```
rerun(object, autofixtau2 = FALSE, extraTries = 10, ...)
```

Arguments

<code>object</code>	An object of either class <code>tssem1FEM</code> , class <code>tssem1REM</code> , class <code>wls</code> , class <code>meta</code> , class <code>reml</code> , class <code>osmasem</code> , class <code>osmasem3L</code> , and class <code>MxModel</code> .
<code>autofixtau2</code>	Logical. Whether automatically fixes elements of <code>tau2</code> with NA of standard errors. It only works for objects of class <code>tssem1REM</code> , class <code>meta</code> , and class <code>osmasem</code> .
<code>extraTries</code>	The number of attempts to run the model in addition to the first.
<code>...</code>	Further arguments to be passed to mxTryHard

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

Examples

```
random1 <- tssem1(Digman97$data, Digman97$n, method="REM", RE.type="Diag")
random1_rerun <- rerun(random1)
summary(random1_rerun)
```

Roorda11	<i>Studies on Students' School Engagement and Achievement Reported by Roorda et al. (2011)</i>
----------	--

Description

The data set includes 45 studies on the influence of affective teacher-student relationships on students' school engagement and achievement reported by Roorda et al. (2011).

Details

The variables are:

data A list of 45 studies of correlation matrices. The variables are *pos* (positive teacher-student relations), *neg* (negative teacher-student relations), *enga* (student engagement), and *achiev* (student achievement).

n A vector of sample sizes

SES A vector of average socio-economic status (SES) of the samples

Source

Roorda, D. L., Koomen, H. M. Y., Spilt, J. L., & Oort, F. J. (2011). The influence of affective teacher-student relationships on students' school engagement and achievement a meta-analytic approach. *Review of Educational Research*, 81(4), 493-529.

References

Jak, S., & Cheung, M. W.-L. (2018). Addressing heterogeneity in meta-analytic structural equation modeling using subgroup analysis. *Behavior Research Methods*, **50**, 1359-1373.

Examples

```
## Random-effects model: First stage analysis
random1 <- tssem1(Cov = Roorda11$data, n = Roorda11$n, method = "REM",
  RE.type = "Diag")
summary(random1)

## Model specification in lavaan syntax
model <- "## Regression paths
  enga ~ b31*pos + b32*neg
  achiev ~ b43*enga
  ## Variances of pos and neg are fixed at 1
  pos ~~ 1*pos
  neg ~~ 1*neg
  ## Correlation between pos and neg
  pos ~~ p21*neg
  ## Error variances
  enga ~~ p33*enga
  achiev ~~ p44*achiev"

RAM <- lavaan2RAM(model, obs.variables=c("pos", "neg", "enga", "achiev"))
RAM

## ## Equivalent RAM specification using create.mxMatrix()
## varnames <- c("pos", "neg", "enga", "achiev")
## A <- create.mxMatrix(c(0,0,0,0,
##                        0,0,0,0,
##                        "0.1*b31", "0.1*b32", 0,0,
##                        0,0,"0.1*b43",0),
##                        type = "Full", nrow = 4, ncol = 4, byrow = TRUE,
##                        name = "A", as.mxMatrix = FALSE)
## dimnames(A) <- list(varnames, varnames)
## A
##
## S <- create.mxMatrix(c(1,
##                        ".5*p21", 1,
##                        0,0,"0.6*p33",
##                        0,0,0,"0.6*p44"),
##                        type="Symm", byrow = TRUE,
##                        name="S", as.mxMatrix = FALSE)
## dimnames(S) <- list(varnames, varnames)
## S

## Random-effects model: Second stage analysis
random2 <- tssem2(random1, RAM=RAM, diag.constraints=TRUE,
  intervals="LB")
```

```
summary(random2)

## Display the model with the parameter estimates
plot(random2)
```

Scalco17*Correlation Matrices from Scalco et al. (2017)*

Description

The data set includes correlation matrices using the theory of planned behavior to predict organic food consumption reported by Scalco17 et al. (2017).

Details

A list of data with the following structure:

data A list of correlation matrices. The variables are *ATT* (attitude), *SN* (subjective norm), *PBC* (perceived behavior control), *BI* (behavioral intention), and *BEH* (behavior)

n A vector of sample sizes

Age A vector of the mean age of the samples

Female A vector of the percentage of the female samples

Source

Scalco, A., Noventa, S., Sartori, R., & Ceschi, A. (2017). Predicting organic food consumption: A meta-analytic structural equation model based on the theory of planned behavior. *Appetite*, **112**, 235-248.

Examples

```
data(Scalco17)
```

sem

*Fit a structural equation model using OpenMx***Description**

It fits a structural equation model by creating a `mxModel` from a RAM object.

Usage

```
sem(
  model.name = "sem",
  RAM = NULL,
  data = NULL,
  Cov = NULL,
  means = NULL,
  numObs,
  intervals.type = c("z", "LB"),
  startvalues = NULL,
  lbound = NULL,
  ubound = NULL,
  replace.constraints = FALSE,
  mxModel.Args = NULL,
  run = TRUE,
  silent = TRUE,
  group = NULL,
  cluster = NULL,
  ...
)
```

Arguments

<code>model.name</code>	A string for the model name in mxModel .
<code>RAM</code>	A RAM object including a list of matrices of the model returned from lavaan2RAM .
<code>data</code>	A data frame or matrix of data.
<code>Cov</code>	A covariance matrix may also be used if <code>data==NULL</code> .
<code>means</code>	A named vector of means (optional) if <code>Cov</code> is used.
<code>numObs</code>	If <code>Cov</code> is used, a sample size must be provided.
<code>intervals.type</code>	Either <code>z</code> (default if missing) or <code>LB</code> . If it is <code>z</code> , it calculates the 95% confidence intervals (CIs) based on the estimated standard error. If it is <code>LB</code> , it calculates the 95% likelihood-based CIs on the parameter estimates.
<code>startvalues</code>	A list of named starting values of the free parameters, e.g., <code>list(a=1, b=2)</code>
<code>lbound</code>	A list of lower bound of the free parameters. If it is not provided, all free parameters are assumed NA.
<code>ubound</code>	A list of upper bound of the free parameters. If it is not provided, all free parameters are assumed NA.

<code>replace.constraints</code>	Logical. If TRUE, the parameters on the left hand side will be replaced by the constraints on the right hand side. That is, the parameters on the left hand side are no longer parameters in the model.
<code>mxModel.Args</code>	A list of arguments passed to <code>mxModel</code> .
<code>run</code>	Logical. If FALSE, only return the mx model without running the analysis.
<code>silent</code>	Logical. An argument is passed to either <code>mxRun</code> or <code>mxTryHard</code>
<code>group</code>	A character string naming the column in data identifying group membership. Required (and only used) when RAM is a multiple-group RAM object, i.e., created by <code>lavaan2RAM</code> with <code>ngroups > 1</code> . <code>data</code> must be a single data frame in this case; it is split internally by the unique values of <code>data[[group]]</code> in <i>first-appearance order in the data</i> (not sorted, and not a factor's own <code>levels()</code> order, even for a factor column – this matches how lavaan itself numbers groups), which must correspond, in order, to the groups used when the model was specified (e.g., lavaan's <code>c(...)</code> syntax numbers groups the same way). A group value that is not itself a legal <code>mxModel</code> name (e.g. purely numeric, or containing a character such as <code>.</code> or <code>-</code>) is sanitized into one internally; the resulting submodel names are available via <code>names(fit\$mx.fit@submodels)</code> , and a mapping back to the original data values via <code>fit\$group.map</code> , so <code>plot.mxsem</code> 's own <code>group</code> argument can still be called with the original, unsanitized group value. An unused factor level in <code>data[[group]]</code> (one that never actually occurs in the data) is simply never treated as a group in the first place (this is what first-appearance order, above, naturally does); if that leaves fewer groups than RAM expects, it is reported as a group-count mismatch. <code>Cov/numObs</code> -based input is not yet supported for multiple-group models. <code>replace.constraints=TRUE</code> IS supported (substituting a constraint into whichever group's A/S/ M matrices contain it, including a constraint that spans multiple groups, e.g. constraining one group's variance to be a multiple of another's); a definition variable (<code>data.*</code>) inside such a constraint resolves against whichever group's own data it is used in, since (unlike two-level's between level) every group already carries its own full data slice.
<code>cluster</code>	A character string naming the column in data identifying level-2 (cluster) membership. Required (and only used) when RAM is a two-level RAM object, i.e., created by <code>lavaan2RAM</code> from lavaan's two-level (<code>level: 1 / level: 2</code>) syntax. <code>data</code> must be a single data frame of individual-level (level-1) rows in this case. <code>Cov/numObs</code> -based input is not yet supported for two-level models, and neither is combining two-level and multiple-group models in the same call. <code>replace.constraints=TRUE</code> IS supported for two-level models (substituting a constraint into whichever of the within-/between-level A/S/M matrices contains it, including a constraint that spans both levels); a definition variable (<code>data.*</code>) may be used in such a constraint for a level-1 (within) parameter, but not yet for a genuine level-2-only (between, cluster-level) covariate, since data passed to the level-2 submodel currently carries only the cluster ID column.
<code>...</code>	Further arguments will be passed to either <code>mxRun</code> or <code>mxTryHard</code>

Value

An object of class `mxsem`

Author(s)

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Examples

```
## Generate data
set.seed(100)
n <- 100
x <- rnorm(n)
y <- 0.5*x + rnorm(n, mean=0, sd=sqrt(1-0.5^2))
my.df <- data.frame(y=y, x=x)

## A regression model
model <- "y ~ x    # Regress y on x
        y ~ 1    # Intercept of y
        x ~ 1    # Mean of x"
plot(model)

RAM <- lavaan2RAM(model, obs.variables=c("y", "x"))

my.fit <- sem(RAM=RAM, data=my.df)
summary(my.fit)

## A meta-analysis
model <- "f =~ 1*yi
        f ~ mu*1      ## Average effect
        f ~~ tau2*f   ## Heterogeneity variance
        yi ~~ data.vi*yi ## Known sampling variance"
plot(model)

## Do not standardize the latent variable (f): std.lv=FALSE
RAM <- lavaan2RAM(model, obs.variables="yi", std.lv=FALSE)

## Use likelihood-based CI
my.fit <- sem(RAM=RAM, data=Hox02, intervals="LB")
summary(my.fit)

## A 2-group regression with a shared slope but group-specific intercepts
x1 <- rnorm(100); y1 <- 2 + 0.5*x1 + rnorm(100)
x2 <- rnorm(100); y2 <- -1 + 0.5*x2 + rnorm(100)
my.df2 <- data.frame(y=c(y1, y2), x=c(x1, x2),
                    g=rep(c("g1", "g2"), each=100))

model <- "y ~ c(b,b)*x + c(a1,a2)*1"
RAM <- lavaan2RAM(model, ngroups=2)
my.fit <- sem(RAM=RAM, data=my.df2, group="g")
summary(my.fit)

## The same 2-group regression, extended with replace.constraints=TRUE:
## group 2's residual variance is constrained to be exactly TWICE group
## 1's, and group 1's own residual variance is reparameterised through
```

```

## exp() (guaranteeing positivity). "resid1" never appears as a free
## parameter itself -- lavaanify() would otherwise reject "a0" as
## undeclared, and an ordinary (non-replaced) "==" constraint cannot
## introduce a new free parameter this way; only replace.constraints=
## TRUE can. The constraint chain is resolved across groups, so
## "resid2" ends up expressed directly in terms of "a0" (not "resid1").
model <- "y ~ c(b,b)*x + c(a1,a2)*1
        y ~~ c(resid1,resid2)*y
        resid1 == exp(a0)
        resid2 == 2*resid1"
RAM <- lavaan2RAM(model, ngroups=2)
my.fit <- sem(RAM=RAM, data=my.df2, group="g", replace.constraints=TRUE)
summary(my.fit)

## A two-level (random-intercept) regression model
set.seed(123)
nClusters <- 60; nPerCluster <- 6
clusterID <- rep(seq_len(nClusters), each=nPerCluster)
u <- rep(rnorm(nClusters, 0, 0.7), each=nPerCluster)
x <- rnorm(nClusters*nPerCluster)
y <- 1.5 + 0.6*x + u + rnorm(nClusters*nPerCluster)
my.df3 <- data.frame(clusterID=clusterID, x=x, y=y)

model <- "level: 1
        y ~ b1*x
        y ~~ residW*y
        level: 2
        y ~ 1
        y ~~ residB*y"
RAM <- lavaan2RAM(model)
my.fit <- sem(RAM=RAM, data=my.df3, cluster="clusterID")
summary(my.fit)

## A 2-group CFA: test whether factor loadings are invariant across groups
set.seed(103)
simGroup <- function(n, loadings) {
  f <- rnorm(n)
  Y <- sapply(loadings, function(l) l*f + rnorm(n, sd=0.5))
  colnames(Y) <- paste0("y", seq_along(loadings))
  data.frame(Y)
}
loadings <- c(1, 0.8, 1.3, 0.9)
my.df4 <- rbind(cbind(simGroup(250, loadings), g="g1"),
               cbind(simGroup(250, loadings), g="g2"))

## Metric invariance: loadings constrained equal across groups.
## std.lv=FALSE: the factor's scale is already identified by fixing
## y1's loading at 1 (c(1,1)*y1), so its variance ("f ~~ c(vf1,vf2)*f")
## is left free and group-specific instead of also being fixed at 1 --
## lavaan2RAM()'s default std.lv=TRUE would otherwise silently discard
## that "f ~~ ..." line, fixing the variance at 1 in both groups with
## no error (see lavaan2RAM()'s "std.lv" argument for details).
model.invariant <- "f =~ c(1,1)*y1 + c(l2,l2)*y2 + c(l3,l3)*y3 + c(l4,l4)*y4

```

```

      f ~~ c(vf1,vf2)*f
      y1 ~ c(m1a,m1b)*1
      y2 ~ c(m2a,m2b)*1
      y3 ~ c(m3a,m3b)*1
      y4 ~ c(m4a,m4b)*1"
fit.invariant <- sem(RAM=lavaan2RAM(model.invariant, ngroups=2, std.lv=FALSE),
  data=my.df4, group="g")

## Configural model: loadings free in each group
model.free <- "f =~ c(1,1)*y1 + c(l2a,l2b)*y2 + c(l3a,l3b)*y3 + c(l4a,l4b)*y4
  f ~~ c(vf1,vf2)*f
  y1 ~ c(m1a,m1b)*1
  y2 ~ c(m2a,m2b)*1
  y3 ~ c(m3a,m3b)*1
  y4 ~ c(m4a,m4b)*1"
fit.free <- sem(RAM=lavaan2RAM(model.free, ngroups=2, std.lv=FALSE),
  data=my.df4, group="g")

summary(fit.invariant)
anova(fit.free, fit.invariant) # likelihood-ratio test of metric invariance

## Group difference in the factor variance, as a derived quantity
model.diff <- paste(model.free, "\ndiff_v := vf2 - vf1")
fit.diff <- sem(RAM=lavaan2RAM(model.diff, ngroups=2, std.lv=FALSE),
  data=my.df4, group="g")
summary(fit.diff) # see "Mxalgebras" in the printed output for diff_v

## A two-level CFA: a factor at the within level and another at the
## between level, with the same three indicators loading on both
set.seed(104)
nClusters <- 100; nPerCluster <- 8; N <- nClusters*nPerCluster
clusterID <- rep(seq_len(nClusters), each=nPerCluster)
fb <- rep(rnorm(nClusters, 0, 1), each=nPerCluster)
fw <- rnorm(N, 0, 1)
ly2 <- 0.8; ly3 <- 1.2
y1 <- fb + fw + rnorm(N, 0, 0.5)
y2 <- ly2*fb + ly2*fw + rnorm(N, 0, 0.5)
y3 <- ly3*fb + ly3*fw + rnorm(N, 0, 0.5)
my.df5 <- data.frame(clusterID=clusterID, y1=y1, y2=y2, y3=y3)

model <- "level: 1
  fw =~ y1 + y2 + y3
level: 2
  fb =~ y1 + y2 + y3"
RAM <- lavaan2RAM(model)
my.fit <- sem(RAM=RAM, data=my.df5, cluster="clusterID")
summary(my.fit)

## A two-level (multilevel) meta-analysis: studies contributing several
## dependent effect sizes each, separating within- from between-study
## heterogeneity while accounting for each effect size's own known
## sampling variance
set.seed(106)

```

```

nStudies <- 60
esPerStudy <- sample(2:4, nStudies, replace=TRUE)
studyID <- rep(seq_len(nStudies), esPerStudy)
N <- length(studyID)
mu <- 0.35; tau2.between <- 0.03; tau2.within <- 0.015
studyEffect <- rep(rnorm(nStudies, mu, sqrt(tau2.between)), esPerStudy)
vi <- runif(N, 0.02, 0.06)
yi <- rnorm(N, studyEffect + rnorm(N, 0, sqrt(tau2.within)), sqrt(vi))
my.df6 <- data.frame(studyID=studyID, yi=yi, vi=vi)

## fw's own variance is the WITHIN-study heterogeneity; yi's own residual
## (after removing fw) is fixed at the KNOWN sampling variance; the
## BETWEEN-study heterogeneity is yi's own variance at level 2
model <- "level: 1
        fw =~ 1*yi
        fw ~~ tauW*fw
        yi ~~ data.vi*yi
level: 2
        yi ~ mu*1
        yi ~~ tauB*yi"
RAM <- lavaan2RAM(model, std.lv=FALSE)
my.fit <- sem(RAM=RAM, data=my.df6, cluster="studyID")
summary(my.fit)

## A two-level location-scale meta-analysis: replace.constraints=TRUE
## lets the WITHIN-study heterogeneity (tauW, on fw) be a function of an
## effect-size-level moderator xi, while yi's own residual stays fixed
## at its KNOWN sampling variance (data.vi), exactly as in the previous
## example. data.xi is a definition variable, so its per-row value is
## looked up from the data instead of estimated. "tauW" never appears
## as a free parameter itself here -- lavaanify() would otherwise reject
## a0/a1 as undeclared, and an ordinary (non-replaced) "==" constraint
## cannot introduce new free parameters this way; only
## replace.constraints=TRUE can.
set.seed(107)
xi <- rnorm(N)
a0 <- -4; a1 <- 0.8
tauWi <- exp(a0 + a1*xi)
yi2 <- rnorm(N, studyEffect + rnorm(N, 0, sqrt(tauWi)), sqrt(vi))
my.df7 <- data.frame(studyID=studyID, yi=yi2, vi=vi, xi=xi)

model <- "level: 1
        fw =~ 1*yi
        fw ~~ tauW*fw
        yi ~~ data.vi*yi
level: 2
        yi ~ mu*1
        yi ~~ tauB*yi
        tauW == exp(a0 + a1*data.xi)"
RAM <- lavaan2RAM(model, std.lv=FALSE)
my.fit <- sem(RAM=RAM, data=my.df7, cluster="studyID",
              replace.constraints=TRUE)
summary(my.fit)

```

smdMES

*Compute Effect Sizes for Multiple End-point Studies***Description**

It computes the standardized mean differences and their asymptotic sampling covariance matrix for two multiple end-point studies with p effect sizes.

Usage

```
smdMES(
  m1,
  m2,
  V1,
  V2,
  n1,
  n2,
  homogeneity = c("covariance", "correlation", "none"),
  bias.adjust = TRUE,
  list.output = TRUE,
  lavaan.output = FALSE
)
```

Arguments

m1	A vector of p sample means of the first group.
m2	A vector of p sample means of the second group.
V1	A p by p sample covariance matrix of the first group.
V2	A p by p sample covariance matrix of the second group.
n1	The sample size of the first group.
n2	The sample size of the second group.
homogeneity	If it is covariance (the default), homogeneity of covariance matrices is assumed. The common standard deviations are used as the standardizers in calculating the effect sizes. If it is correlation, homogeneity of correlation is not assumed. The standard deviations of the first group are used as the standardizer in calculating the effect sizes. If it is none, no homogeneity assumption is made. The standard deviations of the first group are used as the standardizer in calculating the effect sizes.
bias.adjust	If it is TRUE (the default), the effect sizes are adjusted for small bias by multiplying $1 - 3/(4 * (n1 + n2) - 9)$.
list.output	If it is TRUE (the default), the effect sizes and their sampling covariance matrix are outputted as a list. If it is FALSE, they will be stacked into a vector.
lavaan.output	If it is FALSE (the default), the effect sizes and its sampling covariance matrix are reported. If it is TRUE, it outputs the fitted <code>lavaan-class</code> object.

Details

Gleser and Olkin (2009) introduce formulas to calculate the standardized mean differences and their sampling covariance matrix for multiple end-point studies under the assumption of homogeneity of the covariance matrix. This function uses a structural equation modeling (SEM) approach introduced in Chapter 3 of Cheung (2015) to calculate the same estimates. The SEM approach is more flexible in two ways: (1) it allows homogeneity of covariance or correlation matrices or not; and (2) it allows users to test this assumption by checking the fitted `lavaan-class` object.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2015). *Meta-analysis: A structural equation modeling approach*. Chichester, West Sussex: John Wiley & Sons, Inc.
- Cheung, M. W.-L. (2018). Computing multivariate effect sizes and their sampling covariance matrices with structural equation modeling: Theory, examples, and computer simulations. *Frontiers in Psychology*, *9*(1387). <https://doi.org/10.3389/fpsyg.2018.01387>
- Gleser, L. J., & Olkin, I. (2009). Stochastically dependent effect sizes. In H. Cooper, L. V. Hedges, & J. C. Valentine (Eds.), *The handbook of research synthesis and meta-analysis*. (2nd ed., pp. 357-376). New York: Russell Sage Foundation.

See Also

[Gleser94](#), [smdMTS](#), [calEffSizes](#)

Examples

```
## Sample means for the two constructs in Group 1
m1 <- c(2.5, 4.5)

## Sample means for the two constructs in Group 2
m2 <- c(3, 5)

## Sample covariance matrix in Group 1
V1 <- matrix(c(3,2,2,3), ncol=2)

## Sample covariance matrix in Group 2
V2 <- matrix(c(3.5,2.1,2.1,3.5), ncol=2)

## Sample size in Group 1
n1 <- 20

## Sample size in Group 2
n2 <- 25

## SMD with the assumption of homogeneity of covariance matrix
smdMES(m1, m2, V1, V2, n1, n2, homogeneity="cov", bias.adjust=TRUE,
```

```

lavaan.output=FALSE)

## SMD with the assumption of homogeneity of correlation matrix
smdMES(m1, m2, V1, V2, n1, n2, homogeneity="cor", bias.adjust=TRUE,
lavaan.output=FALSE)

## SMD without any assumption of homogeneity
smdMES(m1, m2, V1, V2, n1, n2, homogeneity="none", bias.adjust=TRUE,
lavaan.output=FALSE)

## Output the fitted lavaan model
## It provides a likelihood ratio test to test the null hypothesis of
## homogeneity of variances.
fit <- smdMES(m1, m2, V1, V2, n1, n2, homogeneity="cov", bias.adjust=TRUE,
lavaan.output=TRUE)

lavaan::summary(fit)

lavaan::parameterestimates(fit)

```

smdMTS

Compute Effect Sizes for Multiple Treatment Studies

Description

It computes the standardized mean differences and their asymptotic sampling covariance matrix for k multiple treatment studies. The first group is assumed as the control group.

Usage

```

smdMTS(
  m,
  v,
  n,
  homogeneity = c("variance", "none"),
  bias.adjust = TRUE,
  all.comparisons = FALSE,
  list.output = TRUE,
  lavaan.output = FALSE
)

```

Arguments

<code>m</code>	A vector of k sample means.
<code>v</code>	A vector of k sample variances.
<code>n</code>	A vector of k sample sizes.

homogeneity	If it is variance (the default), homogeneity of variances is assumed. The common standard deviation is used as the standardizer in calculating the effect sizes. If it is none, homogeneity of variances is not assumed. The standard deviation of the first group is used as the standardizer in calculating the effect sizes.
bias.adjust	If it is TRUE (the default), the effect sizes are adjusted for small bias by multiplying $1 - 3/(4 * (n1 + n2) - 9)$.
all.comparisons	If it is FALSE (the default), all groups (except the first group) are compared against the first group. If it is TRUE, all pairwise comparisons are calculated. This may be useful in network meta-analysis.
list.output	If it is TRUE (the default), the effect sizes and their sampling covariance matrix are outputted as a list. If it is FALSE, they will be stacked into a vector.
lavaan.output	If it is FALSE (the default), the effect sizes and its sampling covariance matrix are reported. If it is TRUE, it outputs the fitted <code>lavaan-class</code> object.

Details

Gleser and Olkin (2009) introduce formulas to calculate the standardized mean differences and their sampling covariance matrix for multiple treatment studies under the assumption of homogeneity of the covariance matrix. This function uses a structural equation modeling (SEM) approach introduced in Chapter 3 of Cheung (2015) to calculate the same estimates. The SEM approach is more flexible in three ways: (1) it allows homogeneity of variances or not; (2) it allows users to test the assumption of homogeneity of variances by checking the fitted `lavaan-class` object; and (3) it may calculate all pairwise comparisons.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2015). *Meta-analysis: A structural equation modeling approach*. Chichester, West Sussex: John Wiley & Sons, Inc.
- Cheung, M. W.-L. (2018). Computing multivariate effect sizes and their sampling covariance matrices with structural equation modeling: Theory, examples, and computer simulations. *Frontiers in Psychology*, *9*(1387). <https://doi.org/10.3389/fpsyg.2018.01387>
- Gleser, L. J., & Olkin, I. (2009). Stochastically dependent effect sizes. In H. Cooper, L. V. Hedges, & J. C. Valentine (Eds.), *The handbook of research synthesis and meta-analysis*. (2nd ed., pp. 357-376). New York: Russell Sage Foundation.

See Also

[Gleser94](#), [smdMES](#), [calEffSizes](#)

Examples

```
## Sample means for groups 1 to 3
m <- c(5,7,9)

## Sample variances
v <- c(10,11,12)

## Sample sizes
n <- c(50,52,53)

## Assuming homogeneity of variances
smdMTS(m, v, n, homogeneity = "var", bias.adjust=TRUE, all.comparisons=FALSE,
        lavaan.output=FALSE)

## Not assuming homogeneity of variances and comparing all pairwise groups
## Please note that the SD of the first group is used as the standardizer
smdMTS(m, v, n, homogeneity = "none", bias.adjust=TRUE, all.comparisons=TRUE,
        lavaan.output=FALSE)

## Output the fitted lavaan model
## It provides a likelihood ratio test to test the null hypothesis of
## homogeneity of variances.
fit <- smdMTS(m, v, n, homogeneity = "var", bias.adjust=FALSE, all.comparisons=FALSE,
              lavaan.output=TRUE)

lavaan::summary(fit)

lavaan::parameterestimates(fit)
```

 Stadler15

Correlations from Stadler et al. (2015)

Description

The data set includes correlations between complex problem solving and intelligence reported by Stadler et al. (2015).

Details

A list of data with the following structure:

ID ID of the effect sizes

Authors Authors of the studies

Year Year of the studies

N Sample size

CPSMeasure Complex problem solving (CPS) measure

IntelligenceMeasure Intelligence measure

r Correlation between CPS and intelligence

v Sampling variance of r

Source

Stadler, M., Becker, N., Godker, M., Leutner, D., & Greiff, S. (2015). Complex problem solving and intelligence: A meta-analysis. *Intelligence*, **53**, 92-101.

summary

Summary Method for tssem1, wls, meta, and meta3LFIML Objects

Description

It summaries results for various class.

Arguments

object	An object returned from either class <code>tssem1FEM</code> , class <code>tssem1FEM.cluster</code> , class <code>tssem1REM</code> , class <code>wls</code> , class <code>wls.cluster</code> , class <code>meta</code> , class <code>meta3LFIML</code> , class <code>rem1</code> , class <code>mxsem</code> or class <code>CorPop</code> .
x	An object returned from either class <code>summary.tssem1FEM</code> , class <code>summary.tssem1FEM.cluster</code> , class <code>summary.wls</code> , class <code>summary.meta</code> , class <code>summary.meta3LFIML</code> , class <code>summary.rem1</code> or class <code>summary.CorPop</code> .
homoStat	Logical. Whether to conduct a homogeneity test on the effect sizes.
allX	Logical. Whether to report the predictors and the auxiliary variables.
robust	Logical. Whether to use robust standard error from imxRobustSE .
df.adjustment	Numeric. Adjust the degrees of freedom manually. It may be necessary if the df calculated is incorrect when <code>diag.constraints=TRUE</code> .
probs	Quantiles for the parameter estimates.
cutoff.chisq.pvalue	Cutoff of the p-value for the chi-square statistic.
cutoff.CFI	The cutoff of the CFI.
cutoff.SRM	The cutoff of the SRMR.
cutoff.RMSEA	The cutoff of the RMSEA.
fitIndices	Whether to calculate the chi-square statistic and various goodness-of-fit indices in <code>osmasem</code> . Note. It may take a while since statistics of the saturated and independence models are required.
numObs	The number of observations in calculating the fit statistics in <code>osmasem</code> . If it is missing, the total number of observations is used.
...	Further arguments to be passed to printCoefmat

Note

If the OpenMx status1 is either 0 or 1, the estimation is considered fine. If the OpenMx status1 is other values, it indicates estimation problems. Users should refer to 'OpenMx' website for more details.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[tssem1](#), [wls](#), [meta](#), [reml](#), [rCor](#), [bootuniR2](#), [osmasem](#)

Tenenbaum02

Correlation coefficients reported by Tenenbaum and Leaper (2002)

Description

Forty-eight studies reported by Tenenbaum and Leaper (2002, Table 1).

Details

The variables are:

Authors Authors of the study

Year Year of publication

N Sample size

r Correlation between parents' gender schemas and their offspring's gender-related cognitions.

v Sampling variance of r

Publication_source Publication source: 1="top-tier journal", 2="second-tier journal or book chapter", 3="dissertation", 4="other unpublished study"

Author_gender Gender of the first author: "W"="woman", "M"="man"

Parent_type Parent type: "M"="mother", "F"="father", "MF"="mother and father"

Parent_predictor Parent predictor: "S"="self gender schema", "A"="gender attitudes about others"

Offspring_age Offspring age (months)

Offspring_type Offspring type: "D"="daughter", "S"="son", "DS"="daughter and son"

Offspring_outcome Offspring outcome: "S"="gender schema for self", "A"="gender attitudes toward others", "I"="gender-related interests and preferences", "W"="work-related attitudes"

Source

Tenenbaum, H. R., & Leaper, C. (2002). Are parents' gender schemas related to their children's gender-related cognitions? A meta-analysis. *Developmental Psychology*, 38(4), 615-630. <https://doi.org/10.1037/0012-1649.38.4.615>

Examples

```
data(Tenenbaum02)
```

tssem1	<i>First Stage of the Two-Stage Structural Equation Modeling (TSSEM)</i>
--------	--

Description

It conducts the first stage analysis of TSSEM by pooling correlation/covariance matrices. `tssem1FEM()` and `tssem1REM()` use fixed- and random-effects models, respectively. `tssem1()` is a wrapper of these functions.

Usage

```
tssem1(  
  Cov,  
  n,  
  method = c("REM", "FEM"),  
  cor.analysis = TRUE,  
  cluster = NULL,  
  RE.type = c("Diag", "Symm", "Zero", "User"),  
  RE.startvalues = 0.1,  
  RE.lbound = 1e-10,  
  RE.constraints = NULL,  
  I2 = "I2q",  
  acov = c("weighted", "individual", "unweighted"),  
  asyCovOld = FALSE,  
  model.name = NULL,  
  suppressWarnings = TRUE,  
  silent = TRUE,  
  run = TRUE,  
  ...  
)
```

Arguments

Cov	A list of correlation/covariance matrices
n	A vector of sample sizes
method	Either "REM" (default if missing) or "FEM". If it is "REM", a random-effects meta-analysis will be applied. If it is "FEM", a fixed-effects meta-analysis will be applied.
cor.analysis	Logical. The output is either a pooled correlation or a covariance matrix.

cluster	A character vector in <code>tssem3L1</code> and <code>tssemRobust1</code> or a vector of characters or numbers indicating the clusters in <code>tssem1</code> . Analyses will be conducted for each cluster. It will be ignored when <code>method="REM"</code> .
RE.type	Either "Diag", "Symm", "Zero" or "User". If it is "Diag" (default if missing), a diagonal matrix is used for the random effects meaning that the random effects are independent. If it is "Symm", a symmetric matrix is used for the random effects on the covariances among the correlation (or covariance) vectors. If it is "Zero", there is no random effects which is similar to the conventional Generalized Least Squares (GLS) approach to fixed-effects analysis. If it is "User", the user has to specify the variance component via the <code>RE.constraints</code> argument. This argument will be ignored when <code>method="FEM"</code> .
RE.startvalues	Starting values on the diagonals of the variance component of the random effects. It will be ignored when <code>method="FEM"</code> .
RE.lbound	Lower bounds on the diagonals of the variance component of the random effects. It will be ignored when <code>method="FEM"</code> .
RE.constraints	A $p^* \times p^*$ matrix specifying the variance components of the random effects, where p^* is the number of effect sizes. If the input is not a matrix, it is converted into a matrix by <code>as.matrix()</code> . The default is that all covariance/variance components are free. The format of this matrix follows <code>as.mxMatrix</code> . Elements of the variance components can be constrained equally by using the same labels. If a zero matrix is specified, it becomes a fixed-effects meta-analysis.
I2	Possible options are "I2q", "I2hm" and "I2am". They represent the I2 calculated by using a typical within-study sampling variance from the Q statistic, the harmonic mean and the arithmetic mean of the within-study sampling variances (Xiong, Miller, & Morris, 2010). More than one options are possible. If <code>intervals.type="LB"</code> , 95% confidence intervals on the heterogeneity indices will be constructed.
acov	If it is individual, the sampling variance-covariance matrices are calculated based on individual correlation/covariance matrix. If it is either unweighted or weighted (the default), the average correlation/covariance matrix is calculated based on the unweighted or weighted mean with the sample sizes. The average correlation/covariance matrix is used to calculate the sampling variance-covariance matrices. This argument is ignored with the <code>method="FEM"</code> argument.
asyCovOld	Whether the old <code>asyCov</code> is used. See <code>asyCov</code> .
model.name	A string for the model name in <code>mxModel</code> .
suppressWarnings	Logical. If TRUE, warnings are suppressed. It is passed to <code>mxRun</code> .
silent	Logical. An argument to be passed to <code>mxRun</code>
run	Logical. If FALSE, only return the mx model without running the analysis.
...	Further arguments to be passed to <code>mxRun</code>

Value

Either an object of class `tssem1FEM` for fixed-effects TSSEM, an object of class `tssem1FEM.cluster` for fixed-effects TSSEM with `cluster` argument, or an object of class `tssem1REM` for random-effects TSSEM.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2014). Fixed- and random-effects meta-analytic structural equation modeling: Examples and analyses in R. *Behavior Research Methods*, **46**, 29-40.
- Cheung, M. W.-L. (2013). Multivariate meta-analysis as structural equation models. *Structural Equation Modeling*, **20**, 429-454.
- Cheung, M. W.-L., & Chan, W. (2005). Meta-analytic structural equation modeling: A two-stage approach. *Psychological Methods*, **10**, 40-64.
- Cheung, M. W.-L., & Chan, W. (2009). A two-stage approach to synthesizing covariance matrices in meta-analytic structural equation modeling. *Structural Equation Modeling*, **16**, 28-53.

See Also

[wls](#), [Cheung09](#), [Becker92](#), [Digman97](#), [issp89](#), [issp05](#)

tssemParaVar	<i>Estimate the heterogeneity (SD) of the parameter estimates of the TSSEM object</i>
--------------	---

Description

It estimates the heterogeneity of the parameter estimates of the TSSEM objects using either the bootstrap or the delta methods.

Usage

```
tssemParaVar(
  tssem1.obj,
  tssem2.obj,
  method = c("bootstrap", "delta"),
  interval = 0.8,
  Rep = 50,
  output = c("data.frame", "matrices"),
  nonPD.pop = c("replace", "nearPD", "accept")
)
```

Arguments

tssem1.obj	An object of class tssem1REM returned from tssem1()
tssem2.obj	An object of class wls returned from tssem2() or wls()
method	If it is bootstrap, random correlation matrices are sampled from the tssem1.obj by the parametric bootstrap. If it is delta, the delta method is used to estimate the heterogeneity of the parameter estimates.

interval	The desired interval, e.g., .8 or .95.
Rep	The number of parametric bootstrap. It is ignored when the method is delta.
output	Either a data.frame or matrices of the output.
nonPD.pop	If it is replace, generated non-positive definite matrices are replaced by generated new ones which are positive definite. If it is nearPD, they are replaced by nearly positive definite matrices by calling <code>Matrix::nearPD()</code> . If it is accept, they are accepted.

Details

The bootstrap method is based on the discussion in Cheung (2018) and Yu et al. (2016). The delta method is an alternative method to obtain the heterogeneity.

Value

Either a data.frame or matrices of the output.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

Cheung, M. W.-L. (2018). Issues in solving the problem of effect size heterogeneity in meta-analytic structural equation modeling: A commentary and simulation study on Yu, Downes, Carter, and O’Boyle (2016). *Journal of Applied Psychology*, **103**, 787-803.

Yu, J. (Joya), Downes, P. E., Carter, K. M., & O’Boyle, E. H. (2016). The problem of effect size heterogeneity in meta-analytic structural equation modeling. *Journal of Applied Psychology*, *101*, 1457-1473.

See Also

[bootuniR1](#), [bootuniR2](#), [Nohe15](#)

uniR1	<i>First Stage analysis of the univariate R (uniR) approach</i>
-------	---

Description

It conducts the first stage analysis of the uniR analysis by pooling elements of the correlation coefficients individually.

Usage

`uniR1(Cor, n, ...)`

Arguments

Cor	A list of correlation matrices
n	A vector of sample sizes
...	Further arguments which are currently ignored

Details

This function implements the univariate *r* approach proposed by Viswesvaran and Ones (1995) to conduct meta-analytic structural equation modeling (MASEM). It uses Schmidt and Hunter's approach to combine correlation coefficients. It is included in this package for research interests. The two-stage structural equation modeling (TSSEM) approach is preferred (e.g., Cheung, 2015; Cheung & Chan, 2005).

Value

An object of class `uniR1` of the original data, the sample sizes, the harmonic mean of sample sizes, the average correlation matrix, the standard errors of the correlation matrix, and the standard deviations (heterogeneity) of the correlation matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2015). *Meta-analysis: A structural equation modeling approach*. Chichester, West Sussex: John Wiley & Sons, Inc.
- Cheung, M. W.-L., & Chan, W. (2005). Meta-analytic structural equation modeling: A two-stage approach. *Psychological Methods*, **10**, 40-64.
- Schmidt, F. L., & Hunter, J. E. (2015). *Methods of meta-analysis: Correcting error and bias in research findings (3rd ed.)*. Thousand Oaks, CA: Sage.
- Viswesvaran, C., & Ones, D. S. (1995). Theory testing: Combining psychometric meta-analysis and structural equations modeling. *Personnel Psychology*, **48**, 865-885.

See Also

[uniR2](#), [Becker09](#)

uniR2

*Second Stage analysis of the univariate R (uniR) approach***Description**

It conducts the second stage analysis of the uniR analysis by fitting structural equation models on the average correlation matrix.

Arguments

x	An object of class uniR1 from uniR1 .
RAM	A RAM object including a list of matrices of the model returned from lavaan2RAM .
Amatrix	If RAM is not specified, an Amatrix is required. An asymmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Smatrix	If RAM is not specified, an Smatrix is required. A symmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Fmatrix	If RAM is not specified, an Fmatrix is required. A filter matrix in the RAM specification with MxMatrix-class . If it is NULL (the default), an identity matrix with the same dimensions of Cov will be created. If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function. It is not required when there is no latent variable.
model.name	A string for the model name in mxModel . If it is missing, the default is "UniR2".
suppressWarnings	Logical. If TRUE, warnings are suppressed. It is passed to mxRun .
silent	Logical. An argument to be passed to mxRun
run	Logical. If FALSE, only return the mx model without running the analysis.
model	A model specified using lavaan syntax see model.syntax
...	Further arguments to be passed to either mxRun or sem . For sem , fixed.x=FALSE is passed automatically.

Details

This function implements the univariate r approach proposed by Viswesvaran and Ones (1995) to conduct meta-analytic structural equation modeling (MASEM). It treats the average correlation matrix as if it was a covariance matrix in fitting structural equation models. The harmonic mean of the sample sizes in combining correlation coefficients is used as the sample size in fitting structural equation models. It is included in this package for research interests. The two-stage structural equation modeling (TSSEM) approach is preferred (e.g., Cheung, 2015; Cheung & Chan, 2005).

Value

A fitted object returned from [mxRun](#) or [sem](#).

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Cheung, M. W.-L. (2015). *Meta-analysis: A structural equation modeling approach*. Chichester, West Sussex: John Wiley & Sons, Inc.
- Cheung, M. W.-L., & Chan, W. (2005). Meta-analytic structural equation modeling: A two-stage approach. *Psychological Methods*, **10**, 40-64.
- Viswesvaran, C., & Ones, D. S. (1995). Theory testing: Combining psychometric meta-analysis and structural equations modeling. *Personnel Psychology*, **48**, 865-885.

See Also

[uniR1](#), [lavaan2RAM](#), [Becker09](#)

vanderPol17

Dataset on the effectiveness of multidimensional family therapy in treating adolescents with multiple behavior problems

Description

This dataset includes 61 effect sizes from 19 manuscripts nested from 8 studies reported by van der Pol et al. (2017). It studies the effectiveness of multidimensional family therapy in treating adolescents with multiple behavior problems.

Details

A list of data with the following structure:

Number Number of the effect size.

Study Authors of the studies.

N Total sample size.

N_target Sample size in the target group.

N_control Sample size in the control group.

Comparison_condition Either cognitive behavioral therapy (CBT), combined treatment (CT) or group therapy (Group).

Study_ID Level-3 cluster.

Age_mean Mean age of the participants.

Fllow_up Follow-up duration (in months).

Per_Males Percentage of males.

Per_Minorities Percentage of minorities.

Per_Conduct_disorder Percentage of participants with conduct disorder

Per_Severe_cannabis_users Percentage of participants of severe cannabis use.

Outcome_measure Either substance abuse, delinquency, externalizing and internalizing psychopathology, and family functioning

d Effect size in Cohen's d.

v Sampling variance of d

Source

van der Pol, T. M., Hoeve, M., Noom, M. J., Stams, G. J. J. M., Doreleijers, T. A. H., van Domburgh, L., & Vermeiren, R. R. J. M. (2017). Research Review: The effectiveness of multidimensional family therapy in treating adolescents with multiple behavior problems - a meta-analysis. *Journal of Child Psychology and Psychiatry*, **58**(5), 532-545. <https://doi.org/10.1111/jcpp.12685>

Examples

```
data(vanderPol17)
```

VarCorr

Extract Variance-Covariance Matrix of the Random Effects

Description

It extracts the variance-covariance matrix of the random effects (variance component) from either the meta or osmasem objects.

Usage

```
VarCorr(x, ...)
```

Arguments

x	An object returned from either class meta or osmasem
...	Further arguments; currently none is used

Value

A variance-covariance matrix of the random effects.

Note

It is similar to `coef(object, select="random")` in tssem. The main difference is that `coef()` returns a vector while `VarCorr()` returns its correspondent matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[coef](#), [vcov](#)

Examples

```
## Multivariate meta-analysis on the log of the odds
## The conditional sampling covariance is 0
bcg <- meta(y=cbind(ln_Odd_V, ln_Odd_NV), data=BCG,
            v=cbind(v_ln_Odd_V, cov_V_NV, v_ln_Odd_NV))
VarCorr(bcg)
```

vcov	<i>Extract Covariance Matrix Parameter Estimates from Objects of Various Classes</i>
------	--

Description

It extracts the variance-covariance matrix of the parameter estimates from objects of various classes.

Arguments

object	An object returned from objects of various classes
select	Select all for both fixed- and random-effects parameters, fixed for the fixed-effects parameters or random for the random-effects parameters. For meta3LFIML objects, allX is used to extract all parameters including the predictors and auxiliary variables.
robust	Logical. Whether to use robust standard error from imxRobustSE .
...	Further arguments; currently not in use except for tssemRobust1, which to be passed to robust .

Value

A variance-covariance matrix of the parameter estimates.

Note

vcov returns NA when the diag.constraints=TRUE argument is used in wls objects.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[tssem1](#), [wls](#), [meta](#), [reml](#)

Examples

```
## Random-effects meta-analysis
model1 <- meta(y=yi, v=vi, data=Hox02)
vcov(model1)

## Fixed-effects only
vcov(model1, select="fixed")

## Random-effects only
vcov(model1, select="random")
```

vec2symMat

*Convert a Vector into a Symmetric Matrix***Description**

It converts a vector into a symmetric matrix by filling up the elements into the lower triangle of the matrix.

Usage

```
vec2symMat(x, diag = TRUE, byrow = FALSE)
```

Arguments

x	A vector of numerics or characters
diag	Logical. If it is TRUE (the default), the diagonals of the created matrix are replaced by elements of x; otherwise, the diagonals of the created matrix are replaced by "1".
byrow	Logical. If it is FALSE (the default), the created matrix is filled by columns; otherwise, the matrix is filled by rows.

Value

A symmetric square matrix.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

See Also

[matrix2bdiag](#)

Examples

```
vec2symMat(1:6)
#      [,1] [,2] [,3]
# [1,]  1   2   3
# [2,]  2   4   5
# [3,]  3   5   6

vec2symMat(1:6, diag=FALSE)
#      [,1] [,2] [,3] [,4]
# [1,]  1   1   2   3
# [2,]  1   1   4   5
# [3,]  2   4   1   6
# [4,]  3   5   6   1

vec2symMat(letters[1:6])
#      [,1] [,2] [,3]
# [1,] "a"  "b"  "c"
# [2,] "b"  "d"  "e"
# [3,] "c"  "e"  "f"
```

wls

Conduct a Correlation/Covariance Structure Analysis with WLS

Description

It fits a correlation or covariance structure with weighted least squares (WLS) estimation method where the inverse of the asymptotic covariance matrix is used as the weight matrix. `tssem2` conducts the second stage analysis of the two-stage structural equation modeling (TSSEM). `tssem2` is a wrapper of `wls`.

Usage

```
wls(
  Cov,
  aCov,
  n,
  RAM = NULL,
  Amatrix = NULL,
  Smatrix = NULL,
  Fmatrix = NULL,
  diag.constraints = FALSE,
  cor.analysis = TRUE,
  intervals.type = c("z", "LB"),
  mx.algebras = NULL,
  mxModel.Args = NULL,
  subset.variables = NULL,
  model.name = NULL,
```

```

    suppressWarnings = TRUE,
    silent = TRUE,
    run = TRUE,
    ...
)

tssem2(
  tssem1.obj,
  RAM = NULL,
  Amatrix = NULL,
  Smatrix = NULL,
  Fmatrix = NULL,
  diag.constraints = FALSE,
  intervals.type = c("z", "LB"),
  mx.algebras = NULL,
  mxModel.Args = NULL,
  subset.variables = NULL,
  model.name = NULL,
  suppressWarnings = TRUE,
  silent = TRUE,
  run = TRUE,
  ...
)

```

Arguments

Cov	A $p \times p$ sample correlation/covariance matrix where p is the number of variables.
aCov	A $p^* \times p^*$ asymptotic sampling covariance matrix of either vechs (Cov) or vech (Cov) where $p^* = p(p - 1)/2$ for correlation matrix and $p^* = p(p + 1)/2$ for covariance matrix.
n	Sample size.
RAM	A RAM object including a list of matrices of the model returned from lavaan2RAM .
Amatrix	If RAM is not specified, an Amatrix is required. An asymmetric matrix in the RAM specification with MxMatrix-class . If it is NULL, a matrix of zero will be created. If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Smatrix	If RAM is not specified, an Smatrix is required. A symmetric matrix in the RAM specification with MxMatrix-class . If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function.
Fmatrix	A filter matrix in the RAM specification with MxMatrix-class . If it is NULL (the default), an identity matrix with the same dimensions of Cov will be created. If it is a matrix, it will be converted into MxMatrix-class by the <code>as.mxMatrix</code> function. It is not required when there is no latent variable.
diag.constraints	Logical. This argument is ignored when <code>cor.analysis=FALSE</code> . If <code>diag.constraints=TRUE</code> , the diagonals of the model implied matrix would be constrained at 1 by nonlinear constraints. The drawback is that standard error will not be generated. Parametric bootstrap is used to estimate the standard error by drawing samples from

	<p>$\mathcal{N}(\text{vech}(\text{Cov}), \text{asyCov})$ for covariance analysis and $\mathcal{N}(\text{vechs}(\text{Cov}), \text{asyCov})$ for correlation analysis while <code>asyCov</code> is treated as fixed. This process is computationally intensive. A better approach is to request likelihood-based confidence intervals (CIs) by specifying <code>intervals.type="LB"</code>. If <code>diag.constraints=FALSE</code> and <code>cor.analysis=TRUE</code>, the diagonals are automatically constrained as ones by treating the error variances as computed values rather than as parameters. Since the error variances are not parameters, they are not reported.</p>
<code>cor.analysis</code>	Logical. Analysis of correlation or covariance structure. If <code>cor.analysis=TRUE</code> , <code>vechs</code> is used to vectorize <code>S</code> ; otherwise, <code>vech</code> is used to vectorize <code>S</code> .
<code>intervals.type</code>	Either <code>z</code> (default if missing) or <code>LB</code> . If it is <code>z</code> , it calculates the 95% Wald CIs based on the <code>z</code> statistic. If it is <code>LB</code> , it calculates the 95% likelihood-based CIs on the parameter estimates. Please note that the <code>z</code> values and their associated <code>p</code> values are based on the <code>z</code> statistic. They are not related to the likelihood-based CIs.
<code>mx.algebras</code>	A list of <code>mxMatrix</code> or <code>mxAlgebra</code> objects on the <code>Amatrix</code> , <code>Smatrix</code> , and <code>Fmatrix</code> . It can be used to define new functions of parameters and their LBCIs. For example, if the regression coefficients to calculate an indirect effect are stored in <code>A[1,2]</code> and <code>A[1,3]</code> , we may define <code>list(ind=mxAlgebra(Amatrix[1,2]*Amatrix[1,3], name="ind"))</code> . See the examples in Becker92 and Hunter83 . It should be noted that <code>Fmatrix</code> , <code>Amatrix</code> , <code>Smatrix</code> , <code>Iden</code> (a $p \times p$ identity matrix), <code>sampleS</code> (sample correlation or covariance matrix), <code>impliedS1</code> , <code>impliedS</code> (model implied correlation or covariance matrix), <code>vecS</code> , <code>invAcov</code> , <code>obj</code> , <code>One</code> , <code>select</code> and <code>constraint</code> and <code>Ematrix</code> (computed error variances when <code>diag.constraints=FALSE</code>) have been defined internally. You should not create new matrices using these names.
<code>mxModel.Args</code>	A list of arguments passed to <code>mxModel</code> . These include, for example, additional <code>mxMatrix</code> and <code>mxConstraint</code> .
<code>subset.variables</code>	An optional character vector of variable names to select variables in the analysis. For example, there are 10 variables in <code>Cov</code> , say, <code>x1</code> to <code>x10</code> . We may use <code>c("x1", "x2", "x3")</code> to select three variables in the analysis. Please note that this argument does not reorder the data. That is, <code>c("x3", "x2", "x1")</code> is the same as <code>c("x1", "x2", "x3")</code> .
<code>model.name</code>	A string for the model name in <code>mxModel</code> . If it is missing, the default is "TSSEM2 (or WLS) Analysis of Correlation Structure" for <code>cor.analysis=TRUE</code> and "TSSEM2 (or WLS) Analysis of Covariance Structure" for <code>cor.analysis=FALSE</code> .
<code>suppressWarnings</code>	Logical. If <code>TRUE</code> , warnings are suppressed. The argument to be passed to <code>mxRun</code> .
<code>silent</code>	Logical. An argument to be passed to <code>mxRun</code>
<code>run</code>	Logical. If <code>FALSE</code> , only return the <code>mx</code> model without running the analysis.
<code>...</code>	Further arguments to be passed to <code>mxRun</code> .
<code>tssem1.obj</code>	An object of either class <code>tssem1FEM</code> , class <code>tssem1FEM.cluster</code> or class <code>tssem1REM</code> returned from <code>tssem1()</code>

Value

An object of class `wls` with a list of

<code>call</code>	The matched call
<code>Cov</code>	Input data of either a covariance or correlation matrix
<code>asyCov</code>	The asymptotic covariance matrix of the input data
<code>noObservedStat</code>	Number of observed statistics
<code>n</code>	Sample size
<code>cor.analysis</code>	logical
<code>noConstraints</code>	Number of constraints imposed on S
<code>indepModelChisq</code>	Chi-square statistic of the independent model returned by <code>.indepwlsChisq</code>
<code>indepModelDf</code>	Degrees of freedom of the independent model returned by <code>.indepwlsChisq</code>
<code>mx.fit</code>	A fitted object returned from <code>mxRun</code>

Note

If the input is a list of `tssem1.obj`, it returns a list of results for each cluster.

Author(s)

Mike W.-L. Cheung <mikewlcheung@nus.edu.sg>

References

- Bentler, P.M., & Savalei, V. (2010). Analysis of correlation structures: current status and open problems. In Kolenikov, S., Thombs, L., & Steinley, D. (Eds.). *Recent Methodological Developments in Social Science Statistics* (pp. 1-36). Hoboken, NJ: Wiley.
- Cheung, M. W.-L. (2010). Fixed-effects meta-analyses as multiple-group structural equation models. *Structural Equation Modeling*, **17**, 481-509.
- Cheung, M. W.-L. (2014). Fixed- and random-effects meta-analytic structural equation modeling: Examples and analyses in R. *Behavior Research Methods*, **46**, 29-40.
- Cheung, M. W.-L., & Chan, W. (2005). Meta-analytic structural equation modeling: A two-stage approach. *Psychological Methods*, **10**, 40-64.
- Cheung, M. W.-L., & Chan, W. (2009). A two-stage approach to synthesizing covariance matrices in meta-analytic structural equation modeling. *Structural Equation Modeling*, **16**, 28-53.
- Joreskog, K. G., Sorbom, D., Du Toit, S., & Du Toit, M. (1999). *LISREL 8: New Statistical Features*. Chicago: Scientific Software International.
- McArdle, J. J., & MacDonald, R. P. (1984). Some algebraic properties of the Reticular Action Model for moment structures. *British Journal of Mathematical and Statistical Psychology*, **37**, 234-251.

See Also

[tssem1](#), [Becker92](#), [Digman97](#), [Hunter83](#), [issp89](#), [issp05](#)

Examples

```
#### Analysis of correlation structure
R1.labels <- c("a1", "a2", "a3", "a4")

R1 <- matrix(c(1.00, 0.22, 0.24, 0.18,
               0.22, 1.00, 0.30, 0.22,
               0.24, 0.30, 1.00, 0.24,
               0.18, 0.22, 0.24, 1.00), ncol=4, nrow=4,
             dimnames=list(R1.labels, R1.labels))

n <- 1000
acovR1 <- asyCov(R1, n)

#### One-factor CFA model using lavaan specification
model1 <- "f =~ a1 + a2 + a3 + a4"

RAM1 <- lavaan2RAM(model1, obs.variables=R1.labels)

wls.fit1a <- wls(Cov=R1, aCov=acovR1, n=n, RAM=RAM1,
                 cor.analysis=TRUE, intervals="LB")
summary(wls.fit1a)

## One-factor CFA model using RAM specification
(A1 <- cbind(matrix(0, nrow=5, ncol=4),
             matrix(c("0.2*a1", "0.2*a2", "0.2*a3", "0.2*a4", 0),
                   ncol=1)))

(S1 <- Diag(c("0.2*e1", "0.2*e2", "0.2*e3", "0.2*e4", 1)))

## The first 4 variables are observed while the last one is latent.
(F1 <- create.Fmatrix(c(1,1,1,1,0), name="F1"))

wls.fit1b <- wls(Cov=R1, aCov=acovR1, n=n, Fmatrix=F1, Smatrix=S1, Amatrix=A1,
                 cor.analysis=TRUE, intervals="LB")
summary(wls.fit1b)

## Select 3 variables to analyze
model2 <- "f =~ a1 + a2 + a3"

RAM2 <- lavaan2RAM(model2, obs.variables=R1.labels[-4])

wls.fit1c <- wls(Cov=R1, aCov=acovR1, n=n, RAM=RAM2,
                 cor.analysis=TRUE, subset.variables=c("a1", "a2", "a3"))
summary(wls.fit1c)

#### Multiple regression analysis using lavaan specification
R2.labels <- c("y", "x1", "x2")

R2 <- matrix(c(1.00, 0.22, 0.24,
               0.22, 1.00, 0.30,
               0.24, 0.30, 1.00), ncol=3, nrow=3,
             dimnames=list(R2.labels, R2.labels))
```

```

acovR2 <- asyCov(R2, n)

model3 <- "y ~ x1 + x2
          ## Variances of x1 and x2 are 1
          x1 ~~ 1*x1
          x2 ~~ 1*x2
          ## x1 and x2 are correlated
          x1 ~~ x2"

RAM3 <- lavaan2RAM(model3, obs.variables=R2.labels)

wls.fit2a <- wls(Cov=R2, aCov=acovR2, n=n, RAM=RAM3,
                 cor.analysis=TRUE, intervals="z")
summary(wls.fit2a)

#### Multiple regression analysis using RAM specification

## A2: Regression coefficients
#   y x1 x2
# y   F T  T
# x1  F F  F
# x2  F F  F
(A2 <- mxMatrix("Full", ncol=3, nrow=3, byrow=TRUE,
                 free=c(FALSE, rep(TRUE, 2), rep(FALSE, 6)), name="A2"))

## S2: Covariance matrix of free parameters
#   y x1 x2
# y   T F  F
# x1  F F  F
# x2  F T  F
(S2 <- mxMatrix("Symm", ncol=3, nrow=3, values=c(0.2,0,0,1,0.2,1),
                 labels=c("Var_y", NA, NA, NA, "Cov_x1_x2", NA),
                 free=c(TRUE,FALSE,FALSE,FALSE,TRUE,FALSE), name="S2"))

## F may be ignored as there is no latent variable.
wls.fit2b <- wls(Cov=R2, aCov=acovR2, n=n, Amatrix=A2, Smatrix=S2,
                 cor.analysis=TRUE, intervals="LB")
summary(wls.fit2b)

#### Analysis of covariance structure using lavaan specification
R3.labels=c("a1", "a2", "a3", "a4")

R3 <- matrix(c(1.50, 0.22, 0.24, 0.18,
               0.22, 1.60, 0.30, 0.22,
               0.24, 0.30, 1.80, 0.24,
               0.18, 0.22, 0.24, 1.30), ncol=4, nrow=4,
             dimnames=list(R3.labels, R3.labels))

n <- 1000
acovS3 <- asyCov(R3, n, cor.analysis=FALSE)

model3 <- "f =~ a1 + a2 + a3 + a4"

```

```

RAM3 <- lavaan2RAM(model3, obs.variables=R3.labels)

wls.fit3a <- wls(Cov=R3, aCov=acovS3, n=n, RAM=RAM3,
                 cor.analysis=FALSE)
summary(wls.fit3a)

#### Analysis of covariance structure using RAM specification
(A3 <- cbind(matrix(0, nrow=5, ncol=4),
             matrix(c("0.2*a1", "0.2*a2", "0.2*a3", "0.2*a4", 0), ncol=1)))

(S3 <- Diag(c("0.2*e1", "0.2*e2", "0.2*e3", "0.2*e4", 1)))

F3 <- c(TRUE, TRUE, TRUE, TRUE, FALSE)
(F3 <- create.Fmatrix(F3, name="F3", as.mxMatrix=FALSE))

wls.fit3b <- wls(Cov=R3, aCov=acovS3, n=n, Amatrix=A3, Smatrix=S3,
                 Fmatrix=F3, cor.analysis=FALSE)
summary(wls.fit3b)

```

wvs94a

Forty-four Studies from Cheung (2013)

Description

Between 1990 and 1993, 57,561 adults aged 18 and above from 42 nations were interviewed by local academic institutes in Eastern European nations and by professional survey organizations in other nations. The standardized mean difference (SMD) between males and females on life satisfaction and life control in each country were calculated as the effect sizes. Positive values indicate that males have higher scores than females do.

Details

The variables are:

country Country

lifesat SMD on life satisfaction

lifecon SMD on life control

lifesat_var Sampling variance of lifesat

inter_cov Sampling covariance between lifesat and lifecon

lifecon_var Sampling variance of lifecon

gnp Gross National Product

Source

World Values Study Group. (1994). World Values Survey, 1981-1984 and 1990-1993 [Computer file]. Ann Arbor, MI: Inter-university Consortium for Political and Social Research.

References

- Au, K., & Cheung, M. W.-L. (2004). Intra-cultural variation and job autonomy in 42 countries. *Organization Studies*, **25**, 1339-1362.
- Cheung, M. W.-L. (2013). Multivariate meta-analysis as structural equation models. *Structural Equation Modeling*, **20**, 429-454.

Examples

```
data(wvs94a)

## Random-effects model
random.ma1 <- meta(y=cbind(lifesat, lifecon),
                  v=cbind(lifesat_var, inter_cov, lifecon_var), data=wvs94a,
                  model.name="Random effects model")
summary(random.ma1)

## Random-effects model with both population effect sizes fixed at 0
random.ma2 <- meta(y=cbind(lifesat, lifecon),
                  v=cbind(lifesat_var, inter_cov, lifecon_var), data=wvs94a,
                  intercept.constraints=matrix(0, nrow=1, ncol=2),
                  model.name="Effect sizes are fixed at 0")
summary(random.ma2)

## Compare the nested models
anova(random.ma1, random.ma2)

## Fixed-effects model by fixing the variance component at 0
fixed.ma <- meta(y=cbind(lifesat, lifecon),
                v=cbind(lifesat_var, inter_cov, lifecon_var), data=wvs94a,
                RE.constraints=matrix(0, ncol=2, nrow=2),
                model.name="Fixed effects model")
summary(fixed.ma)

## Mixed-effects model
## gnp is divided by 10000 and centered by using
## scale(gnp/10000, scale=FALSE)
mixed.ma1 <- meta(y=cbind(lifesat, lifecon),
                v=cbind(lifesat_var, inter_cov, lifecon_var),
                x=scale(gnp/10000, scale=FALSE), data=wvs94a,
                model.name="GNP as a predictor")
summary(mixed.ma1)

## Mixed-effects model with equal regression coefficients
mixed.ma2 <- meta(y=cbind(lifesat, lifecon),
                v=cbind(lifesat_var, inter_cov, lifecon_var),
                x=scale(gnp/10000, scale=FALSE), data=wvs94a,
                coef.constraints=matrix(c("0.0*Eq_slope",
                                         "0.0*Eq_slope"), nrow=2),
                model.name="GNP as a predictor with equal slope")
summary(mixed.ma2)
```

```
## Compare the nested models
anova(mixed.ma1, mixed.ma2)

## Plot the multivariate effect sizes
plot(random.ma1, main="Estimated effect sizes and their 95% confidence ellipses",
      axis.label=c("Gender difference on life satisfaction",
                  "Gender difference on life control"))
```

wvs94b

*Forty-four Covariance Matrices on Life Satisfaction, Job Satisfaction,
and Job Autonomy*

Description

Between 1990 and 1993, 57,561 adults aged 18 and above from 42 nations were interviewed by local academic institutes in Eastern European nations and by professional survey organizations in other nations. The covariance matrices among Life Satisfaction, Job Satisfaction, and Job Autonomy were calculated.

Details

The variables are:

data Covariance matrix among Life Satisfaction (LS), Job Satisfaction (JS), and Job Autonomy (JA)

n Sample size in the country

Source

World Values Study Group. (1994). World Values Survey, 1981-1984 and 1990-1993 [Computer file]. Ann Arbor, MI: Inter-university Consortium for Political and Social Research.

References

Au, K., & Cheung, M. W.-L. (2004). Intra-cultural variation and job autonomy in 42 countries. *Organization Studies*, **25**, 1339-1362.

Cheung, M.W.-L., & Cheung, S.-F. (2016). Random-effects models for meta-analytic structural equation modeling: Review, issues, and illustrations. *Research Synthesis Methods*, **7**, 140-155.

Examples

```
data(wvs94b)

## Get the indirect and the direct effects and
## their sampling covariance matrices for each study
indirect1 <- indirectEffect(wvs94b$data, wvs94b$n)
indirect1

## Multivariate meta-analysis on the indirect and direct effects
indirect2 <- meta(indirect1[, c("ind_eff", "dir_eff")],
                  indirect1[, c("ind_var", "ind_dir_cov", "dir_var")])

summary(indirect2)
```

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